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# Variations in dissolved greenhouse gases ( $\text{CO}_2$ , $\text{CH}_4$ , $\text{N}_2\text{O}$ ) in the Congo River network overwhelmingly driven by fluvial-wetland connectivity

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**Abstract.** We carried out 10 field expeditions between 2010 and 2015 in the lowland part of the Congo River network in the eastern part of the basin (Democratic Republic of the Congo), to describe the spatial variations in fluvial dissolved carbon dioxide ( $\text{CO}_2$ ), methane ( $\text{CH}_4$ ) and nitrous oxide ( $\text{N}_2\text{O}$ ) concentrations. We investigate the possible drivers of the spatial variations in dissolved  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$  concentrations by analyzing covariations with several other biogeochemical variables, aquatic metabolic processes (primary production and respiration), catchment characteristics (land cover) and wetland spatial distributions. We test the hypothesis that spatial patterns of  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$  are partly due to the connectivity with wetlands, in particular with a giant wetland of flooded forest in the core of the Congo basin, the “Cuvette Centrale Congolaise” (CCC). Two transects of 1650 km were carried out from the city of Kisangani to the city of Kinshasa, along the longest possible navigable section of the river and corresponding to 41 % of the total length of the main stem. Additionally, three time series of  $\text{CH}_4$  and  $\text{N}_2\text{O}$  were obtained at fixed points in the main stem of the middle Congo (2013–2018, biweekly sampling), in the main stem of the lower Kasai (2015–2017,

monthly sampling) and in the main stem of the middle Oubangui (2010–2012, biweekly sampling). The variations in dissolved  $\text{N}_2\text{O}$  concentrations were modest, with values oscillating around the concentration corresponding to saturation with the atmosphere, with  $\text{N}_2\text{O}$  saturation level (% $\text{N}_2\text{O}$ , where atmospheric equilibrium corresponds to 100 %) ranging between 0 % and 561 % (average 142 %). The relatively narrow range of % $\text{N}_2\text{O}$  variations was consistent with low  $\text{NH}_4^+$  ( $2.3 \pm 1.3 \mu\text{mol L}^{-1}$ ) and  $\text{NO}_3^-$  ( $5.6 \pm 5.1 \mu\text{mol L}^{-1}$ ) levels in these near pristine rivers and streams, with low agriculture pressure on the catchment (croplands correspond to 0.1 % of catchment land cover of sampled rivers), dominated by forests (~70 % of land cover). The covariations in % $\text{N}_2\text{O}$ ,  $\text{NH}_4^+$ ,  $\text{NO}_3^-$  and dissolved oxygen saturation level (% $\text{O}_2$ ) indicate  $\text{N}_2\text{O}$  removal by soil or sedimentary denitrification in low  $\text{O}_2$ , high  $\text{NH}_4^+$  and low  $\text{NO}_3^-$  environments (typically small and organic matter rich streams) and  $\text{N}_2\text{O}$  production by nitrification in high  $\text{O}_2$ , low  $\text{NH}_4^+$  and high  $\text{NO}_3^-$  (typical of larger rivers that are poor in organic matter). Surface waters were very strongly oversaturated in  $\text{CO}_2$  and  $\text{CH}_4$  with respect to atmospheric equilibrium, with values of the partial pressure of  $\text{CO}_2$  ( $p\text{CO}_2$ ) rang-

ing between 1087 and 22 899 ppm (equilibrium  $\sim 400$  ppm) and dissolved CH<sub>4</sub> concentrations ranging between 22 and 71 428 nmol L<sup>-1</sup> (equilibrium  $\sim 2$  nmol L<sup>-1</sup>). Spatial variations were overwhelmingly more important than seasonal variations for *p*CO<sub>2</sub>, CH<sub>4</sub> and %N<sub>2</sub>O as well as day–night variations for *p*CO<sub>2</sub>. The wide range of *p*CO<sub>2</sub> and CH<sub>4</sub> variations was consistent with the equally wide range of %O<sub>2</sub> (0.3%–122.8%) and of dissolved organic carbon (DOC) (1.8–67.8 mg L<sup>-1</sup>), indicative of generation of these two greenhouse gases from intense processing of organic matter either in “terra firme” soils, wetlands or in-stream. However, the emission rate of CO<sub>2</sub> to the atmosphere from riverine surface waters was on average about 10 times higher than the flux of CO<sub>2</sub> produced by aquatic net heterotrophy (as evaluated from measurements of pelagic respiration and primary production). This indicates that the CO<sub>2</sub> emissions from the river network were sustained by lateral inputs of CO<sub>2</sub> (either from terra firme or from wetlands). The *p*CO<sub>2</sub> and CH<sub>4</sub> values decreased and %O<sub>2</sub> increased with increasing Strahler order, showing that stream size explains part of the spatial variability of these quantities. In addition, several lines of evidence indicate that lateral inputs of carbon from wetlands (flooded forest and aquatic macrophytes) were of paramount importance in sustaining high CO<sub>2</sub> and CH<sub>4</sub> concentrations in the Congo river network, as well as driving spatial variations: the rivers draining the CCC were characterized by significantly higher *p*CO<sub>2</sub> and CH<sub>4</sub> and significantly lower %O<sub>2</sub> and %N<sub>2</sub>O values than those not draining the CCC; *p*CO<sub>2</sub> and %O<sub>2</sub> values were correlated to the coverage of flooded forest on the catchment. The flux of greenhouse gases (GHGs) between rivers and the atmosphere averaged 2469 mmol m<sup>-2</sup> d<sup>-1</sup> for CO<sub>2</sub> (range 86 and 7110 mmol m<sup>-2</sup> d<sup>-1</sup>), 12 553 μmol m<sup>-2</sup> d<sup>-1</sup> for CH<sub>4</sub> (range 65 and 597 260 μmol m<sup>-2</sup> d<sup>-1</sup>) and 22 μmol m<sup>-2</sup> d<sup>-1</sup> for N<sub>2</sub>O (range –52 and 319 μmol m<sup>-2</sup> d<sup>-1</sup>). The estimate of integrated CO<sub>2</sub> emission from the Congo River network ( $251 \pm 46$  TgC ( $10^{12}$  gC) yr<sup>-1</sup>), corresponding to nearly half the CO<sub>2</sub> emissions from tropical oceans globally ( $565$  TgC yr<sup>-1</sup>) and was nearly 2 times the CO<sub>2</sub> emissions from the tropical Atlantic Ocean ( $137$  TgC yr<sup>-1</sup>). Moreover, the integrated CO<sub>2</sub> emission from the Congo River network is more than 3 times higher than the estimate of terrestrial net ecosystem exchange (NEE) on the whole catchment ( $77$  TgC yr<sup>-1</sup>). This shows that it is unlikely that the CO<sub>2</sub> emissions from the river network were sustained by the hydrological carbon export from terra firme soils (typically very small compared to terrestrial NEE) but most likely, to a large extent, they were sustained by wetlands (with a much higher hydrological connectivity with rivers and streams).

## 1 Introduction

Emissions to the atmosphere of greenhouse gases (GHGs) such as carbon dioxide (CO<sub>2</sub>), methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O) from inland waters (rivers, lakes and reservoirs) might be quantitatively important for global budgets (Seitzinger and Kroeze, 1998; Cole et al., 2007; Bastviken et al., 2011). Yet, there are very large uncertainties in the estimates of GHGs emission to the atmosphere from rivers, as reflected in the wide range of reported values, between 0.2 and 1.8 PgC ( $10^{15}$  gC) yr<sup>-1</sup> for CO<sub>2</sub> (Cole et al., 2007; Raymond et al., 2013), 2 and 27 TgCH<sub>4</sub> ( $10^{12}$  gCH<sub>4</sub>) yr<sup>-1</sup> for CH<sub>4</sub> (Bastviken et al., 2011; Stanley et al., 2016), and 32 and 2100 GgN<sub>2</sub>O-N ( $10^9$  gN<sub>2</sub>O-N) yr<sup>-1</sup> for N<sub>2</sub>O (Kroeze et al., 2010; Hu et al., 2016). This uncertainty is mainly due to the scarcity of data in the tropics that account for the majority of riverine GHG emissions ( $\sim 80\%$  for CO<sub>2</sub>, Raymond et al., 2013; Borges et al., 2015a, b; Lauerwald et al., 2015, 79% for N<sub>2</sub>O, Hu et al., 2016; 70% for CH<sub>4</sub>, Sawakushi et al., 2014) but also to scaling procedures of varying complexity that use different input data (Raymond et al., 2013; Borges et al., 2015b; Lauerwald et al., 2015), in addition to uncertainty in the estimate of surface area of rivers (Downing et al., 2012; Raymond et al., 2013; Allen and Pavelsky, 2018) and the parameterization of the gas transfer velocity (*k*) (Raymond et al., 2012; Huotari et al., 20013; Maurice et al., 2017; Kokic et al., 2018; McDowell and Johnson, 2018; Ulseth et al., 2019). The exchange of CO<sub>2</sub> between rivers and the atmosphere is in most cases computed from the air–water gradient of the CO<sub>2</sub> concentration and *k* parameterized as a function of stream morphology (e.g., slope or depth) and water flow (or discharge). However, there can be large errors associated with the computation of the dissolved CO<sub>2</sub> concentration from pH and total alkalinity (TA) for low alkalinity waters in particular so that high-quality direct measurements of dissolved CO<sub>2</sub> concentration are preferred (Abril et al., 2015), although scarce. In the tropics, research on GHGs in rivers has mainly focussed on South American rivers and on the central Amazon in particular (Richey et al., 1988, 2002; Melack et al., 2004; Abril et al., 2014; Barbosa et al., 2016; Scofield et al., 2016), while until recently African rivers were nearly uncharted with a few exceptions (Koné et al., 2009, 2010).

There is also a lack of understanding of the drivers of the fluvial concentrations of GHGs, hence, ultimately of the drivers of their exchange with the atmosphere. It is unclear to what extent CO<sub>2</sub> emissions from rivers are sustained by in situ net heterotrophy and/or by lateral inputs of CO<sub>2</sub>. The global organic carbon degradation by net heterotrophy of rivers and streams given by Battin et al. (2008) of  $0.2$  PgC yr<sup>-1</sup> is insufficient to sustain global riverine CO<sub>2</sub> emissions given by the most recent estimates of  $0.7$ – $1.8$  PgC yr<sup>-1</sup> (Raymond et al., 2013; Lauerwald et al., 2015), suggesting an important role of lateral CO<sub>2</sub> inputs in sustaining emissions to the atmosphere from rivers. In a regional

study in the US, Hotchkiss et al. (2015) estimated that instream organic matter degradation could only sustain 14 % and 39 % of CO<sub>2</sub> riverine emissions in small and large systems, respectively. It is also unclear to what extent CO<sub>2</sub> and CH<sub>4</sub> emissions from rivers are sustained by carbon inputs either from the terrestrial biome (*terra firme*) or from wetlands (flooded forests and macrophytes) (Abril and Borges, 2019). This difference has large implications for our fundamental understanding of carbon cycling in rivers and their connectivity with respective catchments but also, consequently, for our capacity to predict how GHG emissions from rivers might be modified in response to climate change (warming and modification of the hydrological cycle), water diversion (damming, water abstraction) or land use change on the catchment (e.g., Klaus et al., 2018). In upland areas, low-order stream CO<sub>2</sub> emissions are undoubtedly related to soil-water and groundwater CO<sub>2</sub> inputs, although CO<sub>2</sub> degassing takes place over very short distances from point sources ( $\leq 200$  m), is highly variable in space and seasonally, and mainly occurs during short-lived high-flow events that promote shallow flow paths (e.g., Duvert et al., 2018). Low-order streams (1–3) might account for about one-third of the global riverine CO<sub>2</sub> emissions (Marx et al., 2017). Recently acquired datasets allowed us to show that inputs from riparian wetlands seem to be of paramount importance in sustaining CO<sub>2</sub> and CH<sub>4</sub> emissions to the atmosphere from large tropical lowland rivers (Abril et al., 2014; Borges et al., 2015a, b). About half of the global surface area of wetlands is located in the tropics and subtropics (33° N–33° S), the rest are in the Northern Hemisphere (Fluet-Chouinard et al., 2015), and more than half of river surface area is located in the tropics and subtropics (Allen and Pavelsky, 2018).

The Congo River (~ 4400 km length, freshwater discharge ~ 44 000 m<sup>3</sup> s<sup>-1</sup>) has a large drainage basin ( $3.7 \times 10^6$  km<sup>2</sup>) covered by evergreen forest (dense and mosaic) (~ 67 %) and savannah (shrubland and grassland) (~ 30 %), owing to the tropical climate (annual average precipitation of 1530 mm and air temperature of 23.7 °C). The Congo basin accounts for 89 % of African rainforests. These rainforests are spread between the Democratic Republic of the Congo (54 %), Gabon (11 %), Cameroon (10 %) and the Republic of the Congo (10 %), the remaining 15 % being shared by several other countries. The mean above-ground biomass of the rainforests in Central Africa (43 kg dry biomass (db) m<sup>-2</sup>) is much higher than the mean in Amazonia (29 kg<sub>db</sub> m<sup>-2</sup>) and nearly equals the mean in the notorious high biomass forests of Borneo (44 kg<sub>db</sub> m<sup>-2</sup>) (Malhi et al., 2013). Current estimates of carbon transport from the Congo River close to the mouth rank it as the Earth's second-largest supplier of organic carbon to the oceans (Coynel et al., 2005). Despite its overwhelming importance, our knowledge of carbon and nutrient cycling in the Congo river basin is limited to some transport flux data from the 1980s, reviewed by Laraque et al. (2009) and a number of more recent small-scale studies (Bouillon et al., 2012, 2014; Spencer et al., 2012; Lambert

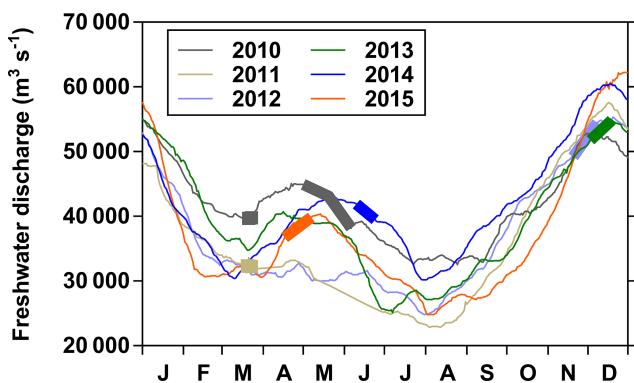
et al., 2016), in sharp contrast to the extensive and sustained work that has been done on the Amazon river basin (Alsdorf et al., 2016). The Congo basin has a wide range of contrasting tributaries (differing in lithology, soil characteristics, vegetation, rainfall patterns, etc.) and extensive flooded forests in its central region, the “Cuvette Centrale Congolaise” (CCC), with an estimated flooded cover of 360 000 km<sup>2</sup>, for a total surface area of 1 760 000 km<sup>2</sup> (Bwangoy et al., 2010). Extensive peat deposits are present beneath the swamp forest of the CCC that store below-ground 31 PgC of organic carbon, a quantity similar to the above-ground carbon stock of the forests of the entire Congo basin (Dargie et al., 2017). The tributaries partly drain semi-humid catchments with alternating dry and wet seasons on both sides of the Equator, resulting in a relatively constant discharge and water level for the main stem Congo River (Runge, 2008). Hence, the CCC is an extended zone of year-round inundation (Bwangoy et al., 2010), in sharp contrast with other large tropical rivers such as the Amazon where floodplain inundation shows clear seasonality (Hamilton et al., 2002).

Data of dissolved GHGs (CH<sub>4</sub>, N<sub>2</sub>O and CO<sub>2</sub>) have been reported in rivers in the western part of the Congo basin in four major river basins in the Republic of the Congo (Alima, Lefini, Sangha, Likouala-Mossaka) (Mann et al., 2014; Upstill-Goddard et al., 2017), and we previously reported GHGs data collected in the eastern part of the basin in the Democratic Republic of the Congo in the framework of a broad synthesis of riverine GHGs at the scale of the African continent (Borges et al., 2015a) and a general comparison of the Congo and the Amazon rivers (Borges et al., 2015b). Here, we describe in more detail the variability of GHGs based on a dataset collected during 10 field expeditions from 2010 to 2015 (Figs. 1 and 2) (Borges and Bouillon, 2019), in particular with regards to spatial and seasonal patterns, as well as with regards to the origin of fluvial CO<sub>2</sub> by integrating metabolic measurements (primary production and respiration), stable isotope ratios of dissolved inorganic carbon (DIC) and characteristics of the catchments with regards to the cover of wetlands. Comparison of data obtained in streams within and outside the giant wetland area of the CCC allows a natural large-scale test of the influence of the connectivity of wetlands on CO<sub>2</sub> and CH<sub>4</sub> dynamics in lowland tropical rivers.

## 2 Material and methods

### 2.1 Field expeditions and fixed monitoring

Samples were collected during a total of 10 field expeditions (Figs. 1 and 2). Three were done from a medium sized boat (22 m long) on which we deployed the equipment for continuous measurements of the partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$ ) (total  $n = 30\,490$ ) as well as the field laboratory for conditioning water samples. Sampling in the



**Figure 1.** Freshwater discharge ( $\text{m}^3 \text{s}^{-1}$ ) of the Congo River at Kinshasa from 2010 to 2015, with an indication of field expedition duration (thick lines).

main stem and large tributaries was made from the medium sized boat, while sampling in smaller tributaries was made with a pirogue. These large-scale field expeditions covered the Kisangani–Kinshasa transect twice (3–19 December 2013 and 10–30 June 2014) and the Kwa river up to Ilebo (16 April–6 May 2015). During the other cruises, the field laboratory was installed in a base camp (typically in a village along the river), and traveling and sampling was made with small pirogues on which it was not possible to deploy the apparatus for continuous measurements of  $p\text{CO}_2$ . Three cruises covered the section from Kisangani to the mouth of the Lomami River (20 November–8 December 2012; 17–26 September 2013; 13–21 March 2014), one cruise covered the section from Kisangani to the mouth of the Itimbiri River (8 May–6 June 2010) and three cruises (previously reported by Bouillon et al., 2012, 2014) covered the Oubangui river network around the city of Bangui (21–23 March 2010, 20–23 March 2011, and 20–24 November 2012).

Fixed sampling was carried out in the Congo main stem in proximity of the city of Kisangani (10 December 2012–16 April 2018), the Oubangui main stem in proximity of the city of Bangui (20 March 2010–31 March 2012) and the Kasai main stem in proximity of the village of Dima, close to the city of Bandundu (14 April 2015–15 May 2017). Sampling was carried out at regular intervals, every 15 d in the Congo and Oubangui main stems and every month in the Kasai main stem. Data from the Oubangui catchment were previously reported by Bouillon et al. (2012, 2014).

## 2.2 Continuous measurements

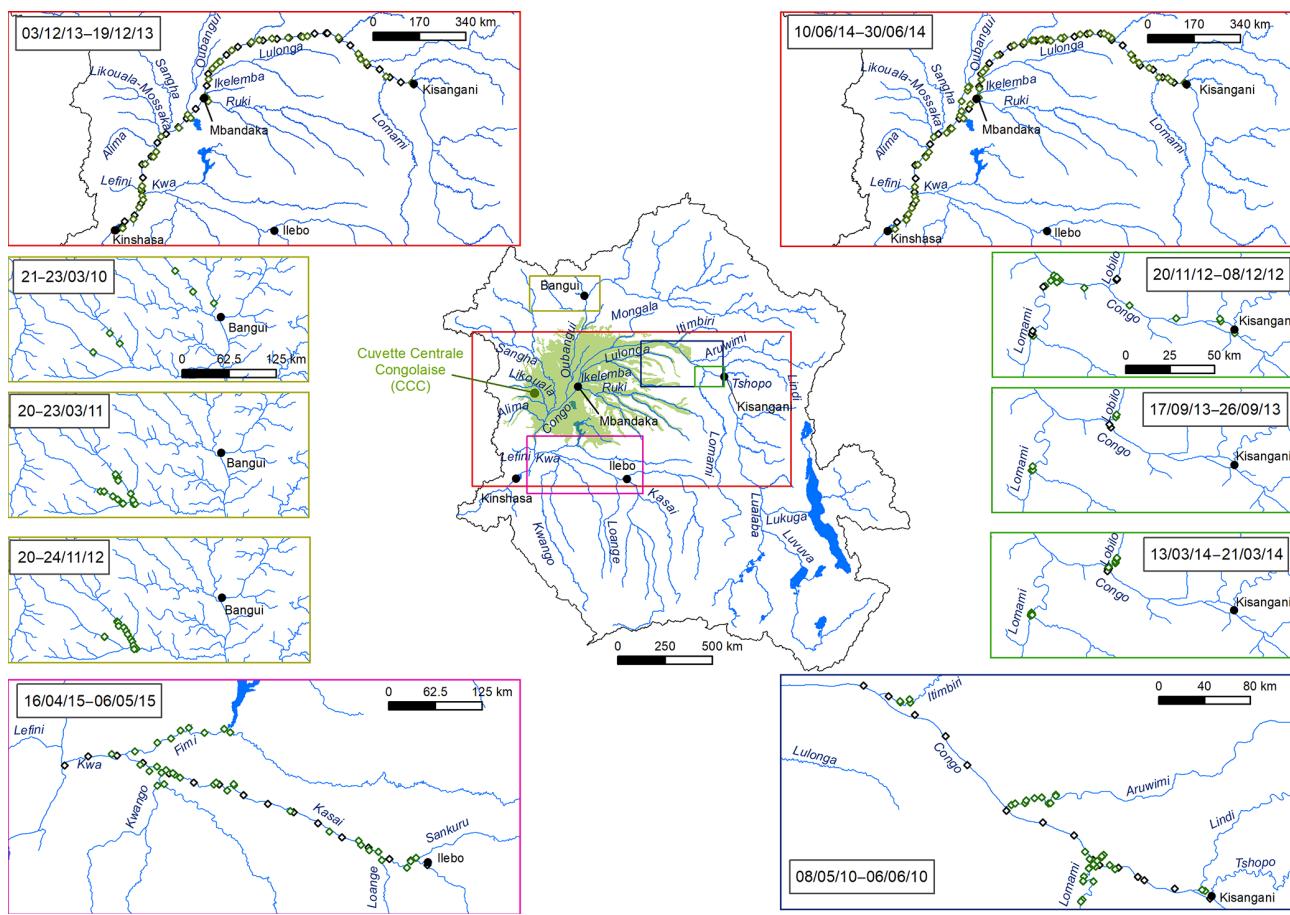
Continuous measurements (1 min interval) of  $p\text{CO}_2$  were made with an equilibrator designed for turbid waters (Frankignoulle et al., 2001) coupled to a nondispersive infrared gas analyzer (IRGA) (Li-Cor 840). The equilibrator consisted of a Plexiglas cylinder (height of 70 cm and internal diameter of 7 cm) filled with glass marbles; pumped water flowed from the top to the bottom of the equilibrator

at a rate of about  $3 \text{ L min}^{-1}$ ; water residence time within the equilibrator was about 10 s, while 99 % of equilibration was achieved in less than 120 s (Frankignoulle et al., 2001). This type of equilibrator system was shown to be the fastest among commonly used equilibration designs (Santos et al., 2012). In parallel to the  $p\text{CO}_2$  measurements, water temperature, specific conductivity, pH, dissolved oxygen saturation level (%O<sub>2</sub>), and turbidity were measured with an YSI multiparameter probe (6600) and position was measured with a Garmin geographical position system (Map 60S) portable probe. Water was pumped to the equilibrator and the multiparameter probe (on deck) with a 12V powered water pump (LVM105) attached on a wooden pole to the side of the boat at about 1 m depth. We did not observe bubble entrainment in water circuit to the equilibrator, as the sailing speed was low (maximum speed  $12 \text{ km h}^{-1}$ ). Instrumentation was powered by 12 V batteries that were recharged in the evening with power generators.

## 2.3 Discrete sampling

In smaller streams, sampling was done from the side of a pirogue with a 1.7 L Niskin bottle (General Oceanics) for gases (CO<sub>2</sub>, CH<sub>4</sub>, N<sub>2</sub>O) and a 5 L polyethylene water container for other variables. Water temperature, specific conductivity, pH and %O<sub>2</sub> were measured in situ with a YSI multiparameter probe (ProPlus).  $p\text{CO}_2$  was measured with a Li-Cor Li-840 IRGA based on the headspace technique with four polypropylene syringes (Abril et al., 2015). During two cruises (20 November–8 December 2012; 17–26 September 2013,  $n = 38$ ), a PP Systems EGM-4 was used as an IRGA instead of the Li-Cor Li-840. During one of the cruises (13–21 March 2014,  $n = 20$ ) the equilibrated headspace was stored in pre-evacuated 12 mL Exetainer (Labco) vials and analyzed in our home laboratory with a gas chromatograph (GC) (see below). Similarly, the equilibrated headspace was stored in pre-evacuated 12 mL Exetainer (Labco) vials for  $p\text{CO}_2$  analysis from the fixed sampling in Kisangani. This approach was preferred to the analysis of  $p\text{CO}_2$  from the samples for CH<sub>4</sub> and N<sub>2</sub>O analysis, as the addition of HgCl<sub>2</sub> to preserve the water sample from biological alteration led to an artificial increase in CO<sub>2</sub> concentrations most probably related to the precipitation of HgCO<sub>3</sub> (Supplement Fig. S1).

Both YSI multiparameter probes were calibrated according to manufacturer's specifications, in air for %O<sub>2</sub> and with standard solutions for other variables: commercial pH buffers (4.00 and 7.00), a  $1000 \mu\text{S cm}^{-1}$  standard for conductivity and a 124 nephelometric turbidity unit (NTU) standard for turbidity. The turbidity data from the YSI 6600 compared satisfactorily with discrete total suspended matter (TSM) measurements (Fig. S2), so hereafter we will refer to TSM for both discrete samples and sensor data. The Li-Cor 840 IRGAs were calibrated before and after each cruise with ultra-pure N<sub>2</sub> and a suite of gas standards (Air Liquide Belgium),



**Figure 2.** Map showing the sampling stations of the 10 field expeditions in the Congo River network.

with CO<sub>2</sub> mixing ratios of 388, 813, 3788 and 8300 ppm. The overall precision of *p*CO<sub>2</sub> measurements was ±2.0 %.

#### 2.4 Metabolism measurements

Primary production (PP) was measured during 2 h incubations along a gradient of light intensity using <sup>13</sup>C-HCO<sub>3</sub><sup>-</sup> as a tracer, as described in detail by Descy et al. (2017). Data were integrated vertically with PAR profiles made with a Li-Cor Li-193 underwater spherical sensor and at daily scale with surface PAR data measured during the cruises with a Li-190R quantum sensor. In order to extend the number of PP data points we developed a very simple model as a function of chlorophyll *a* concentration (Chl *a*) and of Secchi depth (*S*<sub>d</sub>):

$$\text{PP} = -4.4 + 0.166 \times S_d + 3.751 \times \text{Chl } a, \quad (1)$$

where PP is in mmol m<sup>-2</sup> d<sup>-1</sup>, *S*<sub>d</sub> in cm and Chl *a* in µg L<sup>-1</sup>

This approach is inspired from empirical models such as the one developed by Cole and Cloern (1987) that accounts for phytoplankton biomass given by Chl *a*, light extinction given by the photic depth and daily surface irradiance. We use *S*<sub>d</sub> as a proxy for photic depth and, in order to simplify

the computations, we did not include in the model daily surface irradiance, since it is nearly constant year round in our study region close to the Equator. The model satisfactorily predicts the PP (Fig. S3), except at very low Chl *a* values at which the model overestimates PP (due to the *S*<sub>d</sub> term). In order to overcome this, we assumed a zero PP value for Chl *a* concentrations < 0.3 µg L<sup>-1</sup>.

Pelagic community respiration (CR) was determined from the decrease in O<sub>2</sub> in 60 mL biological oxygen demand bottles (Wheaton) over ~24 h incubation periods. The bottles were kept in the dark and close to in situ temperature in a cooler filled with in situ water. The O<sub>2</sub> decrease was determined from triplicate measurements at the start and the end of the incubation with an optical O<sub>2</sub> probe (YSI ProODO). At the end of incubation the sealed samples were homogenized with a magnetic bar prior to the measurement of O<sub>2</sub>. Depth integration was made by multiplying the CR in surface waters by the depth measured at the station with a portable depth meter (Plastimo Echotest-II).

For methane oxidation measurements, seven 60 mL borosilicate serum bottles (Wheaton) were filled sequentially from the Niskin bottle and immediately sealed with butyl

stoppers and crimped with aluminum caps. The butyl stoppers were cleaned of leachable chemicals by boiling in deionized water during 15 min in our home laboratory. The first and the last bottle to be filled were then poisoned with a saturated solution of  $\text{HgCl}_2$  (100  $\mu\text{L}$ ) injected through the butyl stopper with a polypropylene syringe and a steel needle, corresponding to the initial concentration of the incubation (T0). The other bottles were stored in a cooler full of in situ water (to keep samples close to in situ temperature and in the dark) and were poisoned approximately 12 h after T0 (T1) and then approximately every 24 h after T0 (T2, T3, T4 and T5). The difference between the two T0 samples was close to the typical precision of measurements showing that the water was homogeneous within the Niskin bottle with regards to  $\text{CH}_4$  concentration, and no measurable loss of  $\text{CH}_4$  occurred when filling the seven vials. Methane oxidation was computed from the linear decrease in  $\text{CH}_4$  concentration with time.

## 2.5 Sample conditioning and laboratory analysis

Samples for  $\text{CH}_4$  and  $\text{N}_2\text{O}$  were collected from the Niskin bottle with a silicone tube in 60 mL borosilicate serum bottles (Wheaton), poisoned with 200  $\mu\text{L}$  of a saturated solution of  $\text{HgCl}_2$  and sealed with a butyl stopper and crimped with aluminum cap. Measurements were made with the headspace technique (Weiss, 1981) and a GC (SRI 8610C) with a flame ionization detector for  $\text{CH}_4$  (with a methanizer for  $\text{CO}_2$ ) and electron capture detector for  $\text{N}_2\text{O}$  calibrated with  $\text{CO}_2 : \text{CH}_4 : \text{N}_2\text{O} : \text{N}_2$  gas mixtures (Air Liquide Belgium) with mixing ratios of 1, 10, and 30 ppm for  $\text{CH}_4$ ; 404, 1018, and 3961 ppm for  $\text{CO}_2$ ; and 0.2, 2.0, and 6.0 ppm for  $\text{N}_2\text{O}$ . The precision of measurements based on duplicate samples was  $\pm 3.9\%$  for  $\text{CH}_4$  and  $\pm 3.2\%$  for  $\text{N}_2\text{O}$ . The  $\text{CO}_2$  concentration is expressed as partial pressure in parts per million (ppm) and  $\text{CH}_4$  as dissolved concentration ( $\text{nmol L}^{-1}$ ), in accordance with convention in existing topical literature and because both quantities were systematically and distinctly above saturation level (400 ppm and 2–3  $\text{nmol L}^{-1}$ , respectively). Variations in  $\text{N}_2\text{O}$  were modest and concentrations fluctuated around atmospheric equilibrium, so data are presented as percent of saturation level (% $\text{N}_2\text{O}$ , where atmospheric equilibrium corresponds to 100 %), computed from the global mean  $\text{N}_2\text{O}$  air mixing ratios given by the Global Monitoring Division (GMD) of the Earth System Research Laboratory (ESRL) of the National Oceanic and Atmospheric Administration (NOAA) (<https://www.esrl.noaa.gov/gmd/hats/combined/N2O.html>, last access: 15 August 2018) and using the Henry's constant given by Weiss and Price (1980).

The flux ( $F$ ) of  $\text{CO}_2$  ( $F_{\text{CO}_2}$ ),  $\text{CH}_4$  ( $F_{\text{CH}_4}$ ) and  $\text{N}_2\text{O}$  ( $F_{\text{N}_2\text{O}}$ ) between surface waters and the atmosphere was computed according to Liss and Slater (1974):

$$F = k \Delta G, \quad (2)$$

where  $k$  is the gas transfer velocity ( $\text{cm h}^{-1}$ ) and  $\Delta G$  is the air–water gradient of a given gas.

Atmospheric  $p\text{CO}_2$  data from Mount Kenya station (NOAA ESRL GMD) and a constant atmospheric  $\text{CH}_4$  partial pressure of 2 ppm were used. Atmospheric mixing ratios given in dry air were converted to water-vapor-saturated air, using the water vapor formulation of Weiss and Price (1982) as a function of salinity and water temperature. The  $k$  normalized to a Schmidt number of 600 ( $k_{600}$  in  $\text{cm h}^{-1}$ ) were derived from the parameterization as a function slope and stream water velocity given by Eq. 5 of Raymond et al. (2012):

$$k_{600} = 8.42 + 11838 \times VS, \quad (3)$$

where  $V$  is stream velocity ( $\text{m s}^{-1}$ ) and  $S$  is slope (unitless).

We chose this parameterization because it is based on the most exhaustive compilation to date of  $k$  values in streams derived from tracer experiments and was used in the global riverine  $\text{CO}_2$  efflux estimates of Raymond et al. (2013) and Lauerwald et al. (2015). Values of  $V$  and  $S$  were derived from a geographical information system (GIS), as described in Sect. 2.6.

During the June 2014 field expedition, samples for the stable isotope composition of  $\text{CH}_4$  ( $\delta^{13}\text{C}-\text{CH}_4$ ) were collected and preserved similarly to those described above for the  $\text{CH}_4$  concentration. The  $\delta^{13}\text{C}-\text{CH}_4$  was determined with a custom developed interface, whereby a 20 mL He headspace was first created and  $\text{CH}_4$  was flushed out through a double-hole needle, non- $\text{CH}_4$  volatile organic compounds were trapped in liquid  $\text{N}_2$ ,  $\text{CO}_2$  was removed with a soda lime trap,  $\text{H}_2\text{O}$  was removed with a magnesium perchlorate trap, and the  $\text{CH}_4$  was quantitatively oxidized to  $\text{CO}_2$  in an online combustion column similar to that of an elemental analyzer. The resulting  $\text{CO}_2$  was subsequently pre-concentrated by immersion of a stainless steel loop in liquid  $\text{N}_2$ , passed through a micropacked GC column (Restek HayeSep Q 2 m length, 0.75 mm internal diameter) and finally measured on a Thermo DeltaV Advantage isotope ratio mass spectrometer (IRMS). Calibration was performed with  $\text{CO}_2$  generated from certified reference standards (IAEA-CO-1 or NBS-19 and LSVEC) and injected in the line after the  $\text{CO}_2$  trap. Reproducibility of measurements based on duplicate injections of samples was typically better than  $\pm 0.5\text{\textperthousand}$ .

The fraction of  $\text{CH}_4$  removed by methane oxidation ( $F_{\text{ox}}$ ) was calculated with a closed-system Rayleigh fractionation model (Liptay et al., 1998) according to

$$\ln(1 - F_{\text{ox}}) = \left[ \ln \left( \delta^{13}\text{C}-\text{CH}_4_{\text{initial}} + 1000 \right) - \ln \left( \delta^{13}\text{C}-\text{CH}_4 + 1000 \right) \right] / [\alpha - 1], \quad (4)$$

where  $\delta^{13}\text{C}-\text{CH}_4_{\text{initial}}$  is the signature of dissolved  $\text{CH}_4$  as produced by methanogenesis in sediments,  $\delta^{13}\text{C}-\text{CH}_4$  is the signature of dissolved  $\text{CH}_4$  in situ and  $\alpha$  is the fractionation factor.

We used a value of 1.02 for  $\alpha$  based on field measurements in a tropical lake (Morana et al., 2015). For  $\delta^{13}\text{C}-\text{CH}_4$ \_initial, we used a value of  $-60.2\text{\textperthousand}$ , which we measured in bubbles from the sediment trapped with a funnel on one occasion in a river dominated by *Vossia cuspidata* wetlands (16 April–6 May 2015). The model we used to compute  $F_{\text{ox}}$  applies for closed systems, implying that  $\text{CH}_4$  is assumed not to be exchanged with surroundings in contrast to open-system models. Running river water corresponds to a system that is intermediate between closed and open chemical systems, since it is open to the atmosphere and the sediments, but, on the other hand, the water parcel can be partly viewed as a closed system being transported downstream with the flow. As such, the water parcel receives a certain amount of  $\text{CH}_4$  from sediments and then is transported downstream away from the initial input of  $\text{CH}_4$ . We also applied two common open-system models to estimate  $F_{\text{ox}}$  given by Happel et al. (1994) and by Tyler et al. (1997) that have also been applied in lake systems (Bastviken et al., 2002). However, both open-system models gave  $F_{\text{ox}}$  values  $> 1$  in many cases (data not shown) since the difference between  $\delta^{13}\text{C}$  of the  $\text{CH}_4$  source and measured  $\delta^{13}\text{C}$  in dissolved  $\text{CH}_4$  was often much higher than expected from the assumed isotopic fractionation (1.02). The same observation ( $F_{\text{ox}} > 1$ ) was also reported with open-system models in the lakes studied by Bastviken et al. (2002). Since  $F_{\text{ox}}$  values  $> 1$  are not conceptually possible, we preferred to use the results from the closed-system model instead, although we acknowledge that flowing waters are in fact intermediary systems between closed and open and that, consequently, the computed  $F_{\text{ox}}$  values are likely underestimated.

Samples for the stable isotope composition of DIC ( $\delta^{13}\text{C}$ -DIC) were collected from the Niskin bottle with a silicone tube in 12 mL Exetainer vials (Labco) and poisoned with 50  $\mu\text{L}$  of a saturated solution of  $\text{HgCl}_2$ . Prior to the analysis of  $\delta^{13}\text{C}$ -DIC, a 2 mL helium headspace was created and 100  $\mu\text{L}$  of phosphoric acid ( $\text{H}_3\text{PO}_4$ , 99 %) was added in the vial in order to convert  $\text{CO}_3^{2-}$  and  $\text{HCO}_3^-$  to  $\text{CO}_2$ . After overnight equilibration, up to 1 mL of the headspace was injected with a gastight syringe into a coupled elemental analyzer – IRMS (EA-IRMS, Thermo FlashHT or Carlo Erba EA1110 with DeltaV Advantage). The obtained data were corrected for isotopic equilibration between dissolved and gaseous  $\text{CO}_2$ , as described by Gillikin and Bouillon (2007). Calibration was performed with certified standards (NBS-19 or IAEA-CO-1 and LSVEC). Reproducibility of measurements based on duplicate injections of samples was typically better than  $\pm 0.2\text{\textperthousand}$ .

Water was filtered on Whatman glass fiber filters (GF/F grade, 0.7  $\mu\text{m}$  porosity) for TSM (47 mm diameter), particulate organic carbon (POC) and particulate nitrogen (PN) (25 mm diameter) (precombusted at 450  $^{\circ}\text{C}$  for 5 h) and Chl *a* (47 mm diameter). Filters for TSM and POC were stored dry and filters for Chl *a* were stored frozen at  $-20\text{ }^{\circ}\text{C}$ . Filters for POC analysis were decarbonated with HCl fumes for 4 h and dried before encapsulation into silver cups; POC and PN

concentration and carbon stable isotope composition ( $\delta^{13}\text{C}$ -POC) were analyzed on an EA-IRMS (Thermo FlashHT with DeltaV Advantage), with a reproducibility better than  $\pm 0.2\text{\textperthousand}$  for stable isotopic composition and better than  $\pm 5\text{ \%}$  for bulk concentration of POC and PN. Data were calibrated with certified (IAEA-600: caffeine) and in-house standards (leucine and muscle tissue of Pacific tuna) that were previously calibrated versus certified standards. The Chl *a* samples were analyzed by high-performance liquid chromatography according to Descy et al. (2005), with a reproducibility of  $\pm 0.5\text{ \%}$  and a detection limit of  $0.01 \mu\text{g L}^{-1}$ . Part of the Chl *a* data were previously reported by Descy et al. (2017).

The water filtered through GF/F Whatman glass microfiber filters was collected and further filtered through polyethersulfone syringe encapsulated filters (0.2  $\mu\text{m}$  porosity) for stable isotope composition of O of  $\text{H}_2\text{O}$  ( $\delta^{18}\text{O}$ - $\text{H}_2\text{O}$ ), TA, dissolved organic carbon (DOC), major elements ( $\text{Na}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{K}^+$  and dissolved silicate, Si), nitrate ( $\text{NO}_3^-$ ), nitrite ( $\text{NO}_2^-$ ) and ammonium ( $\text{NH}_4^+$ ). Samples for  $\delta^{18}\text{O}$ - $\text{H}_2\text{O}$  were stored at ambient temperature in polypropylene 8 mL vials and measurements were carried out at the International Atomic Energy Agency (IAEA, Vienna), where water samples were pipetted into 2 mL vials and measured twice on different laser water isotope analyzers (Los Gatos Research or Picarro). Isotopic values were determined by averaging isotopic values from the last four out of nine injections, along with memory and drift corrections, with final normalization to the VSMOW/SLAP scales by using two-point lab standard calibrations, as fully described in Wassenaar et al. (2014) and Coplen and Wassenaar (2015). The long-term uncertainty for standard  $\delta^{18}\text{O}$  values was  $\pm 0.1\text{\textperthousand}$ . Samples for TA were stored at ambient temperature in polyethylene 55 mL vials and measurements were carried out by open-cell titration with  $\text{HCl}$  0.1 mol  $\text{L}^{-1}$  according to Gran (1952), and data quality was checked with certified reference material obtained from Andrew Dickson (Scripps Institution of Oceanography, University of California, San Diego, USA), with a typical reproducibility better than  $\pm 3 \mu\text{mol kg}^{-1}$ . DIC was computed from TA and  $p\text{CO}_2$  measurements using the carbonic acid dissociation constants for freshwater of Millero (1979) using the “CO2sys” package. Samples to determine DOC were stored at ambient temperature and in the dark in 40 mL brown borosilicate vials with polytetrafluoroethylene (PTFE)-coated septa and poisoned with 50  $\mu\text{L}$  of  $\text{H}_3\text{PO}_4$  (85 %), and DOC concentration was determined with a wet oxidation total organic carbon analyzer (IO Analytical Aurora 1030W), with a typical reproducibility better than  $\pm 5\text{ \%}$ . Part of the DOC data were previously reported by Lambert et al. (2016). Samples for major elements were stored in 20 mL scintillation vials and preserved with 50  $\mu\text{L}$  of  $\text{HNO}_3$  (65 %). Major elements were measured with inductively coupled plasma MS (ICP-MS; Agilent 7700x) calibrated with the following standards: SRM1640a from National Institute of Standards and Technology, TM-27.3 (lot 0412) and TMRain-04 (lot 0913) from Environ-

ment Canada, and SPS-SW2 Batch 130 from Spectrapure Standard. Limit of quantification was  $0.5 \mu\text{mol L}^{-1}$  for  $\text{Na}^+$ ,  $\text{Mg}^{2+}$ , and  $\text{Ca}^{2+}$ ;  $1.0 \mu\text{mol L}^{-1}$  for  $\text{K}^+$ ; and  $8 \mu\text{mol L}^{-1}$  for  $\text{Si}$ . Samples were collected during four cruises (8 May–6 June 2010, 20 November–8 December 2012, 3–19 December 2013, and 16 April–6 May 2015) for  $\text{NO}_3^-$ ,  $\text{NO}_2^-$  and  $\text{NH}_4^+$  and were stored frozen ( $-20^\circ\text{C}$ ) in 50 mL polypropylene vials.  $\text{NO}_3^-$  and  $\text{NO}_2^-$  were determined with the sulfanilamide, colorimetric was determined with the vanadium reduction method (APHA, 1998) and  $\text{NH}_4^+$  was determined with the dichloroisocyanurate-salicylate-nitroprussiate colorimetric method (SCA, 1981). Detection limits were 0.3, 0.01 and  $0.15 \mu\text{mol L}^{-1}$  for  $\text{NH}_4^+$ ,  $\text{NO}_2^-$  and  $\text{NO}_3^-$ , respectively. Precisions were  $\pm 0.02$ ,  $\pm 0.02$  and  $\pm 0.1 \mu\text{mol L}^{-1}$  for  $\text{NH}_4^+$ ,  $\text{NO}_2^-$  and  $\text{NO}_3^-$ , respectively.

## 2.6 GIS

The limits of the river catchments and Strahler order of rivers and streams were determined from the geospatial HydroSHEDS dataset (<https://hydrosheds.cr.usgs.gov/>, last access: 27 September 2018) using ArcGIS® (10.3.1). Land cover data were extracted from the global land cover (GLC) 2009 dataset ([http://due.esrin.esa.int/page\\_globcover.php](http://due.esrin.esa.int/page_globcover.php), last access: 11 January 2019) from the European Space Agency GlobCover 2009 project for the following classes: croplands, mosaic cropland/vegetation, dense forest, flooded dense forest, open forest/woodland, shrublands, mosaic forest or shrubland/grassland, grasslands, flooded grassland, and water bodies. Shrubland and grassland classes were aggregated for the estimate of savannah. Theoretical contribution of C<sub>4</sub> vegetation were extracted based on the vegetation δ<sup>13</sup>C isoscape for Africa from Still and Powell (2010) but corrected for agro-ecosystems according to the method of Powell et al. (2012).

The geospatial and statistical methods to compute river width, length, Strahler stream order, surface area, slope, flow velocity and discharge throughout the Congo River network are described in detail in the Supplement.

## 2.7 Statistical analysis

Statistical tests were carried out using GraphPad Prism® software at 0.05 level, and the normality of the distribution was tested with the D'Agostino–Pearson omnibus normality test.

## 3 Results and discussion

This section starts with the description of the spatial variations in general limnological variables as well as GHGs along the two main transects (Kinsangani–Kinshasa and Kwa) and as a function of stream order and the presence of the CCC. The following parts of this section deal with the seasonal variability and the drivers of CO<sub>2</sub> dynamics, based on one

hand on the metabolic measurements and on the other hand on stable isotopic composition of DIC. The final part of this section deals with fluxes of GHGs between the river and the atmosphere that are presented and discussed firstly as areal fluxes and finally as integrated fluxes at the scale of the basin and at global scale.

### 3.1 Spatial variations along the Kisangani–Kinshasa transect of general limnological variables and dissolved GHGs

Figures 3 and 4 show the spatial distribution of variables in surface waters of the main stem Congo River and confluence with tributaries along the Kisangani–Kinshasa transect during high water (HW, December 2013) and falling water (FW, June 2014) periods (Figs. 1 and 2). Numerous variables show a regular pattern in the main stem (increase or decrease) due to the gradual inputs from tributaries with a different (higher or lower) value than the main stem. Specific conductivity, TA, δ<sup>13</sup>C-DIC, pH, %O<sub>2</sub>, %N<sub>2</sub>O, TSM, pH and δ<sup>18</sup>O-H<sub>2</sub>O decreased, while pCO<sub>2</sub> and DOC increased in the main stem along the Kisangani–Kinshasa transect. Numerous tributaries had black-water characteristics (low conductivity, TA, pH, %O<sub>2</sub>, TSM and high DOC) while the main stem had generally white-water characteristics. The black-water tributaries were mainly found in the region of the CCC, while tributaries upstream or downstream of the CCC had in general more white-water characteristics. The differences between black-waters and white-waters were apparent in the patterns of continuous measurements of pCO<sub>2</sub>, showing a negative relationship with %O<sub>2</sub>, TSM, pH and specific conductivity (Fig. S4).

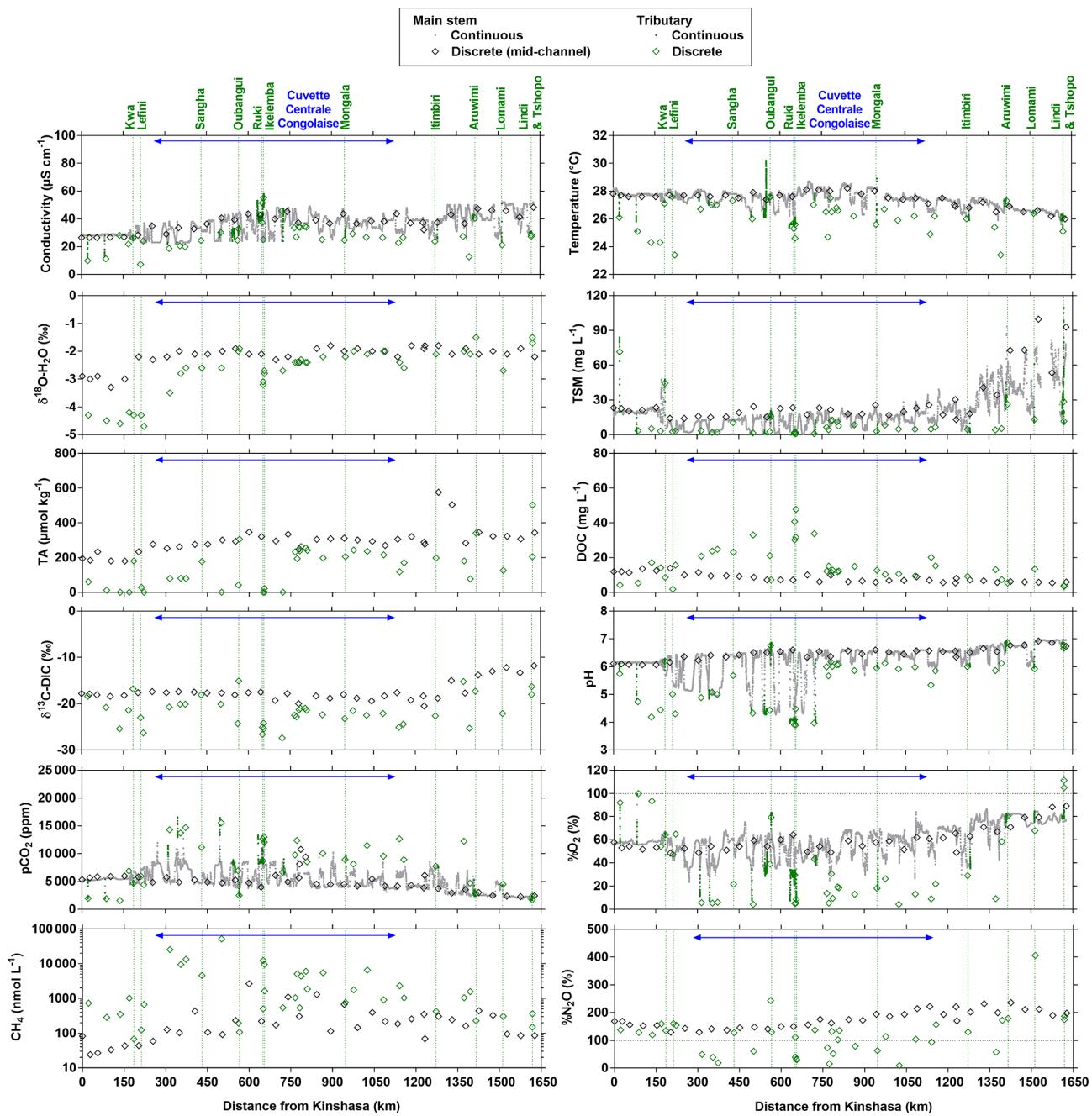
Specific conductivity in the main stem Congo River decreased from  $48.3 \text{ (HW)}$  and  $78.9 \text{ (FW)} \mu\text{S cm}^{-1}$  in Kisangani to  $26.5 \text{ (HW)}$  and  $32.7 \text{ (FW)} \mu\text{S cm}^{-1}$  in Kinshasa (Figs. 3 and 4). This decreasing pattern was related to a gradual dilution with tributary water with lower conductivities, on average  $27.6 \pm 9.9 \text{ (HW)}$  and  $31.9 \pm 17.8 \text{ (FW)} \mu\text{S cm}^{-1}$ . The lowest specific conductivity was measured in the Lefini River that is part of the Téké Plateau, where rainwater infiltrates into deep aquifers across thick sandy horizons leading to water with a low mineralization (Laraque et al., 1998). TA in the main stem decreased from  $344 \text{ (HW)}$  and  $697 \text{ (FW)} \mu\text{mol kg}^{-1}$  in Kisangani to  $195 \text{ (HW)}$  and  $269 \text{ (FW)} \mu\text{mol kg}^{-1}$  in Kinshasa, with an average in tributaries of  $141 \pm 119 \text{ (HW)}$  and  $136 \pm 141 \text{ (FW)} \mu\text{mol kg}^{-1}$ . δ<sup>13</sup>C-DIC roughly followed the patterns of TA, decreasing in the main stem from  $-7.9 \text{ (HW)}$  and  $-13.8 \text{ (FW)} \text{‰}$  in Kisangani to  $-11.8 \text{ (HW)}$  and  $-17.8 \text{ (FW)} \text{‰}$  in Kinshasa, with an average in tributaries of  $-21.7 \pm 3.2 \text{ (HW)}$  and  $-20.9 \pm 4.3 \text{ (FW)} \text{‰}$ . TSM in the main stem decreased from  $92.9 \text{ (HW)}$  and  $23.2 \text{ (FW)} \text{ mg L}^{-1}$  in Kisangani to  $45.4 \text{ (HW)}$  and  $18.2 \text{ (FW)} \text{ mg L}^{-1}$  in Kinshasa, with an average in tributaries of  $9.3 \pm 13.4 \text{ (HW)}$  and  $8.5 \pm 9.3 \text{ (FW)} \text{ mg L}^{-1}$ . The highest TSM values were recorded in the Kwa (44.4

(HW) and 15.8 (FW) mg L<sup>-1</sup>) and in the Nsele, a small stream-draining savannah and flowing into pool Malebo (71.4 [HW] and 34.8 [FW] mg L<sup>-1</sup>). pH in the main stem decreased from 6.73 (HW) and 7.38 (FW) in Kisangani to 6.11 (HW) and 6.29 (FW) in Kinshasa, with an average in tributaries of 5.44 ± 0.88 (HW) and 4.91 ± 1.05 (FW). %O<sub>2</sub> values in the main stem decreased from 89.2 (HW) and 92.6 (FW) % in Kisangani to 57.8 (HW) and 79.7 (FW) % in Kinshasa, with an average in tributaries of 36.7 ± 33.3 (HW) and 43.7 ± 35.8 (LW) %. DOC increased from 5.9 (HW) and 5.1 (FW) mg L<sup>-1</sup> in Kisangani to 11.9 (HW) and 9.4 (FW) mg L<sup>-1</sup> in Kinshasa, with an average in tributaries of 16.1 ± 10.6 (HW) and 17.9 ± 15.2 (FW) mg L<sup>-1</sup>. Extremely low pH values were recorded in rivers draining the CCC, with values as low as 3.6, coinciding with nearly anoxic conditions (%O<sub>2</sub> down to 0.3 %) in surface waters and very high DOC content (up to 67.8 mg L<sup>-1</sup>). δ<sup>18</sup>O-H<sub>2</sub>O in the main stem decreased from -2.2 (HW) and -1.6 (FW) ‰ in Kisangani to -2.9 (HW) and -2.2 (FW) ‰ in Kinshasa, with an average in the tributaries of -2.7 ± 0.9 (HW) and -2.2 ± 0.8 ‰ (FW). Temperature increased in the main stem from 26.0 (HW) and 27.1 (FW) °C in Kisangani to 27.8 (HW) and 27.3 (FW) °C in Kinshasa, due to exposure in the uncovered main stem to solar radiation, as temperature was lower in tributaries with an average of 26.0 ± 1.1 (HW) and 26.1 ± 1.7 (FW) °C due to more shaded conditions (forest cover).

The pCO<sub>2</sub> values in the main stem increased from 2424 (HW) and 1670 (FW) ppm in Kisangani to 5343 (HW) and 2896 (FW) ppm in Kinshasa, with an average in tributaries of 8306 ± 4089 (HW) and 8039 ± 5311 (FW) ppm. pCO<sub>2</sub> in tributaries was in general higher than in the main stem with a few exceptions, namely in rivers close to Kinshasa (1582 to 1903 [HW] and 1087 to 2483 [FW] ppm), due to degassing at waterfalls upstream of the sampling stations, as the terrain is more mountainous in this area than in more gently sloping catchments upstream and also possibly due to a larger contribution of savannah to the catchment cover. The highest pCO<sub>2</sub> values (up to 16 942 ppm) were observed in streams draining the CCC. CH<sub>4</sub> in the main stem decreased from 85 (HW) and 63 (FW) nmol L<sup>-1</sup> in Kisangani to 24 (HW) and 22 (FW) nmol L<sup>-1</sup> just before Kinshasa and then increased again in the Malebo pool (82 [HW] and 78 [FW] nmol L<sup>-1</sup>), possibly related to the shallowness (~3 m in Malebo pool versus ~30 m depth at station just upstream). The general decreasing pattern of CH<sub>4</sub> in the main stem resulted from CH<sub>4</sub> oxidation, as indicated by <sup>13</sup>C-enriched values in the main stem (-38.1 ‰ to -49.4 ‰) with regards to sediment CH<sub>4</sub> (-60.2 ‰) and the increasing <sup>13</sup>C enrichment along the transect (Fig. 5). The calculated fraction of oxidized CH<sub>4</sub> ranged between 0.43 and 0.68 in the main stem and also increased downstream along the transect (Fig. S5). CH<sub>4</sub> in the tributaries showed a very large range of CH<sub>4</sub> concentration (68 to 51 839 nmol L<sup>-1</sup> (HW) and 102 to 56 236 nmol L<sup>-1</sup> (FW)). CH<sub>4</sub> in the tributaries showed a variable degree of <sup>13</sup>C

enrichment compared to sediment CH<sub>4</sub> ( $\delta^{13}\text{C-CH}_4$  between -19.3 ‰ and -56.3 ‰, Fig. 5), and the calculated fraction of oxidized CH<sub>4</sub> ranged between 0.18 and 0.88 (Fig. S5). Unlike the large rivers of the Amazon where a loose negative relation has been reported (Sawakuchi et al., 2016), the  $\delta^{13}\text{C-CH}_4$  values in the Congo River were unrelated to dissolved CH<sub>4</sub> concentrations, and a relatively high <sup>13</sup>C enrichment ( $\delta^{13}\text{C-CH}_4$  up to -19.3 ‰) was observed even at high CH<sub>4</sub> concentrations (correspondingly 3118 nmol L<sup>-1</sup>) (Fig. 5). This lack of correlation between concentration and isotope composition of CH<sub>4</sub> was probably related to spatial heterogeneity of sedimentary (and corresponding water column) CH<sub>4</sub> content over a very large and heterogeneous sampling area. The highest CH<sub>4</sub> concentrations were observed at the mouths of small rivers in the CCC. At the confluence with the Congo main stem, the flow is slowed down, leading to the development of shallow delta-type systems with very dense coverage of aquatic macrophytes (in majority *Vossia cuspidata* with a variable contribution of *Eichhornia crassipes* but also other species, Fig. S6). Such sites are favorable for intense sediment methanogenesis but, due to the stable environment related to near stagnant waters, also very favorable for the establishment of a stable methanotrophic bacterial community in the water column and associated with the root system of floating macrophytes that sustain intense methane oxidation (Yoshida et al., 2014; Kosten et al., 2016). Indeed, we found a very strong relation between CH<sub>4</sub> oxidation and CH<sub>4</sub> concentration on a limited number of incubations carried out in the Kwa river network in April 2015 (Fig. S7). Such conditions can explain the apparently paradoxical combination of high CH<sub>4</sub> concentrations in some cases associated with a high degree of CH<sub>4</sub> oxidation. Microbial oxidation of CH<sub>4</sub> might also explain the occurrence of samples with extremely <sup>13</sup>C depleted POC ( $\delta^{13}\text{C-POC}$  down to -39.0 ‰) observed in low %O<sub>2</sub> and high CH<sub>4</sub> environments (Fig. 6) located in small streams of the CCC that were also characterized by low POC and TSM values (not shown) and high POC:Chl *a* ratios (excluding the possibility that in situ PP would be at the basis of the <sup>13</sup>C depletion). This suggests that in these environments poor in particles (typical of black-water streams) but with high CH<sub>4</sub> concentrations, methanotrophic bacteria that are able to incorporate <sup>13</sup>C-depleted CH<sub>4</sub> into their biomass contribute substantially to POC. While such patterns have been reported at the oxic-anoxic transition zone of lakes with high hypolimnetic CH<sub>4</sub> concentrations such as Lake Kivu (Morana et al., 2015), this has never been reported in surface waters of rivers.

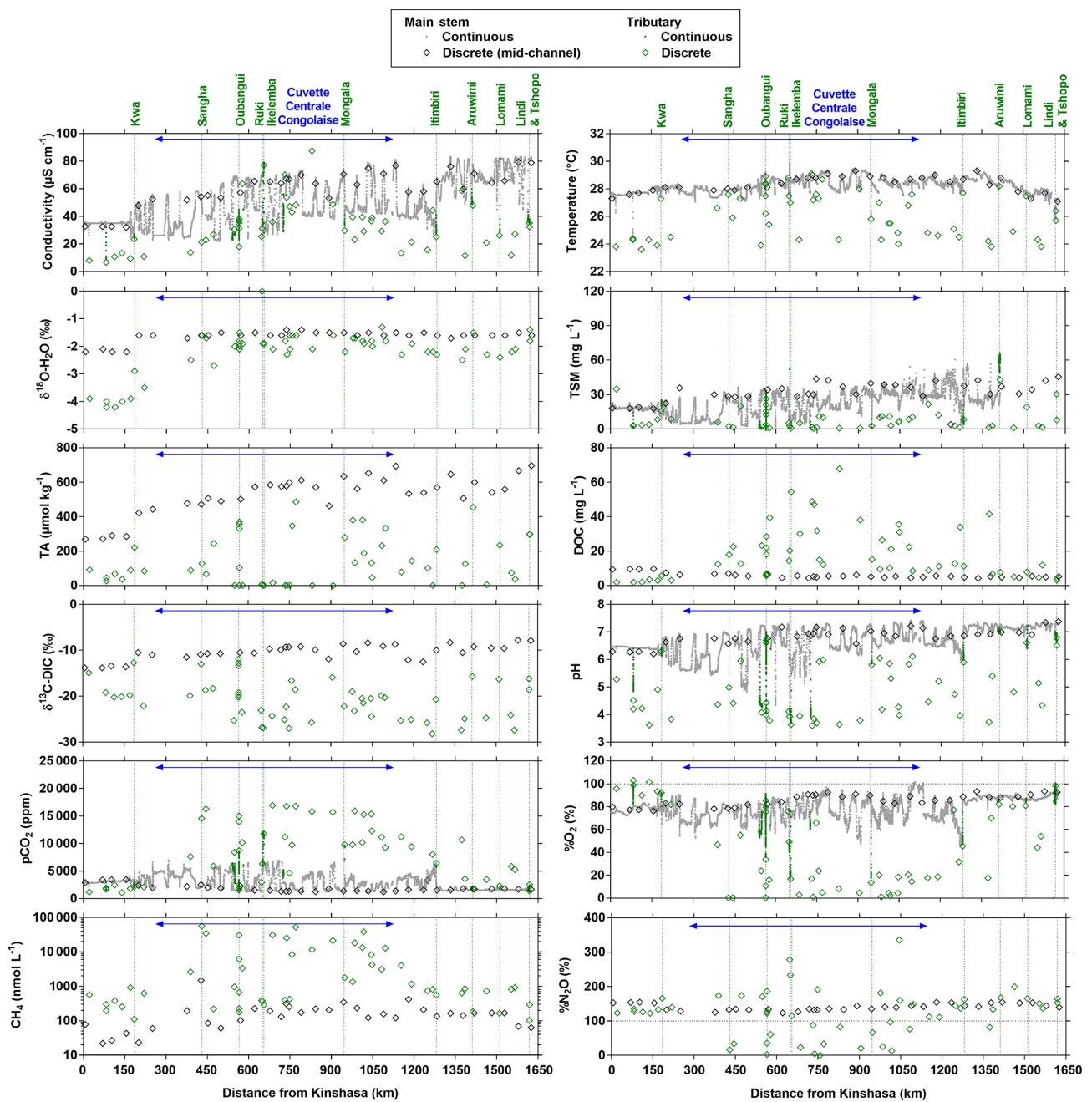
%N<sub>2</sub>O decreased from 198 (HW) and 139 (FW) % in Kisangani to 168 (HW) and 153 (FW) % in Kinshasa, and in most cases %N<sub>2</sub>O was lower in tributaries, on average 114 ± 73 (HW) and 120 ± 69 (FW) %. The undersaturation in N<sub>2</sub>O was observed in streams with high DOC, low %O<sub>2</sub> and relatively low NO<sub>3</sub><sup>-</sup> and is most probably related to denitrification of N<sub>2</sub>O, as also reported in the Amazon river (Richey et al., 1988) and in temperate rivers (Baulch et al.,



**Figure 3.** Variation in surface waters of the specific conductivity ( $\mu\text{S cm}^{-1}$ ), water temperature ( $^{\circ}\text{C}$ ), oxygen stable isotope composition of  $\text{H}_2\text{O}$  ( $\delta^{18}\text{O-H}_2\text{O}$  in ‰), total suspended matter (TSM in  $\text{mg L}^{-1}$ ), total alkalinity (TA in  $\mu\text{mol kg}^{-1}$ ), dissolved organic carbon (DOC in  $\text{mg L}^{-1}$ ), carbon stable isotope composition of dissolved inorganic carbon ( $\delta^{13}\text{C-DIC}$  in ‰), pH, partial pressure of  $\text{CO}_2$  ( $p\text{CO}_2$  in ppm), dissolved  $\text{O}_2$  saturation level ( $\% \text{O}_2$  in %), dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ) and dissolved  $\text{N}_2\text{O}$  saturation level ( $\% \text{N}_2\text{O}$  in %) as a function of the distance upstream of Kinshasa along a transect along the Congo River from Kisangani (3–19 December 2013,  $n = 10505$ ). Grey and black symbols indicate samples from the main stem and green samples from tributaries.

2011). Denitrification could have occurred in river sediments or soils, although the lowest  $\% \text{N}_2\text{O}$  (and  $\% \text{O}_2$ ) occurred in the CCC dominated by flooded soils in the flooded forest. Indeed, there was a general negative relationship between  $\% \text{N}_2\text{O}$  and  $\% \text{O}_2$  (Fig. 7), with an average  $\% \text{N}_2\text{O}$

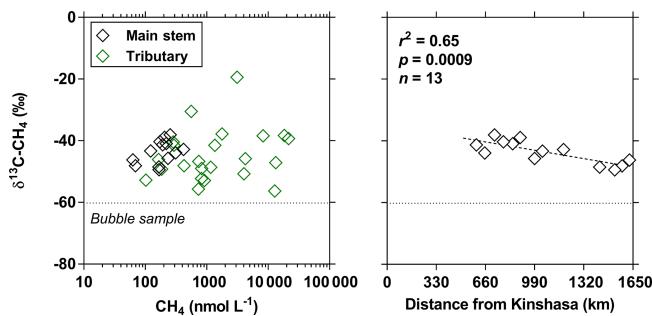
of  $78.5 \pm 59.3\%$  for  $\% \text{O}_2 < 25\%$  and an average  $\% \text{N}_2\text{O}$  of  $155.7 \pm 57.7\%$  for  $\% \text{O}_2 > 25\%$  (Mann–Whitney  $p < 0.0001$ ). The decreasing pattern of  $\text{NH}_4^+ : \text{DIN}$  and increasing pattern of  $\text{NO}_3^- : \text{DIN}$  with  $\% \text{O}_2$  indicated the occurrence of nitrification in oxygenated (typical of high Strahler stream order)



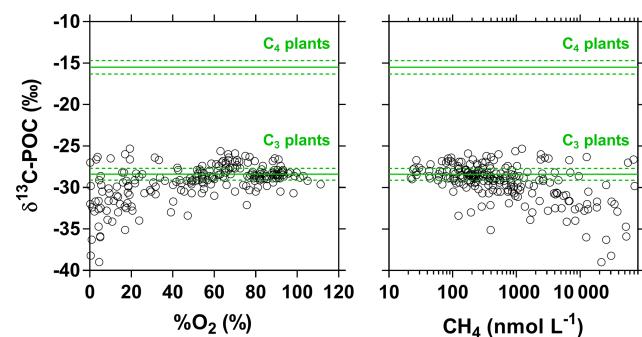
**Figure 4.** Variation in surface waters of the specific conductivity ( $\mu\text{S cm}^{-1}$ ), water temperature ( $^{\circ}\text{C}$ ), oxygen stable isotope composition of  $\text{H}_2\text{O}$  ( $\delta^{18}\text{O-H}_2\text{O}$  in ‰), total suspended matter (TSM in  $\text{mg L}^{-1}$ ), total alkalinity (TA in  $\mu\text{mol kg}^{-1}$ ), dissolved organic carbon (DOC in  $\text{mg L}^{-1}$ ), carbon stable isotope composition of dissolved inorganic carbon ( $\delta^{13}\text{C-DIC}$  in ‰), pH, partial pressure of  $\text{CO}_2$  ( $p\text{CO}_2$  in ppm), dissolved  $\text{O}_2$  saturation level ( $\% \text{O}_2$  in %), dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ) and dissolved  $\text{N}_2\text{O}$  saturation level ( $\% \text{N}_2\text{O}$  in %) as a function of the distance upstream of Kinshasa along a transect along the Congo River from Kisangani (10–30 June 2014,  $n = 12968$ ). Grey and black symbols indicate samples from the main stem and green samples from tributaries.

rivers and prevalence of  $\text{NH}_4^+$  in the more reducing and lower oxygenated (typical of low Strahler order) streams draining the CCC in particular, where  $\text{NO}_3^-$  was probably also removed from the water by sedimentary or soil denitrification (Fig. 7). In addition, in black-water rivers and streams, low

pH (down to 4) might have led to the inhibition of nitrification (Le et al., 2019) and also contributed higher  $\text{NH}_4^+:\text{DIN}$  values. Furthermore, the positive relation between  $\% \text{N}_2\text{O}$  and  $\text{NO}_3^-:\text{DIN}$  and the negative relation between  $\% \text{N}_2\text{O}$  and  $\text{NH}_4^+:\text{DIN}$  (Fig. 8) support the hypothesis of  $\text{N}_2\text{O}$  re-



**Figure 5.** Carbon stable isotope composition of  $\text{CH}_4$  ( $\delta^{13}\text{C-CH}_4$  in ‰) in surface waters of the Congo River main stem (black symbols) and tributaries (green symbols) as a function of dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ) and as a function of the distance upstream of Kinshasa, obtained along a longitudinal transect along the Congo River from Kisangani (10–30 June 2014). The dotted line indicates linear regression.

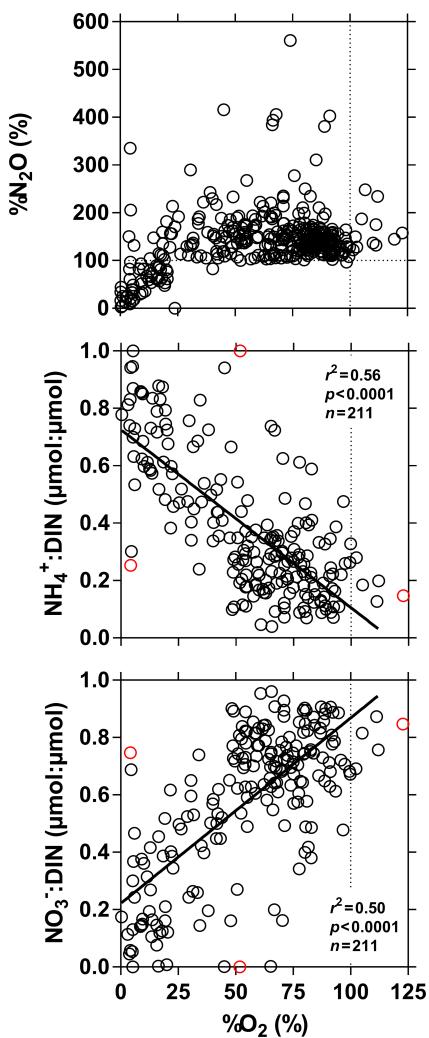


**Figure 6.** Carbon stable isotope composition of particulate organic carbon ( $\delta^{13}\text{C-POC}$  in ‰) in surface waters of the Congo River network as a function of dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %) and dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ). Grey lines indicate average (full line) and standard deviation (dotted lines) of average soil organic carbon stable isotope composition with a dominance of C<sub>4</sub> ( $-15.5 \pm 0.8\text{ ‰}$ ) and C<sub>3</sub> plants ( $-28.4 \pm 0.7\text{ ‰}$ ) (Bird and Pousai, 1997). Data with  $\delta^{13}\text{C-POC} < -30\text{ ‰}$  were associated with significantly lower %O<sub>2</sub> and higher  $\text{CH}_4$  than data with  $\delta^{13}\text{C-POC} > -30\text{ ‰}$  (Mann–Whitney  $p < 0.0001$  for both tests).

removal by denitrification in O<sub>2</sub>-depleted environments (either in stream sediments or soils), while in more oxygenated rivers N<sub>2</sub>O was produced by nitrification. In the main stem of the Congo, there might, in addition, be a loop of nitrogen recycling (ammonification-nitrification) sustained by phytoplankton growth and decay that contributed to maintain over-saturation of N<sub>2</sub>O with respect to atmospheric equilibrium, as phytoplankton growth was only observed in the main stem (Deschy et al., 2017). The generally low %N<sub>2</sub>O values in the Congo River network were probably due to the near pristine nature of these systems with low NH<sub>4</sub><sup>+</sup> ( $2.3 \pm 1.3\text{ }{\mu\text{mol L}}^{-1}$ ) and NO<sub>3</sub><sup>-</sup> ( $5.6 \pm 5.1\text{ }{\mu\text{mol L}}^{-1}$ ) levels, typical of rivers and streams draining a large fraction ( $\sim 70\%$ ) of forests. Croplands only represented at most 17 %, on average, of the land

cover of the studied river catchments where traditional agriculture is practised with little use of artificial fertilizers. This corresponds to an upper bound of cropland surface area since it was estimated aggregating the “cropland” and “mosaic cropland/vegetation” GLC 2009 categories, the latter corresponding to mixed surfaces with < 50 % of cropland. The “cropland” GLC 2009 category only accounts for 0.1 %, on average, of the land cover of the studied river catchments. Nitrogen inputs from waste water can also sustain N<sub>2</sub>O production in impacted rivers and streams (Marwick et al., 2014), but the largest cities along the Congo River main stem are of relatively modest size such as Kisangani (1 600 000 habitants) and Mbandaka (350 000 habitants), especially considering the large dilution due to the massive discharge of the main stem (sampling was done upstream of the influence of the megacity of Kinshasa with 11 900 000 habitants).

The input of the Kwa led to distinct changes of TA,  $\delta^{18}\text{O-H}_2\text{O}$  (decrease) and TSM (increase) of main stem values (comparing values upstream and downstream of the Kwa mouth) (Figs. 3–4). In the main stem, continuous measurements of  $p\text{CO}_2$ , pH, %O<sub>2</sub> and conductivity showed more variability compared to discrete samples acquired in the middle of channel, in particular in the region of CCC (Figs. 3–4). These patterns were related to gradients across the section of channel, as the boat sailed either along the mid-channel or closer to shore. The water from the tributaries flowed along the riverbanks and did not mix with main stem middle channel waters, as visible in natural color remotely sensed images (Fig. S8), leading to strong gradients across the section of main stem channel. During the June 2014 field expedition, this was investigated in more detail by a series of six transects perpendicular to the river main stem channel (Fig. 9). In the upper part (1590 km from Kinshasa), the variables showed little cross section gradients except for a decrease in conductivity towards the right bank due to inputs from the Lindi River that had distinctly lower specific conductivities ( $27.2$  [HW] and  $35.1$  [FW]  $\mu\text{S cm}^{-1}$ ) than the main stem ( $48.3$  [HW] and  $78.9$  [FW]  $\mu\text{S cm}^{-1}$ ). At 795 km from Kinshasa, marked gradients appeared in all variables, with the presence of black-water characteristics close to the right bank (higher  $p\text{CO}_2$  and lower %O<sub>2</sub>, pH, specific conductivity, temperature, and TSM values). This feature was related to inputs from large right-bank tributaries such as the Aruwimi and the Itimbiri (Supplement Table S1). Upstream of this section the only major left-bank tributary is the Lomami, which had white-water characteristics, relatively similar to those of the main stem (Figs. 3–4). The presence of black-water characteristics became apparent also on the left bank, from cross sections at 307 and 254 km upstream of Kinshasa, where the river is particularly wide (> 6 km wide) and received the inputs from the Ruki, the second-largest left-bank tributary (Table S1) with black-water characteristics. The cross section gradients became less marked at 203 km upstream from Kinshasa, as in this region the river becomes more narrow (2 km wide) leading to increased currents and more lateral

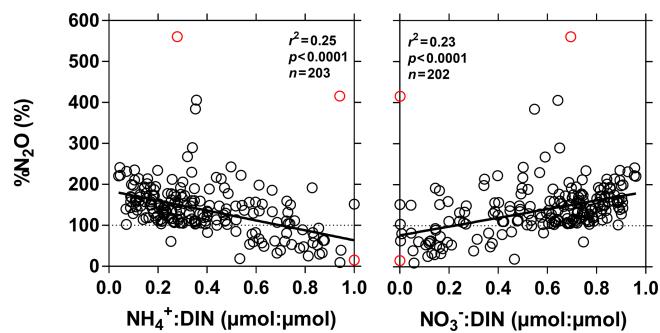


**Figure 7.** Dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %), ratio of NH<sub>4</sub><sup>+</sup> to dissolved inorganic nitrogen (DIN = NH<sub>4</sub><sup>+</sup> + NO<sub>2</sub><sup>-</sup> + NO<sub>3</sub><sup>-</sup>) (μmol : μmol) and ratio of NO<sub>3</sub><sup>-</sup> to DIN (μmol : μmol) in surface waters of the Congo River network as a function of dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %). Outliers (red dots) were identified with a Cook's distance procedure and removed prior to linear regression analysis (solid line).

mixing. The cross section gradients nearly disappeared at 158 km upstream of Kinshasa (and 30 km downstream of the Kwa mouth) due to homogenization by the large Kwa inputs (nearly 20 % of total freshwater discharge from the Congo River, Table S1) into a relatively narrow river section ( $\sim 1$  km).

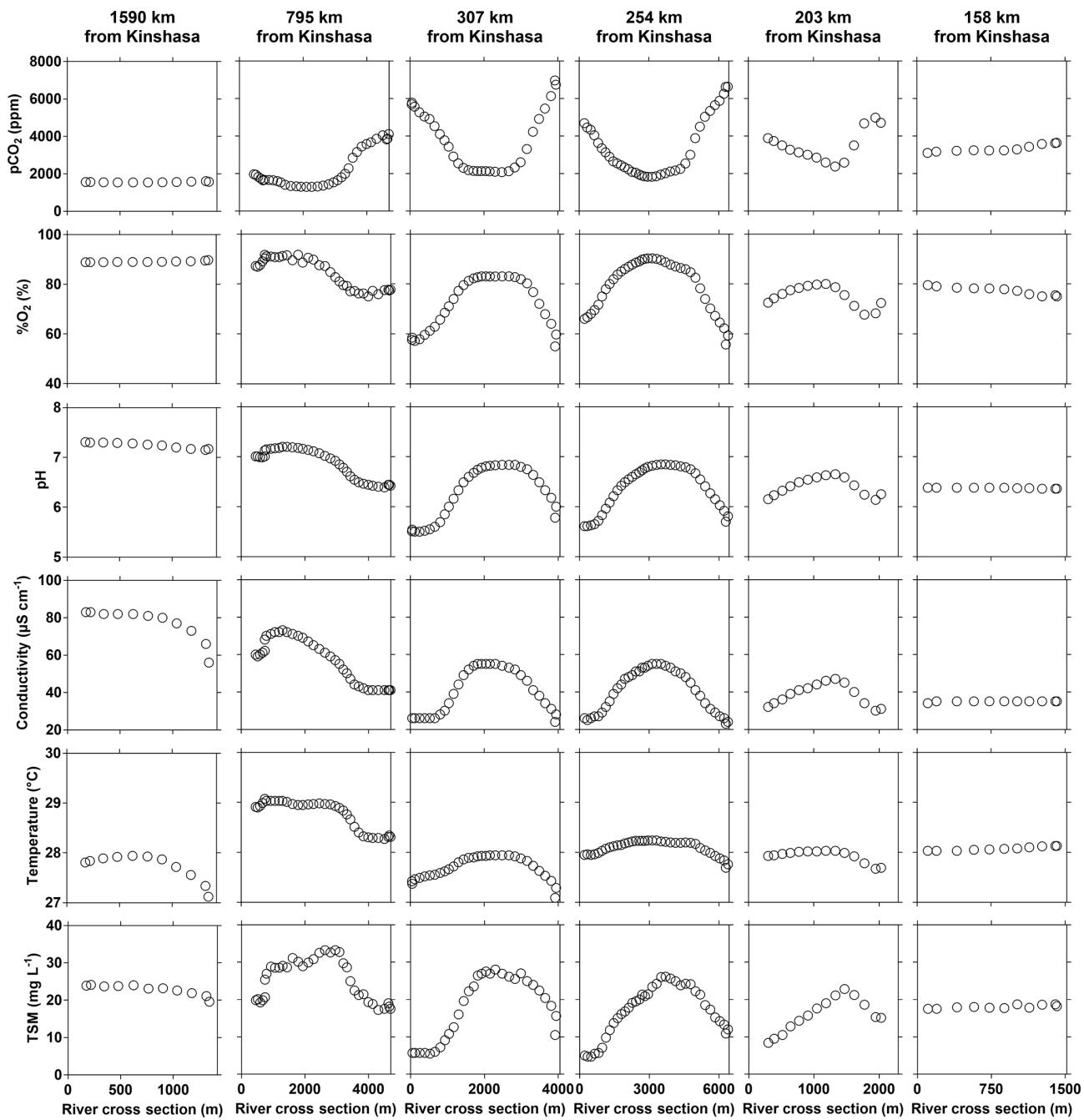
### 3.2 Spatial variations along the Kwa transect

Spatial features of biogeochemical variables along the transect in the Kwa River network (Fig. 10) showed some similarities with two Kisangani–Kinshasa transects along the Congo main stem (Figs. 3–4). The main stem Kwa



**Figure 8.** Dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %), as a function of the ratio of NH<sub>4</sub><sup>+</sup> to dissolved inorganic nitrogen (DIN = NH<sub>4</sub><sup>+</sup> + NO<sub>2</sub><sup>-</sup> + NO<sub>3</sub><sup>-</sup>) (μmol : μmol) and of the ratio of NO<sub>3</sub><sup>-</sup> to DIN (μmol : μmol) in surface waters of the Congo River network. Outliers (red dots) were identified with a Cook's distance procedure and removed prior to linear regression analysis (solid line).

had a higher specific conductivity than the tributaries ( $25.1 \pm 4.2$  versus  $21.2 \pm 11.5 \mu\text{S cm}^{-1}$ ), higher TA ( $281 \pm 64$  versus  $119 \pm 118 \mu\text{mol kg}^{-1}$ ), higher temperatures ( $27.7 \pm 0.7$  versus  $26.3 \pm 2.2^\circ\text{C}$ ), higher TSM ( $40.1 \pm 8.9$  versus  $15.2 \pm 35.1 \text{ mg L}^{-1}$ ), higher pH ( $6.1 \pm 0.2$  versus  $4.5 \pm 0.7$ ), higher %O<sub>2</sub> ( $67.0 \pm 7.0$  versus  $37.9 \pm 26.8 \%$ ), higher δ<sup>13</sup>C-DIC ( $-16.5 \pm 1.2$  versus  $-22.6 \pm 3.4 \text{ ‰}$ ) and lower DOC ( $5.2 \pm 1.4$  versus  $13.0 \pm 8.5 \text{ mg L}^{-1}$ ). Unlike the Kisangani–Kinshasa transects along the Congo main stem, the δ<sup>18</sup>O-H<sub>2</sub>O was lower in the main stem Kwa ( $-4.3 \pm 0.3 \text{ ‰}$ ) than in the tributaries ( $-3.3 \pm 0.6 \text{ ‰}$ ). Note that both the main upstream branches of the Kwa (Kasai and Sankuru) had low δ<sup>18</sup>O-H<sub>2</sub>O values. The δ<sup>18</sup>O-H<sub>2</sub>O values of the upper Kasai and Sankuru ( $-4.4 \text{ ‰}$  and  $-4.7 \text{ ‰}$ , respectively) were lower than those of the Lualaba (Congo at Kisangani) ( $-2.2$  [HW] and  $-1.6$  [FW] ‰). This difference can be in part explained by the spatial patterns of δ<sup>18</sup>O-H<sub>2</sub>O in rainwater, as the annual averages over of river catchments are  $-3.1 \text{ ‰}$  and  $-4.2 \text{ ‰}$ , for the Lualaba and the Kasai, respectively, based on the global grids of the O isotope composition of precipitation given by Bowen et al. (2005). The river δ<sup>18</sup>O-H<sub>2</sub>O was close to those of rain for the Kasai but less depleted in <sup>18</sup>O for the Lualaba. This was probably related to the lower evapotranspiration over the catchments of the upper Kasai and Sankuru than those of Lualaba (Bultot, 1972), leading to more <sup>18</sup>O-depleted water (e.g., Simpson and Herczeg, 1991). Additionally, the catchment of Kwa has a high fraction of unconsolidated sedimentary (41.4 %) and siliciclastic sedimentary (44.3 %) rocks than the catchment of the Lualaba that is dominated by metamorphic rocks (68.4 %). Unconsolidated sedimentary and siliciclastic sedimentary rocks are more favorable to the infiltration of water and development of aquifers that will minimize evaporation and <sup>18</sup>O enrichment, unlike catchments dominated by metamorphic rocks. Note that the tributaries with the lowest δ<sup>18</sup>O-H<sub>2</sub>O values were situated downstream of the Kwa and upstream of Kinshasa

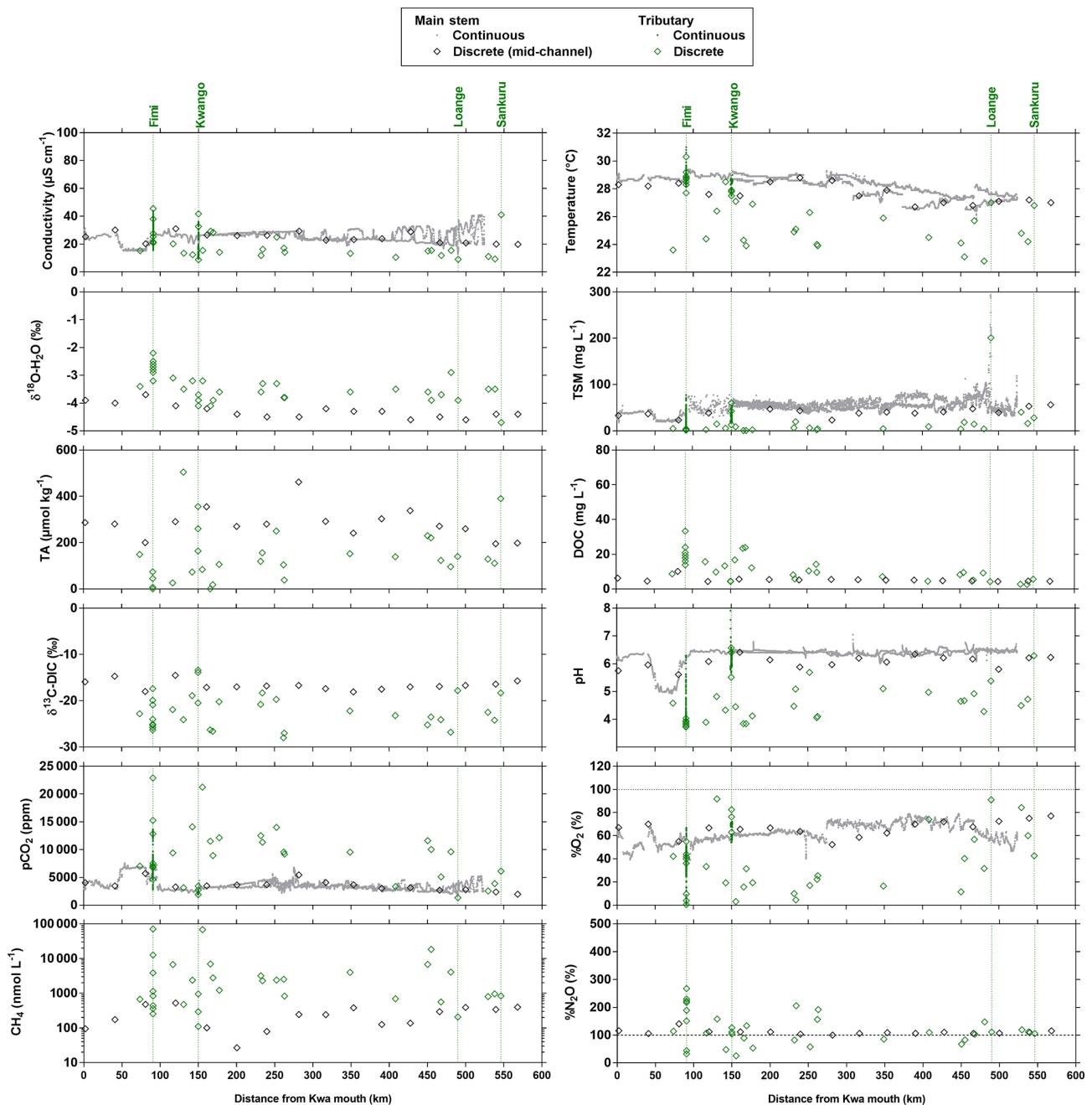


**Figure 9.** Variations in surface water of the partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm), dissolved oxygen saturation level (%O<sub>2</sub> in %), pH, specific conductivity ( $\mu\text{S cm}^{-1}$ ), water temperature (°C) and total suspended matter (TSM in  $\text{mg L}^{-1}$ ) along perpendicular transects to the main stem Congo River as a function of distance from the left bank (m) and at a variable distance from Kinshasa (10–30 June 2014).

(Figs. 3–4) and are part of the Téké Plateau. These rivers are fed by deep aquifers derived from infiltration of rain through sandy soils (Laraque et al., 1998).

Another difference with the Kisangani–Kinshasa transects along the Congo main stem relates to N<sub>2</sub>O values that were closer to saturation in the Kwa main stem ( $110.1 \pm 8.8\%$ ), while surface waters oscillated from undersaturation to over-

saturation in the tributaries ( $122.4 \pm 59.5\%$ ). This difference could be due to variations in biogeochemical cycling or in physical settings leading to changes in  $k$ . The latter seems more likely due to the strong flow in the Kwa that probably led to high gas transfer velocities and strong degassing of N<sub>2</sub>O to the atmosphere. In the Kwa main stem,  $p\text{CO}_2$  ( $3473 \pm 974$  ppm) and CH<sub>4</sub> ( $255 \pm 150$  nmol L<sup>-1</sup>) val-



**Figure 10.** Variation in surface waters of the specific conductivity ( $\mu\text{S cm}^{-1}$ ), water temperature ( $^{\circ}\text{C}$ ), oxygen stable isotope composition of H<sub>2</sub>O ( $\delta^{18}\text{O-H}_2\text{O}$  in ‰), total suspended matter (TSM in  $\text{mg L}^{-1}$ ), total alkalinity (TA in  $\mu\text{mol kg}^{-1}$ ), dissolved organic carbon (DOC in  $\text{mg L}^{-1}$ ), carbon stable isotope composition of dissolved inorganic carbon ( $\delta^{13}\text{C-DIC}$  in ‰), pH, partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm), dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %), dissolved CH<sub>4</sub> concentration ( $\text{nmol L}^{-1}$ ) and dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %) as a function of the distance upstream of the Kwa mouth along a transect along the Kwa River (16 April–6 May 2015,  $n = 7017$ ). Grey and black symbols indicate samples from the main stem and green samples from tributaries.

ues were lower than in tributaries ( $8804 \pm 5108$  ppm,  $6783 \pm 16479$  nmol L<sup>-1</sup>). The highest  $p\text{CO}_2$  and CH<sub>4</sub> values of the entire dataset in the Congo River network were observed in a tributary of the Fimi (22 899 ppm and 71 428 nmol L<sup>-1</sup>) that is bordered by very extensive meadows of the aquatic macro-

phyte *Vossia cuspidata* and unrelated to inputs from the shallow Lake Mai Ndombé that showed lower  $p\text{CO}_2$  and CH<sub>4</sub> values (3,143 ppm and 250 nmol L<sup>-1</sup>, respectively).

### 3.3 Spatial variations as a function of stream order and as a function of the influence of the CCC

The large differences in  $p\text{CO}_2$ ,  $\text{CH}_4$ ,  $\%O_2$  and  $\%N_2O$  (Figs. 3, 4, 10) among the various sampled tributaries of the Congo River can be analyzed in terms of size classes as given by Strahler order (Fig. 11). There were distinct patterns in  $\text{CH}_4$  and  $p\text{CO}_2$  versus Strahler order, with a decrease in the central value (median and average) for both quantities as a function of Strahler order in streams draining and not draining the CCC (Fig. 11). For nearly all the stream orders, the streams draining the CCC had significantly higher  $p\text{CO}_2$  and  $\text{CH}_4$  values than streams not draining the CCC (Fig. 11). The  $\%O_2$  values per Strahler order did not show any distinct pattern (increase or decrease) in the streams not draining the CCC, but the streams draining the CCC showed an increasing pattern as a function of Strahler order. The  $\%O_2$  values were significantly lower in the streams draining the CCC than those not draining the CCC (Fig. 11). For  $\%N_2O$ , the tendency of the central value (median and average) as a function of Strahler order did not show a clear pattern for streams not draining the CCC; however, there was a clear increasing pattern with Strahler order for streams draining the CCC. In addition, the  $\%N_2O$  values were significantly lower for half of the cases, in streams draining the CCC compared to those not draining it (Fig. 11).

In US rivers, a decreasing pattern as a function of Strahler order has previously been reported for  $p\text{CO}_2$  (Butman and Raymond, 2001; Liu and Raymond, 2018). This has been interpreted as reflecting inputs of soil-water enriched in terrestrial respiration  $\text{CO}_2$  that have a stronger impact in smaller and lower Strahler order systems, in particular headwater streams, followed by degassing of  $\text{CO}_2$  in higher Strahler order rivers (Hotchkiss et al., 2015), although soil-water  $\text{CO}_2$  inputs in headwater streams are seasonally variable and spatially heterogeneous (Duvert et al., 2018). Nevertheless, all of the low-Strahler-order streams we sampled were in lowlands, so the decreasing pattern of  $p\text{CO}_2$  as a function of Strahler order could alternatively reflect the stronger influence of riparian wetlands on smaller streams, rather than larger systems. The mechanism remains the same, a high ratio of lateral inputs to water volume in small streams that is related to soil-water in temperate streams such as in the US but related in addition to riparian wetlands in tropical systems such as those sampled in the Congo River network.

The influence of riparian wetlands on stream  $p\text{CO}_2$ ,  $\text{CH}_4$  and  $\%N_2O$  can be also highlighted when data were separated into rivers draining or not draining the CCC but aggregating into systems smaller or larger than Strahler order 5 to account simultaneously for the effect of stream size (Fig. 12). The  $p\text{CO}_2$  values were statistically higher in rivers draining the CCC than those not draining it, with median values more than 2-fold higher in both small and large rivers. Conversely,  $\%O_2$  levels were statistically lower in rivers draining the CCC than those not draining it, with median val-

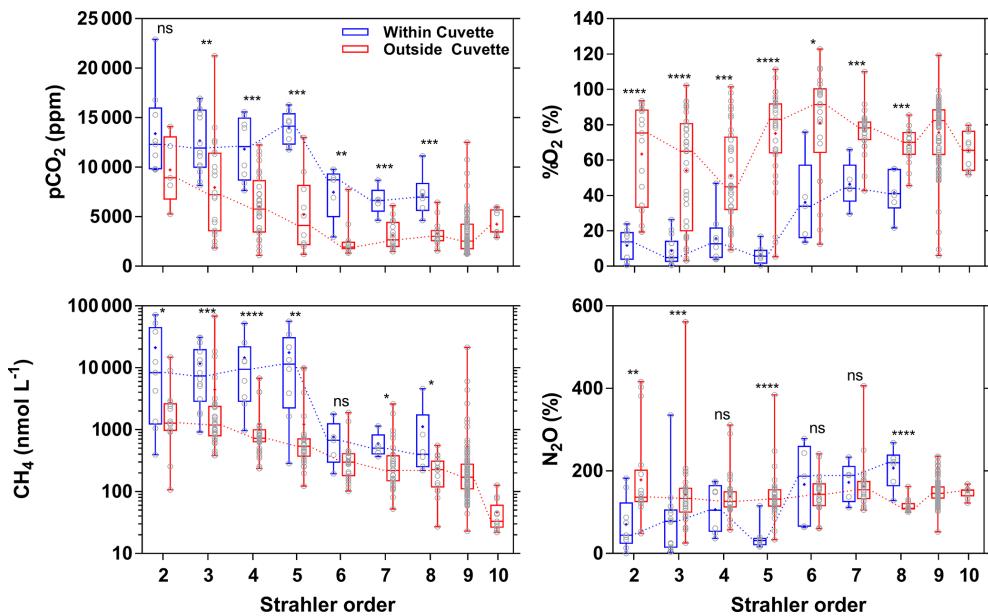
ues 11-fold and 2-fold lower in small and large rivers, respectively. Additional evidence on the influence of the connectivity of wetlands with rivers in sustaining high  $p\text{CO}_2$  and low  $\%O_2$  values was provided by the positive relationship between  $p\text{CO}_2$  and flooded dense forest cover and the converse negative relationship between  $\%O_2$  and flooded dense forest cover (Fig. 13). These patterns were also consistent with the positive relation between DOC concentration and flooded dense forest reported by Lambert et al. (2016). Note that aquatic macrophytes (*Vossia cuspidata*) also most probably strongly contributed, in addition to flooded forest, to high  $p\text{CO}_2$  and low  $\%O_2$  levels, based on visual observations of dense coverage (Fig. S9), although macrophytes have not been systematically mapped and GIS data are unavailable (as for flooded dense forest).

Wetlands coverage had also a major importance on  $\text{CH}_4$  distribution, as the  $\text{CH}_4$  values were statistically higher in rivers draining the CCC than those not draining it (Fig. 12), with median values 10-fold and 2-fold higher in small and large rivers, respectively.  $\%N_2O$  was also statistically lower in rivers draining the CCC than those not draining it, with median values 2.7-fold lower in small rivers but 1.4-fold higher in large rivers. The pattern of  $\%N_2O$  followed the one of  $\%O_2$ , and the very low to null  $\text{N}_2O$  values were observed in the systems draining the CCC where the lowest  $\%O_2$  values (close to 0) were also observed due to sedimentary or soil organic matter degradation, leading to a decrease in  $O_2$  and  $\text{N}_2O$  in surface waters (consumption of  $\text{N}_2O$  by denitrification).

The  $\text{CH}_4 : \text{CO}_2$  molar ratio ranged between 0.0001 and 0.1215, with a mean of  $0.0097 \pm 0.018$ . Such ratios were distinctly higher than those typically observed in marine waters (0.0005) and in the atmosphere (0.005). The  $\text{CH}_4 : \text{CO}_2$  molar ratio strongly increased with the decrease in  $\%O_2$  and was significantly higher in small rivers draining the CCC (Fig. 14). These patterns were probably related to inputs of organic matter from wetlands and in particular aquatic macrophytes that lead to important organic matter transfer to sediments and high sedimentary degradation of organic matter. This led to  $\%O_2$  decrease in surface waters and a large fraction of organic matter degradation by anaerobic processes compared to aerobic degradation, leading to an increase in the  $\text{CH}_4 : \text{CO}_2$  ratio. The decrease in  $O_2$  and increase in  $\text{CO}_2$  in the water in presence of floating macrophytes was probably in part also related to autotrophic root respiration and not fully related to microbial heterotrophic respiration.

### 3.4 Drivers of $\text{CO}_2$ dynamics – metabolic measurements and daily variations in $\text{CO}_2$

We compared the balance of depth-integrated planktonic PP to water column CR and compared it to  $\text{FCO}_2$  to test if in situ net heterotrophy was sufficient to sustain the emissions of  $\text{CO}_2$  to the atmosphere (Fig. 15), or if alterna-

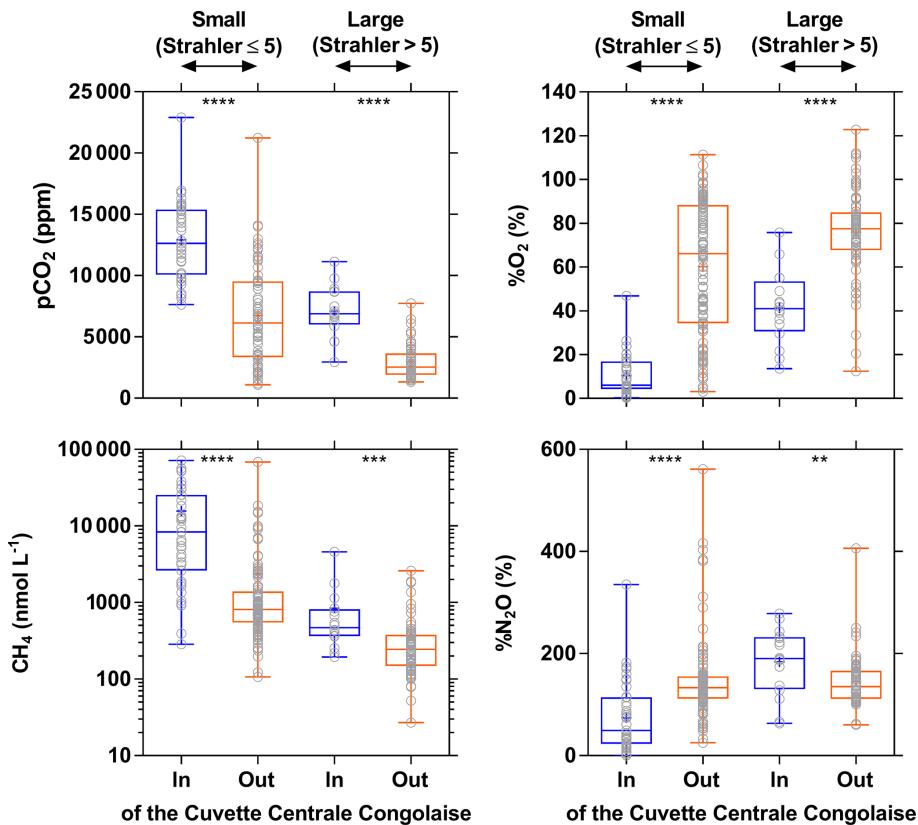


**Figure 11.** Box plot as a function of Strahler stream order of the partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm), dissolved CH<sub>4</sub> concentration (nmol L<sup>-1</sup>), dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %), and dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %) for rivers and streams of the Congo River network draining and not draining the Cuvette Centrale Congolaise. The box represents the first and third quartile, the horizontal line corresponds to the median, the cross corresponds to the average, the error bars correspond to the maximum and minimum, and the symbols show all data points. A Mann–Whitney test was used to test statistical differences: ns represents not significant, \*\*\*\* =  $p < 0.0001$ ; \*\*\* =  $p < 0.001$ ; \*\* =  $p < 0.01$ ; \* =  $p < 0.05$ .

tively fluvial CO<sub>2</sub> had a lateral origin (soils or wetlands). A detailed description of spatial and seasonal variations in PP as well as main phytoplankton communities is given by Descy et al. (2017). In brief, phytoplankton biomass was mainly confined to the main stem and was low in most tributaries. The PP values in the Congo River ranged between 0.0 and 57.5 mmol m<sup>-2</sup> d<sup>-1</sup> and were higher than previously reported in tropical river channels, whereas in other tropical rivers phytoplankton production mainly occurred in the floodplain lakes. This is due to generally lower TSM values in the Congo and to its relative shallowness that allows net phytoplankton growth in the mainstream unlike other deeper and more turbid tropical rivers such as the Amazon. Measured CR was lower than PP 3 out of 49 times, and on average the PP : CR ratio was 0.28. Volumetric rates of CR ranged between 0.7 and 46.6 mmol m<sup>-3</sup> d<sup>-1</sup>, while integrated rates of CR ranged between 3.1 and 790.4 mmol m<sup>-2</sup> d<sup>-1</sup>. CR was unrelated to TSM, POC, NH<sub>4</sub><sup>+</sup> and Chl *a* but showed a positive relation with DOC after binning the data (Fig. S10). The same pattern emerged when using modeled PP, to extend the number of data points, with PP higher than CR 2 out of 169 times and a PP : CR ratio of 0.15 on average. This indicates that a generalized and strongly net heterotrophic metabolism was encountered in the sampled sites. Yet, in 174 out of 187 cases,  $F\text{CO}_2$  was higher than CR, and in 162 out of 169 cases,  $F\text{CO}_2$  was higher than net community production (NCP). CR averaged 81 mmol m<sup>-2</sup> d<sup>-1</sup>, NCP averaged

–75 mmol m<sup>-2</sup> d<sup>-1</sup> and the corresponding average  $F\text{CO}_2$  was 740 mmol m<sup>-2</sup> d<sup>-1</sup>. The  $F\text{CO}_2$  : CR ratio was higher in lower-order streams than in higher-order streams, with median values ranging between 21 and 139 in stream orders 2–5 and between 3 and 17 in stream orders 6–10 (Fig. 16). This indicates a prevalence of lateral CO<sub>2</sub> inputs either from soil-water or riparian wetlands in sustaining  $F\text{CO}_2$  in lower-order streams over higher-order streams where in-stream CO<sub>2</sub> production from net heterotrophy is more important. These patterns are in general agreement with the conceptual framework developed by Hotchkiss et al. (2015), although lateral CO<sub>2</sub> inputs were exclusively attributed by these authors to soil-water or ground-water inputs and riparian wetlands were not considered. These patterns are also in agreement with the results reported by Ward et al. (2018), who show that in large high-order rivers of the lower Amazon, in-stream production of CO<sub>2</sub> from respiration is sufficient to sustain CO<sub>2</sub> emissions to the atmosphere.

CR was estimated from measurements of O<sub>2</sub> concentration decrease in bottles that were not rotated, and this has been shown to lead to an underestimation of CR up to a factor of 2 (Richardson et al., 2013; Ward et al., 2018). The underestimation of our CR measurements due to the absence of rotation is most likely not as severe as in the Richardson et al. (2013) and Ward et al. (2018) studies, as the organic matter in our samples was mostly in dissolved form (median DOC of 8.6 mg L<sup>-1</sup>), with a low particulate load (me-

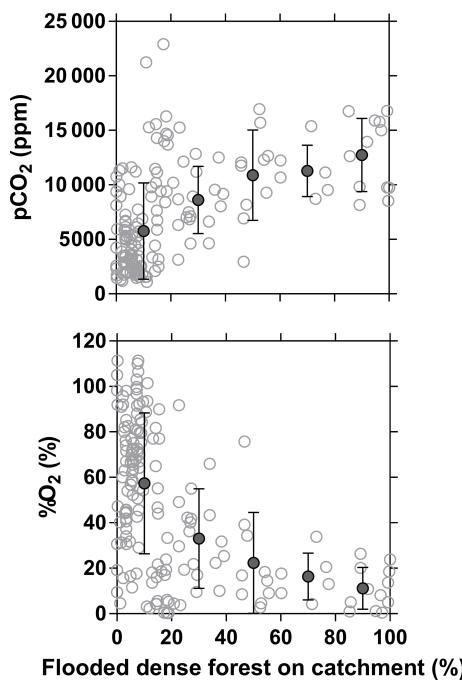


**Figure 12.** Box plot of the partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm), dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %), dissolved CH<sub>4</sub> concentration (nmol L<sup>-1</sup>), and dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %) in surface waters of rivers and streams of the Congo river network draining and not draining the Cuvette Centrale Congolaise for small and large systems (Strahler stream order 5 ≤ and > 5, respectively) (3–19 December 2013; 10–30 June 2014). The box represents the first and third quartile, the horizontal line corresponds to the median, the cross corresponds to the average, the error bars correspond to the maximum and minimum, and the symbols show all data points. A Mann–Whitney test was used to test statistical differences: \*\*\*\* =  $p < 0.0001$ ; \*\*\* =  $p < 0.001$ ; \*\* =  $p < 0.01$ .

dian TSM of 14 mg L<sup>-1</sup> and POC of 1.3 mg L<sup>-1</sup>), while the median of TSM at the sites studied by Ward et al. (2018) in the Amazon was higher (28.5 mg L<sup>-1</sup> based on data reported by Ward et al., 2015). We acknowledge that our CR measurements might be underestimated due to bottle effects and lack of rotation up to a factor of 2 based on the studies of Richardson et al. (2013) and Ward et al. (2018); nevertheless, it seems unrealistic to envisage an underestimation of CR by a factor of 10 that would allow reconciling the CR (and NCP) estimates with those of FCO<sub>2</sub>. Although we did not measure sediment respiration, the average value reported by Cardoso et al. (2014) of 21 mmol m<sup>-2</sup> d<sup>-1</sup> for tropical rivers and streams does not allow accounting for the imbalance between FCO<sub>2</sub> and NCP. This then suggests that the emission of CO<sub>2</sub> from the Congo lowland river network is to a large extent sustained by lateral inputs rather than by in-stream production of CO<sub>2</sub> by net heterotrophy. It remains to be determined to what extent this lateral input of CO<sub>2</sub> is sustained by riparian wetlands or soil-groundwater from terra firme.

The low PP : CR ratio of 0.15 to 0.28 on average, and generally low PP values (on average 12 mmol m<sup>-2</sup> d<sup>-1</sup>) were

also reflected in the low diurnal variations in  $p\text{CO}_2$ . We did not carry out dedicated 24 h cycles to look at the day–night variability of  $p\text{CO}_2$ , due to lack of opportunity given the important navigation time to cover large distances, but we compared the data acquired at the anchoring site on shore (typically around 17:00 universal time, UT, just before dusk) with the data on the same spot the next day (typically around 04:30 UT, just after dawn) (Fig. S11). Unsurprisingly, water temperature measured just before dusk was significantly higher than just after dawn (on average 0.5 °C higher), while specific conductivity was not significantly different, indicating that the same water mass was sampled at dusk and dawn (Fig. S11). The  $p\text{CO}_2$  and O<sub>2</sub> concentration measured just before dusk were not significantly different than just after dawn, showing that daily variability in these variables was low (Fig. S11). The difference between  $p\text{CO}_2$  at dusk and dawn ranged between −2307 and 1186 ppm and averaged 39 ppm ( $n = 39$ ). The wide range of values of the difference of  $p\text{CO}_2$  at dusk and at dawn might reflect occasional small-scale variability of  $p\text{CO}_2$ , as the boat anchored for the night close to shore, frequently in close proximity to riparian veg-



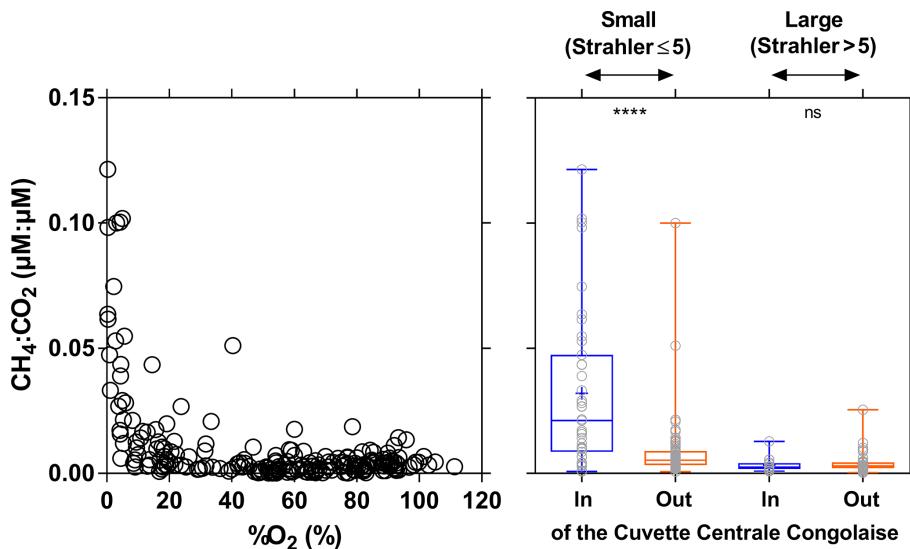
**Figure 13.** Partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm) and dissolved O<sub>2</sub> saturation level (%O<sub>2</sub> in %) in surface waters of rivers and streams of the Congo River network as a function of the flooded dense forest over the respective catchment (GLC, 2009). Grey open dots are individual data points, and black full dots are binned averages ( $\pm$  standard deviation) by intervals of 20 %.

etation. Nevertheless, the average difference is not of the expected sign (in the case of a strong diurnal change of  $p\text{CO}_2$  due to PP and CR,  $p\text{CO}_2$  should have been lower at dusk than dawn, so the difference should have been negative). This difference was also very small compared to the overall range of spatial variations in  $p\text{CO}_2$  (1087 to 22 899 ppm). Day–night variations in  $p\text{CO}_2$  have been reported in temperate headwater and low-order streams and in one lowland river with an amplitude from  $\sim 50$  to  $\sim 700$  ppm (Lynch et al., 2010; Dinsmore et al., 2013; Peter et al., 2014; Crawford et al., 2017; Reiman and Xu, 2019), although daily signals of  $p\text{CO}_2$  were not systematically observed and were absent for instance in streams covered by forest canopy (Crawford et al., 2017). In a low turbidity and very shallow low-order stream of the Tana River network, Tamoooh et al. (2013) reported on one occasion day–night variation in  $p\text{CO}_2$  with an amplitude of  $\sim 400$  ppm, and in the Zambezi river, during the dry season, corresponding to very low TSM values ( $< 10 \text{ mg L}^{-1}$ ), Teodoru et al. (2015) reported day–night  $p\text{CO}_2$  variation in the range of 475 ppm. In both cases, the day–night variations were also small compared to spatial variations of 300 to 5204 ppm in the Tana and 300 to 14 004 ppm in the Zambezi. In floodplain lakes of the Amazon, daily variations in  $p\text{CO}_2$  can be intense (with an amplitude up to  $\sim 2000$  ppm) during cyanobacterial blooms (Abril et al., 2013; Amaral et

al., 2018) but have not been documented in the river channels of the Amazon. Our data show that in nutrient-poor and light-limited lowland tropical rivers such as the Congo River, where pelagic PP is low, day–night variations in  $p\text{CO}_2$  were negligible compared to spatial variations in  $p\text{CO}_2$ . We conclude that accounting for day–night variations in  $p\text{CO}_2$  should not lead to a dramatic revision of global CO<sub>2</sub> emissions, unlike the recent claim based on data from a lowland temperate river by Reiman and Xu (2019), given that tropical rivers account for 80 % of CO<sub>2</sub> emissions (Raymond et al., 2013; Borges et al., 2015a, b; Lauerwald et al., 2015).

### 3.5 Drivers of CO<sub>2</sub> dynamics – stable isotope composition of DIC

The stable isotope composition of DIC can provide information on the origin of CO<sub>2</sub>, although the signal depends on the combination of the biological processes that remove or add CO<sub>2</sub> to the water column (CR and PP), rock weathering that adds HCO<sub>3</sub><sup>-</sup>, and outgassing, which removes CO<sub>2</sub> that is <sup>13</sup>C-depleted relative to HCO<sub>3</sub><sup>-</sup>. The variable contribution to DIC in time and in space of CO<sub>2</sub> relative to HCO<sub>3</sub><sup>-</sup> complicates the interpretation of  $\delta^{13}\text{C}$ -DIC data. The stable isotope composition of DIC due to rock weathering will depend on the type of rock (silicate or carbonate) and on the origin of the CO<sub>2</sub> involved in rock dissolution (either CO<sub>2</sub> from the atmosphere or from respiration of soil organic matter). The stable isotopic composition of DIC due to the degradation of organic matter will depend in part on whether the organic matter is derived from terrestrial vegetation following the C<sub>3</sub> photosynthetic pathway (woody plants and trees, temperate grasses;  $\delta^{13}\text{C} \sim -27\text{\textperthousand}$ ) compared to the less fractionating C<sub>4</sub> photosynthetic pathway (largely tropical and subtropical grasses;  $\delta^{13}\text{C} \sim -13\text{\textperthousand}$ ) (Hedges et al., 1986; Bird et al., 1994). Spatial and temporal changes of  $\delta^{13}\text{C}$  are related to the change of the relative abundance of HCO<sub>3</sub><sup>-</sup> over CO<sub>2</sub>. The degassing of CO<sub>2</sub> to the atmosphere and the addition of HCO<sub>3</sub><sup>-</sup> from rock weathering lead to  $\delta^{13}\text{C}$ -DIC values becoming dominated by those related to HCO<sub>3</sub><sup>-</sup>. Since CO<sub>2</sub> is isotopically depleted relative to HCO<sub>3</sub><sup>-</sup>, CO<sub>2</sub> degassing leads to a gradual enrichment of the remaining DIC pool (e.g., Doctor et al., 2008; Deirmendjian and Abril, 2018). The combination of these processes can lead to spatial changes of  $\delta^{13}\text{C}$ -DIC that covary with CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> concentrations. For instance, in the Tana River network, an altitudinal gradient of  $\delta^{13}\text{C}$ -DIC was attributed to a downstream accumulation of HCO<sub>3</sub><sup>-</sup> due to rock weathering combined to CO<sub>2</sub> degassing (Bouillon et al., 2009; Tamoooh et al., 2003). Figure 17 shows  $\delta^{13}\text{C}$ -DIC as a function of the TA : DIC ratio in the rivers and streams of the Congo, with a general increase in  $\delta^{13}\text{C}$ -DIC from TA : DIC = 0 (all of the DIC is in the form of CO<sub>2</sub>) towards TA : DIC = 1 (nearly all of the DIC is in the form of HCO<sub>3</sub><sup>-</sup>). This pattern reflects the mixing of two distinct types of rivers and streams: the lowland systems draining the CCC with low rock weathering due to dominance of



**Figure 14.** Ratio of dissolved CH<sub>4</sub> and CO<sub>2</sub> concentration (μmol:μmol) plotted as a function of O<sub>2</sub> saturation level (%O<sub>2</sub> in %) and plotted in box plots draining and not draining the Cuvette Centrale Congolaise for small and large systems (Strahler stream order 5 ≤ and >5, respectively) (3–19 December 2013; 10–30 June 2014). The box represents the first and third quartile, the horizontal line corresponds to the median, the cross corresponds to the average, the error bars correspond to the maximum and minimum, and the symbols show all data points. A Mann–Whitney test was used to test statistical differences: ns=not significant; \*\*\* =  $p < 0.0001$ .

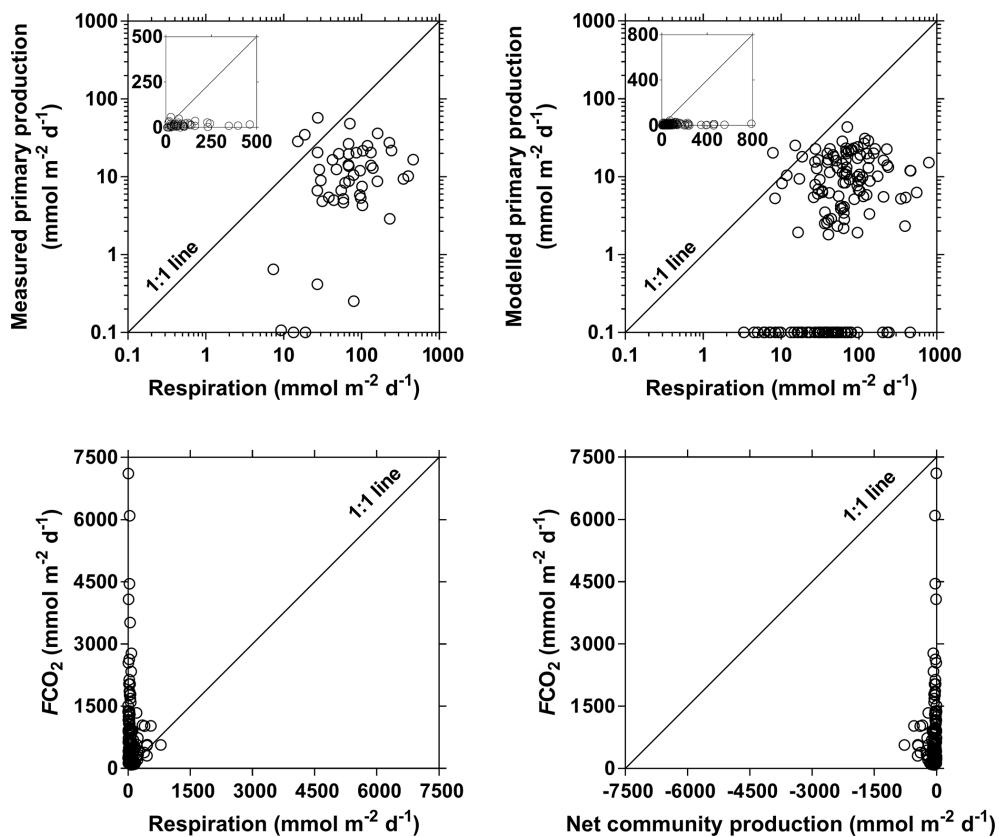
deep organic soils (low HCO<sub>3</sub><sup>-</sup>) and high CO<sub>2</sub> from respiration leading to TA : DIC close to 0 with low δ<sup>13</sup>C-DIC and the systems draining highland regions (Lualaba and Kasaï) with high rock weathering and lower generation of CO<sub>2</sub> from respiration and/or higher CO<sub>2</sub> degassing leading to TA : DIC close to 1 with high δ<sup>13</sup>C-DIC. The plots of TA : Na<sup>+</sup> and Mg<sup>2+</sup> : Na<sup>+</sup> versus Ca<sup>2+</sup> : Na<sup>+</sup> (Fig. 18) showed aggregation of data close to what would be expected for silicate rock weathering based on the average values proposed by Gaillardet et al. (1999). This is in agreement with the dominance of silicate rocks over carbonate rocks in the Congo basin (Nkounkou and Probst, 1987). Note that TA values from the Congo are generally very low compared to other large rivers globally (Meybeck, 1987), due to the large proportion of relatively insoluble rocks on the catchment (70 % of metamorphic rocks) and a small proportion of low soluble rocks such as siliciclastic sedimentary rocks (10 % mainly as sand stone), unconsolidated sediments (17 % as sand and clays) and a very small proportion of highly soluble volcanic rocks (1 %). The high TA values in the Lualaba are due to a larger proportion of volcanic rocks in high-altitude areas, such as the Virunga region, which is rich in volcanic rocks (including basalts) and has been shown to be a hotspot of chemical weathering (Balagizi et al., 2015).

At TA : DIC = 0, the δ<sup>13</sup>C-DIC values are exclusively related to those of CO<sub>2</sub> and might be indicative of the source of mineralized organic matter. The δ<sup>13</sup>C-DIC values ranged between −28.2‰ and −15.9‰ and averaged −24.1‰ (Fig. 17) indicating that CO<sub>2</sub> was produced from the degradation of mixture of organic matter from C<sub>3</sub> and C<sub>4</sub> ori-

gin. Furthermore, δ<sup>13</sup>C-DIC was positively related to pCO<sub>2</sub> (Fig. 17), indicating that in the streams with high pCO<sub>2</sub> values, the CO<sub>2</sub> was generated from the degradation organic matter with a higher contribution from C<sub>4</sub> plants. The δ<sup>13</sup>C-DIC values were unrelated to the contribution of C<sub>4</sub> vegetation on the catchment (terra firme), as modeled by Still and Powell (2010), and the cover by savannah on the catchment given by GLC2009 (Fig. S12). Further, the most enriched δ<sup>13</sup>C-DIC values (>−21‰), corresponded on average to a low contribution of C<sub>4</sub> vegetation on the catchment (7.4 %) and low contribution of savannah cover on the catchment (0.8 %). These patterns are inconsistent with an origin from terra firme of the C<sub>4</sub> material that led to high pCO<sub>2</sub> in streams (Fig. 17) but is rather more consistent with a larger contribution of degradation of C<sub>4</sub> aquatic macrophyte material in high CO<sub>2</sub> streams.

### 3.6 Seasonal variations

The difference between the HW and FW in the main stem along the Kisangani–Kinshasa transects (Figs. 3 and 4) were relatively modest with higher specific conductivity, TA, δ<sup>13</sup>C-DIC, and TSM (at Kisangani); biogeochemical signatures of higher surface run-off during the HW sampling (December 2013); and water flows from deeper soil horizons (or ground-water) during the FW sampling (June 2014). The comparison of tributaries that were sampled during both HW and FW periods along the Kisangani–Kinshasa transects shows that pCO<sub>2</sub> was significantly higher (Wilcoxon match-pairs signed rank test  $p = 0.078$ ) during HW for large systems (freshwater discharge  $\geq 300 \text{ m}^3 \text{ s}^{-1}$ , Table S1) but

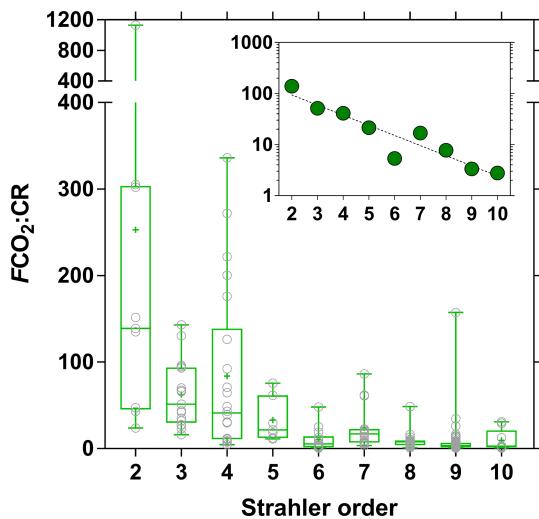


**Figure 15.** Primary production (measured and modeled) ( $\text{mmol m}^{-2} \text{d}^{-1}$ ) as a function of community respiration ( $\text{mmol m}^{-2} \text{d}^{-1}$ ) and air–water  $\text{CO}_2$  fluxes ( $\text{FCO}_2$  in  $\text{mmol m}^{-2} \text{d}^{-1}$ ) as a function of community respiration ( $\text{mmol m}^{-2} \text{d}^{-1}$ ) and net community production ( $\text{mmol m}^{-2} \text{d}^{-1}$ ) in surface waters of rivers and streams of the Congo River network. Insets show the data on a linear scale (instead of a log–log scale).

not significantly different for small systems (freshwater discharge  $< 300 \text{ m}^3 \text{s}^{-1}$ ), irrespective of whether it was from the left or right bank (Fig. 19). However, no significant differences among the HW and FW periods occurred in % $\text{O}_2$ ,  $\text{CH}_4$  and % $\text{N}_2\text{O}$  for either small or large systems, irrespective of whether they were taken from the left or right bank (Fig. 19). This indicates that across the basin, spatial differences among tributaries are more important than seasonal variations within a given tributary. The average difference for large rivers between HW and FW was on average only 1745 ppm for  $p\text{CO}_2$  when the respective range of variation for the whole dataset was from 1087 to 22 899 ppm.

Yearly cycles of  $\text{CH}_4$  and  $\text{N}_2\text{O}$  were established on the Lualaba (6 years), the Oubangui (2 years) and the Kasai (2 years) (Figs. 20 and 21), while  $p\text{CO}_2$  is only available during 2 years in the Lualaba. In the Oubangui and the Kasai,  $\text{CH}_4$  concentration peaked with the onset of rising water and decreased as water level continued to increase. The decrease in  $\text{CH}_4$  as discharge peaked in the Oubangui and the Kasai is most probably related to dilution by surface runoff, as also testified by the decrease in specific conductivity and TA (not shown). The increase at the onset of rising water could be

related to initial flushing of soil atmosphere enriched in  $\text{CH}_4$  as rain penetrates superficial layers of soils. In the Lualaba the  $\text{CH}_4$  seasonal variations seem to follow more closely those of freshwater discharge (for instance in 2014). Unlike the Kasai and the Oubangui, the Lualaba has very extensive permanent marshes and swamps such as the Upemba wetland system (inundated area of  $18\,000 \text{ km}^2$ ) and the Luama swamps (inundated area of  $6000 \text{ km}^2$ ) (Hughes and Hughes, 1992). Yet, it remains to be determined if the seasonality of  $\text{CH}_4$  observed in Lualaba can be attributed to higher inputs during high water from these major wetlands located, respectively, 1000 and 600 km upstream of Kisangani. Even so, there are numerous more modest marshes and swamps that border upstream tributaries of the Lualaba closer to Kisangani (Hughes and Hughes, 1992). The seasonal amplitude of dissolved  $\text{CH}_4$  in the Oubangui was about 10 times higher than in the Kasai and the Oubangui, and the seasonal amplitude of  $\text{CH}_4$  in these three rivers seemed to be related to the relative seasonal amplitude of freshwater discharge, as indicated by the positive relation with the ratio of maximum and minimum of freshwater discharge (Fig. 22).



**Figure 16.** Ratio of air–water  $CO_2$  fluxes ( $FCO_2$  in  $mmol\ m^{-2}\ d^{-1}$ ) and community respiration ( $mmol\ m^{-2}\ d^{-1}$ ) as a function of Strahler stream order for rivers and streams of the Congo River network. The box represents the first and third quartile, the horizontal line corresponds to the median, the cross corresponds to the average, the error bars correspond to the maximum and minimum, and the symbols show all data points. Inset shows on a log scale the median of  $FCO_2 : CR$  and an exponential fit ( $r^2 = 0.88$ ).

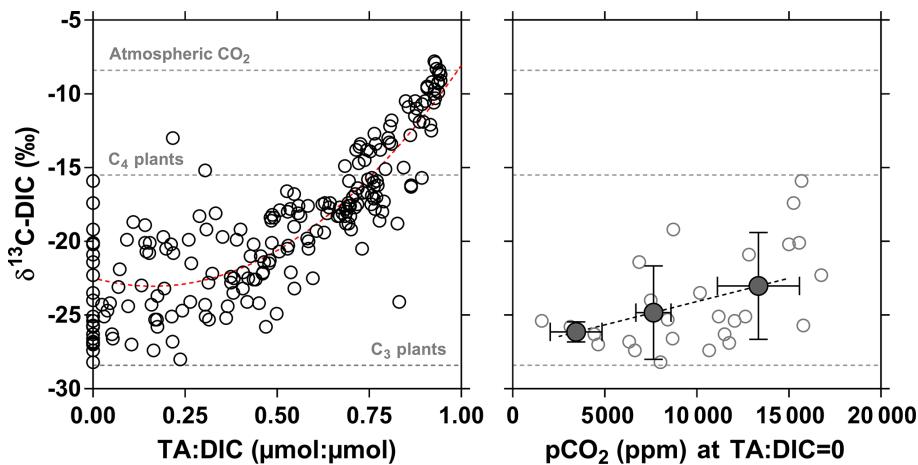
The seasonal cycles of  $\%N_2O$  show patterns that were consistent with those  $CH_4$ , with a loose parallelism of  $\%N_2O$  and discharge in the Lualaba, an inverse relation in the Kasaï, and peak of  $\%N_2O$  during rising waters (although delayed with respect to the  $CH_4$  peak) in the Oubangui. In addition to the seasonal cycle there seemed to be a longer-term decrease in the annual average of  $N_2O$  in the Lualaba (Fig. S13). There were no significant long-term changes of the annual average of other variables such as POC and PN (not shown). So the observed decrease in annual  $N_2O$  is probably unrelated to changes in terrestrial productivity (Zhou et al., 2014) or nitrogen content of terrestrial vegetation (Craine et al., 2018) that act at longer timescales (several decades) and seem to be related to long-term changes in climate (precipitation). Freshwater discharge showed an increasing pattern during the time period (Fig. S13). An increasing discharge could lead to increased gas transfer velocities and a loss of  $N_2O$  to the atmosphere. Water temperature in the Congo River is already close to the optimum of denitrification ( $\sim 27^\circ C$ , Canion et al., 2014), so an enhancement of denitrification with increase in temperature is unlikely. Water temperature did not show a clear pattern, but freshwater discharge increased during the 2013–2018 period, excluding the year 2017. It is likely that the decreasing trend in  $N_2O$  during the 2013–2018 period is a transient feature in response to inter-annual fluctuations in hydrology that led to a period of sustained increase in freshwater discharge.

The  $pCO_2$  time series in the Lualaba at Kisangani is shorter than for  $CH_4$ , but a general positive relationship between  $pCO_2$  and discharge was observed (Fig. S14). A similar positive relationship between  $pCO_2$  and discharge was also observed in the Oubangui (Bouillon et al., 2012) and the Madeira River (Almeida et al., 2017), as well in several rivers in the Amazon (Richey et al., 2002) based on  $pCO_2$  calculated from pH and TA. Such  $pCO_2$  discharge patterns were interpreted as resulting from higher connectivity during high water between the river main stem and the floodplains and wetlands. In large temperate rivers, a negative relationship between  $pCO_2$  and discharge was observed in the Meuse (Borges et al., 2018) and in more than half of the US rivers analyzed by Liu and Raymond (2018). The difference in the relationship of  $pCO_2$  versus discharge between tropical and temperate large rivers might be related to lower interactions between river and floodplains in temperate rivers, particularly in highly human-impacted and channelized rivers such as the Meuse. This is in agreement with the analysis of Aho and Raymond (2019), who reported, in the Salmon River network, positive relationships between  $pCO_2$  and flow in water watersheds with a high presence of wetlands and negative relationships between  $pCO_2$  and flow in watersheds with low presence of wetlands. Also, in temperate rivers, temperature covaries strongly with discharge in temperate rivers, so that the warmer months that promote biological production of  $CO_2$  are also characterized by lower discharge (Borges et al., 2018).

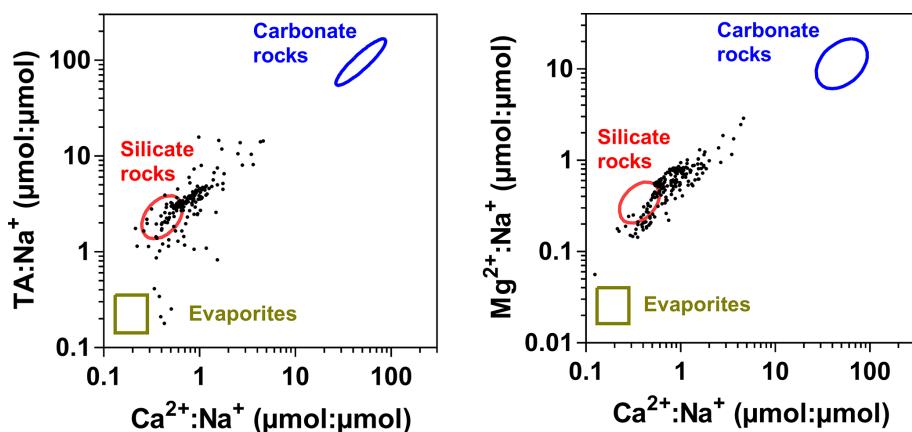
The seasonal amplitude of  $CH_4$  ( $\sim 50\ nmol\ L^{-1}$  in the Kasaï and the Congo, versus  $200$ – $400\ nmol\ L^{-1}$  in the Oubangui) and  $\%N_2O$  (20 %–90 % in the 3 rivers) was overall much lower than the spatial gradients across the basin of 22 and  $71\ 428\ nmol\ L^{-1}$  for  $CH_4$  and 0 and 561 % for  $N_2O$ .

### 3.7 Variability of GHG fluxes in the Congo river network

Since  $pCO_2$  and  $CH_4$  and  $N_2O$  concentrations followed distinct patterns as a function of Strahler stream order (Fig. 11) we used  $k_{600}$  and stream surface area as a function of Strahler stream order (Fig. S15) to compute the air–water GHG fluxes and to integrate them at basin scale. This was done separating data for streams draining and not draining the CCC since  $pCO_2$  and  $CH_4$  and  $N_2O$  patterns were very different (Figs. 11 and 12). The stream surface area decreased regularly with increasing Strahler stream order but showed a large increase for Strahler stream order 9 (Fig. S15). The latter mainly corresponds to the Congo main stem that downstream of Kisangani is characterized by anastomosing river channels with extended sand bars and numerous islands (Runge, 2008; O'Loughlin et al., 2013). In particular along the about 500 km long section between the Mbandaka and Kwa mouths, the main stem river channel undergoes a general expansion and width increases from  $\sim 4$  to  $\sim 10\ km$ . This corresponds to the area of CCC depre-



**Figure 17.** Carbon stable isotope composition of dissolved inorganic carbon (DIC) ( $\delta^{13}\text{C-DIC}$  in ‰) as a function of the total alkalinity (TA) to DIC ratio ( $\mu\text{mol} : \mu\text{mol}$ ) and as a function of the partial pressure of  $\text{CO}_2$  ( $p\text{CO}_2$  in ppm) for a TA:DIC ratio equal to zero in surface waters of rivers and streams of the Congo River network. Open dots indicate individual data points, full dots indicate binned averages ( $\pm$  stand deviation) (bins < 5000, 5000–10 000 and > 10 000 ppm). Horizontal dotted lines indicate the  $\delta^{13}\text{C}$  values of atmospheric  $\text{CO}_2$  and of average soil organic carbon stable isotope composition with a dominance of  $\text{C}_4$  ( $-15.5 \pm 0.8\text{ ‰}$ ) and  $\text{C}_3$  plants ( $-28.4 \pm 0.7\text{ ‰}$ ) (Bird and Pousai, 1997). Dotted red line provides polynomial fit ( $\delta^{13}\text{C-DIC} = -22.5 + 21.37 \times \text{TA : DIC} - 6.97 \times (\text{TA : DIC})^2$ ,  $r^2 = 0.76$ ).

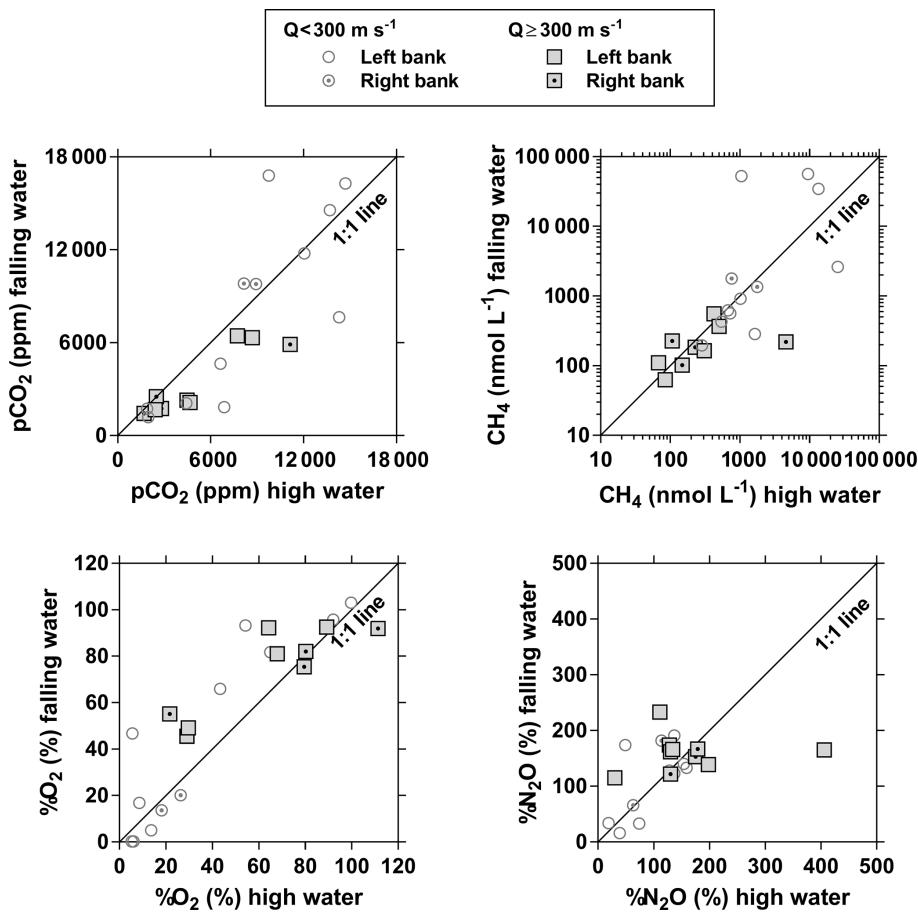


**Figure 18.** Total alkalinity (TA) and  $\text{Mg}^{2+}$  as a function of  $\text{Ca}^{2+}$  of the surface waters of rivers and streams of the Congo River network, in  $\text{Na}^+$  normalized plots ( $\mu\text{mol} : \mu\text{mol}$ ) showing the composition fields for rivers draining different lithologies from a global compilation of the 60 largest rivers in the world (Gaillardet et al., 1999).

sion with a corresponding decrease in the slope from 6 to  $3\text{ cm km}^{-1}$  (O'Loughlin et al., 2013). The calculated  $k_{600}$  decreased regularly with increasing Strahler stream order, as previously reported (Butman and Raymond, 2011; Raymond et al., 2012; Deirmendjian and Abril, 2018; Liu and Raymond, 2018), due to higher turbulence in low-order streams associated with higher stream flow due to steeper slopes. HydroSHEDS stream order classification is missing at least 1 stream order because small streams are not correctly represented (Benstead and Leigh, 2012; Raymond et al., 2013). Hence, to correct this bias, we added 1 to stream orders determined by HydroSHEDS, meaning that the lowest stream order to which GHG were attributed was 2. We then extrapolated  $p\text{CO}_2$  and  $\text{CH}_4$  and  $\text{N}_2\text{O}$  concentrations to stream or-

der 1, separating streams draining and not draining the CCC, using a linear regression with higher orders or by using the same value as for order 2 (Fig. S16).

An error analysis on the GHG flux computation and upscaling was carried out by error propagation of the GHG concentration measurements, the  $k$  value estimates and the estimate of surface areas of river channels to scale the areal fluxes, using a Monte Carlo simulation with 1000 iterations. The uncertainty in the GHG concentrations led to an uncertainty of areal fluxes of  $\pm 1.2\%$ ,  $\pm 2.3\%$  and  $\pm 7.1\%$  for  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$ , respectively. The uncertainty in  $k$  derived from tracer experiments is typically  $\pm 30.0\%$  (Ulseth et al., 2019). This leads to a cumulated uncertainty of areal fluxes of  $\pm 17.6\%$ ,  $\pm 17.9\%$  and  $\pm 19.0\%$  for  $\text{CO}_2$ ,  $\text{CH}_4$  and

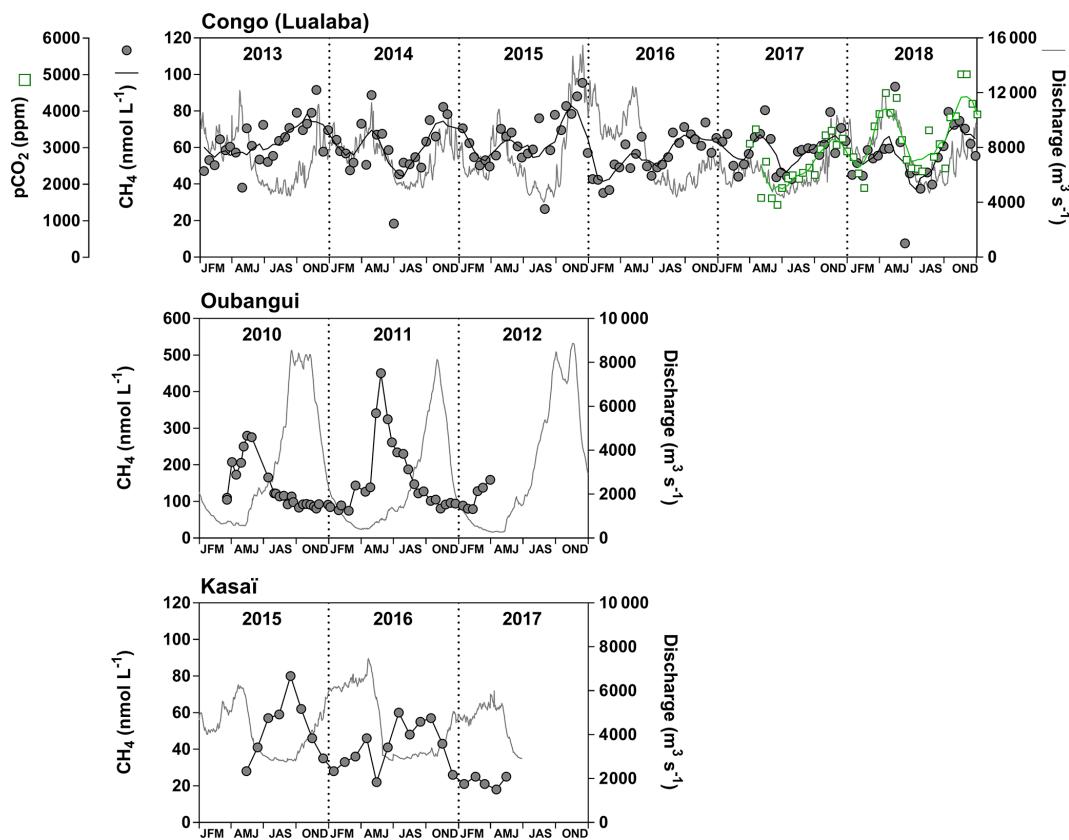


**Figure 19.** Comparison of the partial pressure of  $\text{CO}_2$  ( $\text{pCO}_2$  in ppm), dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ), dissolved  $\text{O}_2$  saturation level ( $\% \text{O}_2$  in %), and dissolved  $\text{N}_2\text{O}$  saturation level ( $\% \text{N}_2\text{O}$  in %) in surface waters of the Congo River tributaries sampled during both high water (3–19 December 2013) and falling water periods (10–30 June 2014). Tributaries were separated into left and right bank, as well as into large and small systems, with a freshwater discharge ( $Q$ )  $<$  and  $\geq 300 \text{ m}^3 \text{s}^{-1}$ , respectively.

$\text{N}_2\text{O}$ , respectively. The uncertainty of the river and stream surface areas based on GIS analysis of Allen and Pavelsky (2018) is estimated to  $\pm 10 \%$  leading to an overall uncertainty of integrated fluxes of  $\pm 18.3 \%$ ,  $\pm 18.3 \%$  and  $\pm 19.6 \%$  for  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$ , respectively.

The calculated  $F\text{CO}_2$  ranged between 86 and  $7110 \text{ mmol m}^{-2} \text{ d}^{-1}$ , averaging  $2469 \pm 435 \text{ mmol m}^{-2} \text{ d}^{-1}$  (weighted by surface area of Strahler stream order), encompassing the range  $F\text{CO}_2$  reported by Mann et al. (2014) ( $312$  to  $1429 \text{ mmol m}^{-2} \text{ d}^{-1}$ ) in 25 sites during a single period (November 2010) from four major river basins in the Republic of the Congo (Alima, Lefini, Sangha, Likouala-Mossaka). The  $\text{pCO}_2$  values ranged between 1087 and  $22\,899 \text{ ppm}$ , also encompassing the values reported by Mann et al. (2014) (2600 to  $15\,802 \text{ ppm}$ ) that were not measured directly but computed from pH and DIC measurements, although pH measurements in black-water rivers can be biased by the presence of humic-dissolved organic matter (Abril et al., 2015) and the addition of  $\text{HgCl}_2$  seems to alter the  $\text{CO}_2$  content of samples (Fig. S1). The calculated  $F\text{CH}_4$

ranged between 65 and  $597\,260 \mu\text{mol m}^{-2} \text{ d}^{-1}$ , averaging  $12\,553 \pm 2247 \mu\text{mol m}^{-2} \text{ d}^{-1}$  (weighted by surface area of Strahler order), encompassing the range  $F\text{CH}_4$  reported by Upstill-Goddard et al. (2017) ( $33$  to  $48\,705 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ) in 41 sites draining the Congo basin in the Republic of the Congo (November 2010 and August 2011) in the same four river basins sampled by Mann et al. (2014). The  $\text{CH}_4$  values ranged between 22 and  $71\,428 \text{ nmol L}^{-1}$ , also encompassing the values reported by Upstill-Goddard et al. (2017) ( $11$  to  $9553 \text{ nmol L}^{-1}$ ). The calculated  $F\text{N}_2\text{O}$  ranged between  $-52$  and  $319 \mu\text{mol m}^{-2} \text{ d}^{-1}$ , averaging  $22 \pm 4 \mu\text{mol m}^{-2} \text{ d}^{-1}$  (weighted by surface area of Strahler order), encompassing the range  $F\text{N}_2\text{O}$  reported by Upstill-Goddard et al. (2017) ( $-19$  to  $67 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ). The  $\% \text{N}_2\text{O}$  values ranged between 0 % and 561 %, also encompassing the values reported by Upstill-Goddard et al. (2017) (6 % to 266 %). The wider ranges of GHGs and respective fluxes we report compared to those of Mann et al. (2014) and Upstill-Goddard et al. (2017) reflect the larger number of river systems sampled over a wider geographical area ( $n = 25$  versus  $n = 278$  for



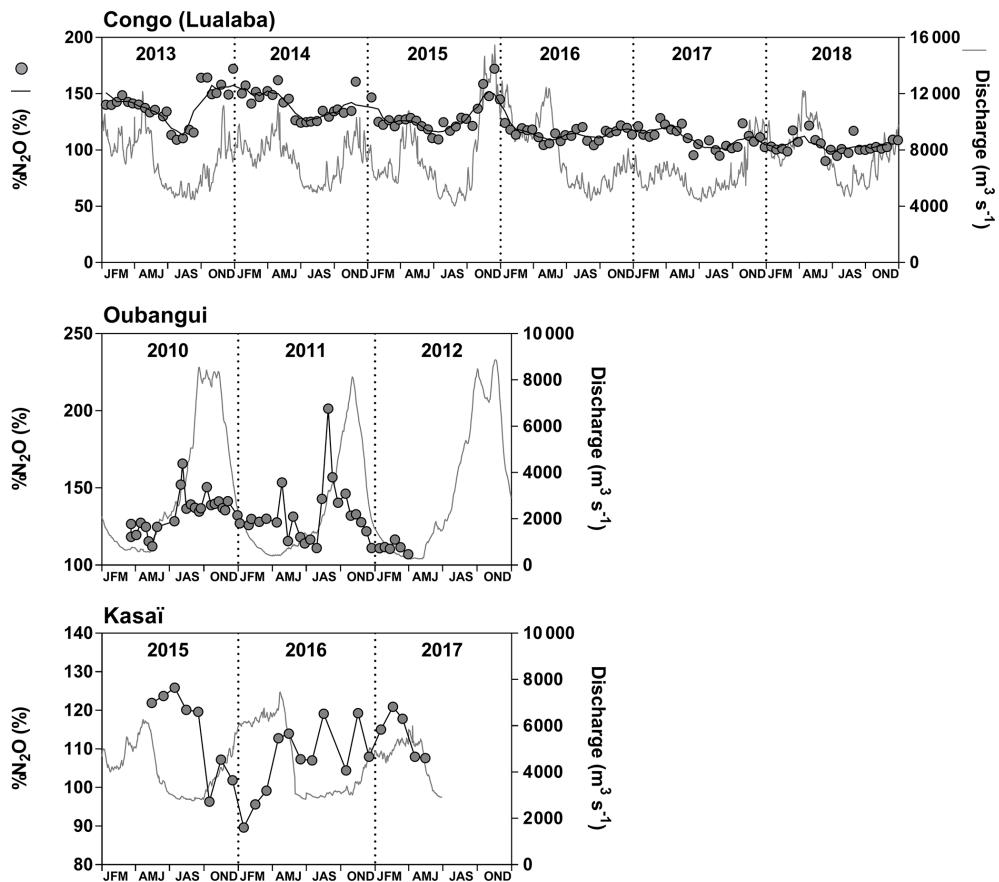
**Figure 20.** Time series of dissolved  $\text{CH}_4$  concentration ( $\text{nmol L}^{-1}$ ) in surface waters and freshwater discharge (grey line) in the Congo (at Kisangani, 2013–2018), the Oubangui (at Bangui, 2010–2012) and the Kasaï (at Dima, 2015–2017) rivers. The black line shows a five sample running average. Time series of the partial pressure of  $\text{CO}_2$  ( $p\text{CO}_2$  in ppm) in surface waters was also obtained in the Congo River (at Kisangani, 2017–2018). Data in the Oubangui were previously reported by Bouillon et al. (2012, 2013).

$p\text{CO}_2$  and  $n = 41$  versus  $n = 367$  for  $\text{CH}_4$  and  $\text{N}_2\text{O}$ ), hence, representing a wider range of river types, morphologies, catchment characteristics and wetland densities.

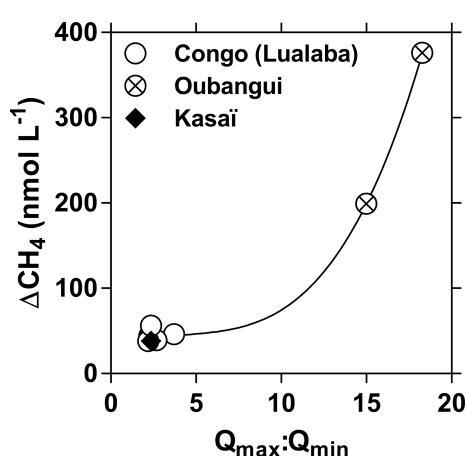
Information on the seasonal variability of concurrent  $F\text{CO}_2$ ,  $F\text{CH}_4$  and  $F\text{N}_2\text{O}$  values was only available in the Lualaba (at Kisangani), where the three GHGs were measured simultaneously (Fig. S17).  $F\text{CO}_2$ ,  $F\text{CH}_4$  and  $F\text{N}_2\text{O}$  were loosely positively correlated with freshwater discharge, as the seasonal variations in  $k_{600}$  were small (ranging between  $23.4$  and  $30.3 \text{ cm h}^{-1}$ ), and although  $p\text{CO}_2$  was correlated to freshwater discharge (Fig. S14) this was not the case for  $\text{CH}_4$  and  $\text{N}_2\text{O}$  concentrations (Figs. 20 and 21). The range of seasonal variations at Kisangani of  $F\text{CO}_2$  ( $234$  and  $948 \text{ mmol m}^{-2} \text{ d}^{-1}$ ),  $F\text{CH}_4$  ( $116$  and  $876 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ) and  $F\text{N}_2\text{O}$  ( $-2$  and  $40 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ) was small compared to the range of spatial variations in  $F\text{CO}_2$  ( $86$  and  $7,110 \text{ mmol m}^{-2} \text{ d}^{-1}$ ),  $F\text{CH}_4$  ( $65$  and  $597,260 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ) and  $F\text{N}_2\text{O}$  ( $-52$  and  $319 \mu\text{mol m}^{-2} \text{ d}^{-1}$ ).

### 3.8 Significance of integrated GHG fluxes at basin and global scales

The  $F\text{CO}_2$  and  $F\text{CH}_4$  decreased with increasing Strahler order, as given by surface area, and also when integrated by surface area of the streams (Table 1). Strahler orders 1–2 accounted for nearly 80 % of the integrated  $F\text{CO}_2$  (79.6 %) and  $F\text{CH}_4$  (77.0 %), while Strahler orders 1–4 accounted for >90 % of the integrated  $F\text{CO}_2$  (90.7 %) and  $F\text{CH}_4$  (91.9 %). Strahler orders 5–10 only accounted for 9.3 % of integrated  $F\text{CO}_2$  and 8.1 % of integrated  $F\text{CH}_4$ . The rivers draining the CCC contributed to 6 % of the basin-wide emissions for  $\text{CO}_2$  and 22 % for  $\text{CH}_4$ , although the contribution in stream surface area was only 11 %. The low contribution of  $F\text{CO}_2$  from the CCC to the basin-wide emissions was due to the lower  $k_{600}$  values, although  $p\text{CO}_2$  values were higher than the rest of the basin (Table 1). In the case of  $F\text{CH}_4$ , the much higher  $\text{CH}_4$  concentrations in the CCC overcome the lower  $k_{600}$  values.  $F\text{N}_2\text{O}$  per surface area in rivers and streams outside the CCC were relatively similar for Strahler orders 10 to 3 and increased for Strahler orders 1 and 2 (Table 1).  $F\text{N}_2\text{O}$  per surface area in rivers and streams draining the CCC steadily



**Figure 21.** Time series of dissolved N<sub>2</sub>O saturation level (%N<sub>2</sub>O in %) in surface waters and freshwater discharge (grey line) in the Congo (at Kisangani, 2013–2018), the Oubangui (at Bangui, 2010–2012) and the Kasai (at Dima, 2015–2017) rivers. The black line shows a five sample running average. Data in the Oubangui were previously reported by Bouillon et al. (2012, 2013).



**Figure 22.** Seasonal amplitude of dissolved CH<sub>4</sub> concentration ( $\Delta\text{CH}_4$  in nmol L<sup>-1</sup>) in surface waters as a function of the ratio of seasonal maximum and minimum of freshwater discharge ( $Q_{\text{max}} : Q_{\text{min}}$ ) in the Congo (at Kisangani, 2013–2018), the Oubangui (at Bangui, 2010–2012) and the Kasai (at Dima, 2015–2017) rivers (Fig. 20).

decreased from Strahler order 8 to 1, with rivers of orders 5, 3, 2 and 1 acting as sinks for N<sub>2</sub>O. Consequently, the relative contribution per Strahler order of integrated FN<sub>2</sub>O was less skewed than for integrated FCO<sub>2</sub> and FCH<sub>4</sub>. Strahler orders 1–4 contributed 69.9 % of integrated FN<sub>2</sub>O compared to > 90 % for FCO<sub>2</sub> and FCH<sub>4</sub>.

The rivers and streams draining the CCC were a very small sink of atmospheric N<sub>2</sub>O ( $-0.01 \text{ GgN}_2\text{O-N yr}^{-1}$ ) while the rivers and streams outside the CCC were a source of N<sub>2</sub>O ( $5.1 \text{ GgN}_2\text{O-N yr}^{-1}$ ). The integrated FN<sub>2</sub>O for the Congo River network was  $5.1 \pm 1.0 \text{ GgN}_2\text{O-N yr}^{-1}$  corresponded to 14 %–17 % of total riverine emissions of N<sub>2</sub>O reported by Hu et al. (2016). Note that the N<sub>2</sub>O riverine emissions computed by Hu et al. (2016) were indirectly computed from data on global nitrogen deposition on catchments and on emission factors rather than derived from direct measurements of dissolved N<sub>2</sub>O concentrations. The estimates given by Hu et al. (2016) are more conservative than older estimates (e.g., Kroese et al., 2010) because they are based on revised emission factors and converge with a similar more recent study by Maavara et al. (2018). Our estimate of the integrated FN<sub>2</sub>O is consistent with the range of N<sub>2</sub>O emissions of 3.8

**Table 1.** Partial pressure of CO<sub>2</sub> ( $p\text{CO}_2$  in ppm); dissolved CH<sub>4</sub> concentration (mmol L<sup>-1</sup>); dissolved N<sub>2</sub>O concentration (mmol L<sup>-1</sup>); air–water fluxes of CO<sub>2</sub> ( $F\text{CO}_2$  in mmol m<sup>-2</sup> d<sup>-1</sup> and in Tg C yr<sup>-1</sup>), CH<sub>4</sub> ( $F\text{CH}_4$  in μmol m<sup>-2</sup> d<sup>-1</sup> and in Tg CH<sub>4</sub> yr<sup>-1</sup>), and N<sub>2</sub>O ( $F\text{N}_2\text{O}$  in μmol m<sup>-2</sup> d<sup>-1</sup> and in Gg N<sub>2</sub>O-N yr<sup>-1</sup>); stream surface area (S.A. in km<sup>2</sup>); and as a function of Strahler stream order (S.O.) in the Congo River network for rivers and streams draining and not draining the Cuvette Centrale Congolaise. Values in italic were extrapolated to stream order 1, using a linear regression with higher orders or by using the same value as for stream order 2 (Fig. S16).

S.O.	$p\text{CO}_2$ ppm	CH <sub>4</sub> nmol L <sup>-1</sup>	N <sub>2</sub> O nmol L <sup>-1</sup>	%N <sub>2</sub> O	$k_{600}$ cm h <sup>-1</sup>	Temp. °C	$F\text{CO}_2$ mmol m <sup>-2</sup> d <sup>-1</sup>	$F\text{CH}_4$ μmol m <sup>-2</sup> d <sup>-1</sup>	$F\text{N}_2\text{O}$ μmol m <sup>-2</sup> d <sup>-1</sup>	S.A. km <sup>2</sup>	$F\text{CO}_2$ Tg C yr <sup>-1</sup>	$F\text{CH}_4$ Tg CH <sub>4</sub> yr <sup>-1</sup>	$F\text{N}_2\text{O}$ Gg N <sub>2</sub> O-N yr <sup>-1</sup>	
Draining the Cuvette Centrale Congolaise														
1	<i>12304</i>	<i>8349</i>	<i>0.5</i>	<i>7.3</i>	<i>26.4</i>	<i>27.0</i>	<i>2892</i>	<i>61 732</i>	<i>-43.1</i>	<i>350</i>	<i>4.4</i>	<i>0.126</i>	<i>-0.154</i>	
2	<i>12304</i>	<i>8349</i>	<i>2.7</i>	<i>44.1</i>	<i>16.1</i>	<i>27.0</i>	<i>1766</i>	<i>37 747</i>	<i>-15.9</i>	<i>335</i>	<i>2.6</i>	<i>0.074</i>	<i>-0.054</i>	
3	<i>11923</i>	<i>7411</i>	<i>4.9</i>	<i>77.5</i>	<i>13.0</i>	<i>26.1</i>	<i>1376</i>	<i>25 765</i>	<i>-5.2</i>	<i>324</i>	<i>2.0</i>	<i>0.049</i>	<i>-0.017</i>	
4	<i>12065</i>	<i>9409</i>	<i>6.5</i>	<i>104.5</i>	<i>12.3</i>	<i>26.7</i>	<i>1323</i>	<i>33 431</i>	<i>0.9</i>	<i>291</i>	<i>1.7</i>	<i>0.057</i>	<i>0.003</i>	
5	<i>13705</i>	<i>9533</i>	<i>2.0</i>	<i>32.2</i>	<i>12.5</i>	<i>27.0</i>	<i>1525</i>	<i>33 255</i>	<i>-14.8</i>	<i>278</i>	<i>1.9</i>	<i>0.054</i>	<i>-0.042</i>	
6	<i>7830</i>	<i>532</i>	<i>13.2</i>	<i>214.7</i>	<i>12.2</i>	<i>27.5</i>	<i>834</i>	<i>1855</i>	<i>24.7</i>	<i>480</i>	<i>1.8</i>	<i>0.005</i>	<i>0.121</i>	
7	<i>6643</i>	<i>503</i>	<i>11.3</i>	<i>188.8</i>	<i>11.7</i>	<i>27.5</i>	<i>670</i>	<i>1582</i>	<i>18.2</i>	<i>163</i>	<i>0.5</i>	<i>0.002</i>	<i>0.030</i>	
8	<i>6977</i>	<i>401</i>	<i>12.9</i>	<i>219.8</i>	<i>13.6</i>	<i>28.7</i>	<i>824</i>	<i>1581</i>	<i>28.6</i>	<i>366</i>	<i>1.3</i>	<i>0.003</i>	<i>0.107</i>	
Not draining the Cuvette Centrale Congolaise														
1	<i>10719</i>	<i>1584</i>	<i>9.4</i>	<i>136.6</i>	<i>161.6</i>	<i>24.1</i>	<i>15 417</i>	<i>66 695</i>	<i>107.2</i>	<i>2235</i>	<i>150.9</i>	<i>0.871</i>	<i>2.449</i>	
2	<i>8921</i>	<i>1275</i>	<i>9.4</i>	<i>135.8</i>	<i>56.5</i>	<i>24.1</i>	<i>4450</i>	<i>19 171</i>	<i>36.4</i>	<i>2143</i>	<i>41.8</i>	<i>0.240</i>	<i>0.798</i>	
3	<i>7225</i>	<i>1185</i>	<i>9.3</i>	<i>133.4</i>	<i>31.1</i>	<i>24.0</i>	<i>1972</i>	<i>9554</i>	<i>18.9</i>	<i>1883</i>	<i>16.3</i>	<i>0.105</i>	<i>0.364</i>	
4	<i>5766</i>	<i>726</i>	<i>8.5</i>	<i>126.3</i>	<i>21.5</i>	<i>24.3</i>	<i>1065</i>	<i>4098</i>	<i>10.1</i>	<i>1752</i>	<i>8.2</i>	<i>0.042</i>	<i>0.181</i>	
5	<i>4110</i>	<i>538</i>	<i>8.6</i>	<i>131.4</i>	<i>17.2</i>	<i>25.5</i>	<i>589</i>	<i>2452</i>	<i>9.6</i>	<i>1688</i>	<i>4.4</i>	<i>0.024</i>	<i>0.165</i>	
6	<i>1929</i>	<i>299</i>	<i>9.3</i>	<i>143.4</i>	<i>16.8</i>	<i>25.6</i>	<i>239</i>	<i>1328</i>	<i>12.9</i>	<i>1772</i>	<i>1.9</i>	<i>0.014</i>	<i>0.233</i>	
7	<i>2667</i>	<i>220</i>	<i>9.3</i>	<i>153.6</i>	<i>16.3</i>	<i>27.5</i>	<i>343</i>	<i>1030</i>	<i>15.4</i>	<i>2168</i>	<i>3.3</i>	<i>0.013</i>	<i>0.340</i>	
8	<i>3010</i>	<i>226</i>	<i>6.7</i>	<i>109.2</i>	<i>12.9</i>	<i>27.7</i>	<i>310</i>	<i>837</i>	<i>2.1</i>	<i>1696</i>	<i>2.3</i>	<i>0.008</i>	<i>0.036</i>	
9	<i>2521</i>	<i>170</i>	<i>8.7</i>	<i>145.7</i>	<i>10.9</i>	<i>27.7</i>	<i>217</i>	<i>533</i>	<i>8.8</i>	<i>4639</i>	<i>4.4</i>	<i>0.014</i>	<i>0.419</i>	
10	<i>3445</i>	<i>33</i>	<i>9.4</i>	<i>153.1</i>	<i>20.4</i>	<i>27.7</i>	<i>574</i>	<i>178</i>	<i>19.2</i>	<i>646</i>	<i>1.6</i>	<i>0.001</i>	<i>0.127</i>	
Total													251	1.7
5.1														

to  $4.3 \text{ GgN}_2\text{O-N yr}^{-1}$  given by Maavara et al. (2018) for the Congo river network, also based on an indirect calculation based on nitrogen deposition and emission factors.

The integrated  $F\text{CH}_4$  for the Congo River network of  $1.7 \pm 0.3 \text{ TgCH}_4 \text{ yr}^{-1}$  is nearly 2 times higher than the estimate given by Bastviken et al. (2011) for all tropical rivers and corresponds to 6 % of the global emission of  $\text{CH}_4$  from rivers given by Stanley et al. (2016), while the surface area of the Congo River network corresponds to a lower proportion (3 %) of the global riverine surface area ( $773\,000 \text{ km}^2$ , Allen and Pavelsky). Note that the meta-analysis of Stanley et al. (2016) includes part of our dataset from the Congo River, as published by Borges et al. (2015a). The integrated  $F\text{CH}_4$  we report for the Congo River network is also more than 3 times higher than the estimate of  $\text{CH}_4$  emissions for Amazonian large rivers reported by Sawakuchi et al. (2014) ( $0.49 \text{ TgCH}_4 \text{ yr}^{-1}$ ). The integrated  $F\text{CH}_4$  we report for the Congo River network corresponds to 7 % of the emission from the Congo wetlands inferred from remotely sensed atmospheric  $\text{CH}_4$  data (Bloom et al., 2010) ( $25.7 \text{ TgCH}_4 \text{ yr}^{-1}$ ). However, the top-down estimate given by Bloom et al. (2011) includes the  $\text{CH}_4$  emission from all ecosystems over the Congo basin and should also include the fluvial emissions. Note that the  $\text{CH}_4$  emissions we report for the Congo River basin only include the diffusive flux component, when the ebullitive  $\text{CH}_4$  emission component represents the majority of  $\text{CH}_4$  emissions from inland waters (Bastviken et al., 2011), which would be consistent with the gap between our emission estimates and those from atmospheric  $\text{CH}_4$  inventories. Finally, note that the  $\text{CH}_4$  emissions were only calculated and integrated for the rivers and streams draining the CCC but not for the actual wetland flooded area of the CCC. The emission of  $\text{CH}_4$  from actual wetland flooded area of the CCC can be estimated to a massive  $51 \text{ TgCH}_4 \text{ yr}^{-1}$  by extrapolating the area averaged  $F\text{CH}_4$  from the streams ( $24\,468 \mu\text{mol m}^{-2} \text{ d}^{-1}$ , Table 1) to the flooded extent ( $360\,000 \text{ km}^2$ , Bwangoy et al., 2010). This corresponds to about 29 % of the  $\text{CH}_4$  emissions from natural wetlands ( $\sim 180 \text{ TgCH}_4 \text{ yr}^{-1}$ , Saunois et al., 2010) and would in this case be higher than the estimate of  $\text{CH}_4$  emissions from the Congo wetlands inferred from remotely sensed atmospheric  $\text{CH}_4$  data (Bloom et al., 2010).

The integrated  $F\text{CO}_2$  for the Congo River network is  $251 \pm 46 \text{ TgC yr}^{-1}$  and is equivalent to the  $\text{CO}_2$  emission value for rivers globally given by Cole et al. (2007) and to 14 % and 39 % of the  $\text{CO}_2$  emission value for rivers globally given by Raymond et al. (2013) ( $18\,000 \text{ TgC yr}^{-1}$ ) and Lauerwald et al. (2015) ( $650 \text{ TgC yr}^{-1}$ ), respectively. The integrated  $\text{CO}_2$  emission from the Congo River network corresponds to 44 % of the  $\text{CO}_2$  emissions from tropical ( $24^\circ \text{ N}$ – $24^\circ \text{ S}$ ) oceans globally ( $565 \text{ TgC yr}^{-1}$ ) and 183 % of  $\text{CO}_2$  emissions from the tropical Atlantic Ocean ( $137 \text{ TgC yr}^{-1}$ ), based on the Takahashi et al. (2002)  $F\text{CO}_2$  climatology. The terrestrial net ecosystem exchange (NEE) of the watershed of the Congo River can be estimated based on the NEE

estimate of  $23 \text{ gC m}^{-2} \text{ d}^{-1}$  for savannahs given by Ciais et al. (2011) and of  $20 \text{ gC m}^{-2} \text{ d}^{-1}$  for forests given by Fisher et al. (2013) and based on the respective land cover from GLC2009 (30 % savannah and 70 % forest). The corresponding terrestrial NEE of  $77 \text{ TgC yr}^{-1}$  is more than 3 times lower than the riverine  $\text{CO}_2$  emission from the Congo River. This is extremely surprising since the hydrological export from terra firme forests of DOC and DIC that are assumed to sustain fluvial emissions are typically 2 %–3 % compared to terrestrial NEE (Kindler et al., 2011; Deirmendjian et al., 2018). Hydrological carbon export is higher compared to NEE in European grasslands (on average 22 %) (Kindler et al., 2011). We ignore if this is transposable to tropical grasslands, such as savannahs, although they only occupy 30 % of the Congo catchment surface. Accordingly, the  $\text{CO}_2$  emission from the Congo River network should have been an order of magnitude lower than the estimates of terrestrial NEE from terra firme biomes rather than more than 3 times higher. However, in wetlands such as peatlands in Europe, the hydrological export of DOC and DIC represent 109 % of NEE and is enough to sustain the riverine  $\text{CO}_2$  emission that represents 17 % of NEE (Billett et al., 2004). Indeed, the carbon export from flooded forests to riverine waters of the Congo basin can be roughly estimated to  $396 \text{ TgC yr}^{-1}$  and is in excess of the integrated  $F\text{CO}_2$  (calculated from the export per surface area of flooded forest of  $1100 \text{ gC m}^{-2} \text{ yr}^{-1}$  reported by Abril et al., 2014, for the central Amazon and surface area of flooded forest of the CCC,  $360\,000 \text{ km}^2$ , Bwangoy et al., 2010). Altogether, this would then strongly suggest that the  $\text{CO}_2$  emission from the lowland Congo River network is to a large extent sustained by another source of carbon than from the terrestrial terra firme biome. The most likely alternative source would be wetlands (flooded forest and aquatic macrophytes), in agreement with the analysis in the central Amazon River by Abril et al. (2013).

#### 4 Conclusions

Net heterotrophy in rivers and lakes sustained by inputs of organic matter from the terrestrial vegetation on the catchments is the prevailing paradigm to explain oversaturation of  $\text{CO}_2$  in inland surface waters and corresponding emissions to the atmosphere, based on process studies, the earliest in the Amazon (Wissmar et al., 1981) and boreal systems (Del Giorgio et al., 1999; Prairie et al., 2002), and then generalized at global scales for lakes (Cole et al., 1994) and rivers (Cole and Caraco, 2001). Yet, the comparison of 169 measurements of aquatic NCP and  $F\text{CO}_2$  estimates in the central Congo River network covering a wide range of size and type of rivers and streams shows that the aquatic NCP cannot account for fluvial  $F\text{CO}_2$ . This implies that lateral inputs of  $\text{CO}_2$  sustain a large part of the  $\text{CO}_2$  emissions from rivers and streams in the Congo River network.

The comparison of the integrated CO<sub>2</sub> emission from the Congo River network with terrestrial terra firme NEE shows that it unlikely that fluvial CO<sub>2</sub> emissions are sustained by lateral hydrological transfer of carbon from terra firme. Indeed, integrated  $F\text{CO}_2$  from the river network was more than 3 times higher than terrestrial NEE, when forests typically only export a very small fraction of NEE as carbon to rivers that can sustain fluvial CO<sub>2</sub> emissions. It is then likely that the fluvial CO<sub>2</sub> emissions from the central Congo River are sustained by organic matter inputs as well as direct CO<sub>2</sub> inputs from extensive riparian wetlands (flooded forest and aquatic floating macrophytes). This is consistent with the stable isotopic signature of DIC, the differences in the spatial distribution of dissolved CO<sub>2</sub>, %O<sub>2</sub> and CH<sub>4</sub> between rivers and streams draining or not draining the CCC, a large wetland region in the core of the basin, and based on the correlation between  $p\text{CO}_2$  levels and the cover of flooded forest in the catchment. Indeed, the calculated export of carbon from the CCC to the riverine network is sufficient to sustain the fluvial CO<sub>2</sub> emission from the Congo.

The fact that fluvial CO<sub>2</sub> emissions in lowland rivers are to a large extent sustained by carbon inputs from wetlands in addition to those from terra firme has consequences for the conceptualization of statistical and mechanistic models of carbon cycling in river networks. While progress has been made in integrating wetland connectivity in mechanistic regional models (Lauerwald et al., 2017), this has not been the case so far for statistical global models that rely on terrestrial (terra firme) productivity (Lauerwald et al., 2015). The comparison of the output of such a statistical model for the Congo River with observational data (Fig. S18) shows that the model fails to represent spatial gradients and the higher  $p\text{CO}_2$  values of streams and rivers draining the CCC in particular. This illustrates how ignoring the river–wetland connectivity can lead to the misrepresentation of  $p\text{CO}_2$  dynamics in river networks, in particular tropical ones that account for the vast majority (80 %) of global riverine CO<sub>2</sub> emissions.

**Data availability.** Full dataset is available at <https://zenodo.org/record/3413449> (Borges and Bouillon, 2019).

**Supplement.** The supplement related to this article is available online at: <https://doi.org/10.5194/bg-16-3801-2019-supplement>.

**Author contributions.** AVB and SB conceived the study. AVB, FD, TL, ET, ATS, TB, JB, CRT and SB collected field samples. AVB, FD, TL, CM, CRT, JPD and SB made laboratory analysis. GHA and TL carried out GIS analysis. AVB drafted the manuscript with substantial inputs from SB. All authors contributed to the manuscript.

**Competing interests.** The authors declare that they have no conflict of interest.

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## References

- Abril, G. and Borges, A. V.: Carbon leaks from flooded land: do we need to re-plumb the inland water active pipe?, *Biogeosciences*, 16, 769–784, <https://doi.org/10.5194/bg-16-769-2019>, 2019.
- Abril, G., Martinez, J.-M., Artigas, L. F., Moreira-Turcq, P., Benedetti, M. F., Vidal, L., Meziane, T., Kim, J.-H., Bernardes, M. C., Savoye, N., Deborde, J., Albéric, P., Souza, M. F. L., Souza, E. L., and Roland, F.: Amazon river carbon dioxide outgassing fuelled by wetlands, *Nature*, 505, 395–398, <https://doi.org/10.1038/nature12797>, 2014.
- Abril, G., Bouillon, S., Darchambeau, F., Teodoru, C. R., Marwick, T. R., Tambooh, F., Omengo, F. O., Geeraert, N., Deirmendjian, L., Polsenaere, P., and Borges A. V.: Technical note: Large overestimation of  $p\text{CO}_2$  calculated from pH and alkalinity in acidic, organic-rich freshwaters, *Biogeosciences*, 12, 67–78, <https://doi.org/10.5194/bg-12-67-2015>, 2015.
- Aho, K. S. and Raymond, P. A.: Differential response of greenhouse gas evasion to storms in forested and wetland streams, *J. Geophys. Res.*, 124, 649–662, <https://doi.org/10.1029/2018JG004750>, 2019.

- Allen, G. H. and Pavelsky, T. M.: Global extent of rivers and streams, *Science*, 28, eaat0636, <https://doi.org/10.1126/science.aat0636>, 2018.
- Almeida, R. M., Pacheco, F. S., Barros, N., Rosi, E., and Roland, F.: Extreme floods increase CO<sub>2</sub> outgassing from a large Amazonian river, *Limnol. Oceanogr.*, 62, 989–999, <https://doi.org/10.1002/lno.10480>, 2017.
- Alsdorf, D., Beighley, E., Laraque, A., Lee, H., Tshimanga, R., O'Loughlin, F., Mahé, G., Dinga, B., Moukandi, G., and Spencer, R. G. M.: Opportunities for hydrologic research in the Congo Basin, *Rev. Geophys.*, 54, 378–409, <https://doi.org/10.1002/2016RG000517>, 2016.
- Amaral, J. H. F., Borges, A. V., Melack, J. M., Sarmento, H., Barbosa, P. M., Kasper, D., Melo, M. L., de Fex Wolf, D., da Silva, J. S., and Forsberg, B. R.: Influence of plankton metabolism and mixing depth on CO<sub>2</sub> dynamics in an Amazon floodplain lake, *Sci. Total Environ.*, 630, 1381–1393, <https://doi.org/10.1016/j.scitotenv.2018.02.331>, 2018.
- APHA: Standard methods for the examination of water and wastewater, American Public Health Association, 1325 pp., 1998.
- Balagizi, C. M., Darchambeau, F., Bouillon, S., Yalire, M. M., Lambert, T., and Borges, A. V.: River geochemistry, chemical weathering and atmospheric CO<sub>2</sub> consumption rates in the Virunga Volcanic Province (East Africa), *Geochem. Geophys. Geosy.*, 16, 2637–2660, <https://doi.org/10.1002/2015GC005999>, 2015.
- Barbosa, P. M., Melack, J. M., Farjalla, V. F., Amaral, J. H. F., Scofield, V., and Forsberg, B. R.: Diffusive methane fluxes from Negro, Solimões and Madeira rivers and fringing lakes in the Amazon basin, *Limnol. Oceanogr.*, 61, S221–S237, <https://doi.org/10.1002/lno.10358>, 2016.
- Bastviken, D., Ejlertsson, J., and Tranvik, L.: Measurement of methane oxidation in lakes: A comparison of methods, *Environ. Sci. Technol.*, 36, 3354–3361, <https://doi.org/10.1021/es010311p>, 2002.
- Bastviken, D., Tranvik, L. J., Downing, J. A., Crill, P. M., and Enrich-Prast, A.: Freshwater methane emissions offset the continental carbon sink, *Science*, 331, p. 50, <https://doi.org/10.1126/science.1196808>, 2011.
- Battin, T. J., Kaplan, L. A., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., Newbold, J. D., and Sabater, F.: Biophysical controls on organic carbon fluxes in fluvial networks, *Nat. Geosci.*, 1, 95–100, <https://doi.org/10.1038/ngeo101>, 2008.
- Baulch, H. M., Schiff, S. L., Maranger, R., and Dillon, P. J.: Nitrogen enrichment and the emission of nitrous oxide from streams, *Global Biogeochem. Cy.*, 25, GB4013, <https://doi.org/10.1029/2011GB004047>, 2011.
- Benstead, J. P. and Leigh, D. S.: An expanded role for river networks, *Nat. Geosci.*, 5, 678–679, <https://doi.org/10.1038/ngeo1593>, 2012.
- Billett, M. F., Palmer, S. M., Hope, D., Deacon, C., Storeton-West, R., Hargreaves, K. J., Flechard, C., and Fowler, D.: Linking land-atmosphere-stream carbon fluxes in a lowland peatland system, *Global Biogeochem. Cy.*, 18, GB1024, <https://doi.org/10.1029/2003GB002058>, 2004.
- Bird, M. I. and Pousai, P.: Variations of δ<sup>13</sup>C in the surface soil organic carbon pool, *Global Biogeochem. Cy.*, 11, 313–322, <https://doi.org/10.1029/97GB01197>, 1997.
- Bird, M. I., Giresse, P., and Chivas, A. R.: Effect of forest and savanna vegetation on the carbon-isotope composition from the Sanaga River, Cameroon, *Limnol. Oceanogr.*, 39, 1845–1854, <https://doi.org/10.4319/lo.1994.39.8.1845>, 1994.
- Bowen, G. J., Wassenaar, L. I., and Hobson, K. A.: Global application of stable hydrogen and oxygen isotopes to wildlife forensics, *Oecologia*, 143, 337–348, <https://doi.org/10.1007/s00442-004-1813-y>, 2005.
- Bloom, A. A., Palmer, P. I., Fraser, A., Reay, D. S., and Frankenberg, C.: Large-scale controls of methanogenesis inferred from methane and gravity spaceborne data, *Science*, 327, 322–325, <https://doi.org/10.1126/science.1175176>, 2010.
- Borges, A. V., Darchambeau, F., Teodoru, C. R., Marwick, T. R., Tamooij, F., Geeraert, N., Omengo, F. O., Guérin, F., Lambert, T., Morana, C., Okuku, E., and Bouillon, S.: Globally significant greenhouse gas emissions from African inland waters, *Nat. Geosci.*, 8, 637–642, <https://doi.org/10.1038/NGEO2486>, 2015a.
- Borges, A. V., Abril, G., Darchambeau, F., Teodoru, C. R., Deborde, J., Vidal, L. O., Lambert, T., and Bouillon, S.: Divergent biophysical controls of aquatic CO<sub>2</sub> and CH<sub>4</sub> in the World's two largest rivers, *Sci. Rep.*, 5, 15614, <https://doi.org/10.1038/srep15614>, 2015b.
- Borges, A. V., Darchambeau, F., Lambert, T., Bouillon, S., Morana, C., Brouyère, S., Hakoun, V., Jurado, A., Tseng, H.-C., Descy, J.-P., and Roland, F. A. E.: Effects of agricultural land use on fluvial carbon dioxide, methane and nitrous oxide concentrations in a large European river, the Meuse (Belgium), *Sci. Total Environ.*, 610/611, 342–355, <https://doi.org/10.1016/j.scitotenv.2017.08.047>, 2018.
- Borges, A. V. and Bouillon, S.: Data-base of CO<sub>2</sub>, CH<sub>4</sub>, N<sub>2</sub>O and ancillary data in the Congo River, available at: <https://zenodo.org/record/3413449#.XYm2eUYzaUk>, last access: 24 September 2019.
- Bouillon, S., Abril, G., Borges, A. V., Dehairs, F., Govers, G., Hughes, H. J., Merckx, R., Meysman, F. J. R., Nyunja, J., Osburn, C., and Middelburg, J. J.: Distribution, origin and cycling of carbon in the Tana River (Kenya): a dry season basin-scale survey from headwaters to the delta, *Biogeosciences*, 6, 2475–2493, <https://doi.org/10.5194/bg-6-2475-2009>, 2009.
- Bouillon, S., Yambélé, A., Spencer, R. G. M., Gillikin, D. P., Hernes, P. J., Six, J., Merckx, R., and Borges, A. V.: Organic matter sources, fluxes and greenhouse gas exchange in the Oubangui River (Congo River basin), *Biogeosciences*, 9, 2045–2062, <https://doi.org/10.5194/bg-9-2045-2012>, 2012.
- Bouillon, S., Yambélé, A., Gillikin, D. P., Teodoru, C., Darchambeau, F., Lambert, T., and Borges, A. V.: Contrasting biogeochemical characteristics of right-bank tributaries and a comparison with the mainstem Oubangui River, Central African Republic (Congo River basin), *Sci. Rep.*, 4, 5402, <https://doi.org/10.1038/srep05402>, 2014.
- Bultot, F.: Atlas Climatique du Bassin Congolais Publications de L'Institut National pour L'Etude Agronomique du Congo (I.N.E.A.C.), Troisième Partie, Température et Humidité de L'Air, Rosee, Température du Sol, 253 pp., 1972.
- Butman, D. and Raymond, P. A.: Significant efflux of carbon dioxide from streams and rivers in the United States, *Nat. Geosci.*, 4, 839–842, <https://doi.org/10.1038/NGEO1294>, 2011.
- Bwangoy, J.-R. B., Hansen, M. C., Roy, D. P., De Grandi, G., and Justice, C. O.: Wetland mapping in the Congo Basin using optical and radar remotely sensed data and derived

- topographical indices, *Remote Sens. Environ.*, 114, 73–86, <https://doi.org/10.1016/j.rse.2009.08.004>, 2010.
- Canion, A., Overholt, W. A., Kostka, J. E., Huettel, M., Lavik, G., and Kuypers, M. M. M.: Temperature response of denitrification and anaerobic ammonium oxidation rates and microbial community structure in Arctic fjord sediments, *Environ. Microbiol.*, 16, 3331–3344, <https://doi.org/10.1111/1462-2920.12593>, 2014.
- Cardoso, S. J., Enrich-Prast, A., Pace, M. L., and Roland, F.: Do models of organic carbon mineralization extrapolate to warmer tropical sediments? *Limnol. Oceanogr.*, 59, 48–54, <https://doi.org/10.4319/lo.2014.59.1.0048>, 2014.
- Ciais, P., Bombelli, A., Williams, M., Piao, S. L., Chave, J., Ryan, C. M., Henry, M., Breden, P., and Valentini, R.: The carbon balance of Africa: synthesis of recent research studies, *Philos. T. R. Soc. A*, 369, 2038–2057, <https://doi.org/10.1098/rsta.2010.0328>, 2013.
- Coplen, T. B. and Wassenaar, L. I.: LIMS for Lasers 2015 for achieving long-term accuracy and precision of  $\delta^2\text{H}$ ,  $\delta^{17}\text{O}$ , and  $\delta^{18}\text{O}$  of waters using laser absorption spectrometry, *Rapid Commun. Mass Spectr.*, 29, 2122–2130, <https://doi.org/10.1002/rcm.7372>, 2015.
- Cole, B. E. and Cloern, J. E.: An empirical model for estimating phytoplankton productivity in estuaries, *Mar. Ecol. Prog. Ser.*, 36, 299–305, <https://doi.org/10.3354/meps036299>, 1987.
- Cole, J. J. and Caraco, N. F.: Carbon in catchments: connecting terrestrial carbon losses with aquatic metabolism, *Mar. Fresh. Res.*, 52, 101–110, <https://doi.org/10.1071/MF00084>, 2001.
- Cole, J. J., Caraco, N. F., Kling, G. W., and Kratz, T. K.: Carbon dioxide supersaturation in the surface waters of lakes, *Science*, 265, 1568–1570, <https://doi.org/10.1126/science.265.5178.1568>, 1994.
- Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., Duarte, C. M., Kortelainen, P., Downing, J. A., Middelburg, J. J., and Melack, J.: Plumbing the global carbon cycle: Integrating inland waters into the terrestrial carbon budget, *Ecosystems*, 10, 171–184, <https://doi.org/10.1007/s10021-006-9013-8>, 2007.
- Coynel, A., Seyler, P., Etcheber, H., Meybeck, M., and Orange, D.: Spatial and seasonal dynamics of total suspended sediment and organic carbon species in the Congo River, *Global Biogeochem. Cy.*, 19, GB4019, <https://doi.org/10.1029/2004GB002335>, 2005.
- Craine, J. M., Elmore, A. J., Wang, L., Aranibar, J., Bauters, M., Boeckx, P., Crowley, B. E., Dawes, M. A., Delzon, S., Fajardo, A., Fang, Y., Fujiyoshi, L., Gray, A., Guerrieri, R., Gundale, M. J., Hawke, D. J., Hietz, P., Jonard, M., Kearsley, E., Kenzo, T., Makarov, M., Marañón-Jiménez, S., McGlynn, T. P., McNeil, B. E., Mosher, S. G., Nelson, D. M., Peri, P. L., Roggy, J. C., Sanders-DeMott, R., Song, M., Szpak, P., Templer, P. H., Van der Colff, D., Werner, C., Xu, X., Yang, Y., Yu, G., and Zmudczyńska-Skarbek, K.: Isotopic evidence for oligotrophication of terrestrial ecosystems, *Nat. Ecol. Evol.*, 2, 1735–1744, <https://doi.org/10.1038/s41559-018-0694-0>, 2018.
- Crawford, J. T., Stanley, E. H., Dornblaser, M., and Striegl, R. G.: CO<sub>2</sub> time series patterns in contrasting headwater streams of North America, *Aquat. Sci.*, 79, 473–486, <https://doi.org/10.1007/s00027-016-0511-2>, 2017.
- Dargie, G. C., Lewis, S. L., Lawson, I. T., Mitchard, E. T. A., Page, S. E., Bocko, Y. E., and Ifo, S. A.: Age, extent and carbon storage of the central Congo Basin peatland complex, *Nature*, 542, 86–90, <https://doi.org/10.1038/nature21048>, 2017.
- Deirmendjian, L. and Abril, G.: Carbon dioxide degassing at the groundwater-stream-atmosphere interface: isotopic equilibration and hydrological mass balance in a sandy watershed, *J. Hydrol.*, 558, 129–143, 2018.
- Deirmendjian, L., Loustau, D., Augusto, L., Lafont, S., Chipeaux, C., Poirier, D., and Abril, G.: Hydro-ecological controls on dissolved carbon dynamics in groundwater and export to streams in a temperate pine forest, *Biogeosciences*, 15, 669–691, <https://doi.org/10.5194/bg-15-669-2018>, 2018.
- Dinsmore, K. J., Wallin, M. B., Johnson, M. S., Billett, M. F., Bishop, K., Pumpanen, J., and Ojala, A.: Contrasting CO<sub>2</sub> concentration discharge dynamics in headwater streams: a multi-catchment comparison, *J. Geophys. Res.*, 118, 445–461, <https://doi.org/10.1002/jgrg.20047>, 2013.
- Del Giorgio, P. A., Cole, J. J., Caraco, N. F., and Peters, R. H.: Linking planktonic biomass and metabolism to net gas fluxes in northern temperate lakes, *Ecology*, 80, 1422–1431, [https://doi.org/10.1890/0012-9658\(1999\)080\[1422:LPBAMT\]2.0.CO;2](https://doi.org/10.1890/0012-9658(1999)080[1422:LPBAMT]2.0.CO;2), 1999.
- Desyc, J.-P., Hardy, M.-A., Sténuite, S., Pirlot, S., Leporcq, B., Kimirei, I., Sekadende, B., Mwaitega, S. R., and Sinyenza, D.: Phytoplankton pigments and community composition in Lake Tanganyika, *Freshwater Biol.*, 50, 668–684, <https://doi.org/10.1111/j.1365-2427.2005.01358.x>, 2005.
- Desyc, J.-P., Darchambeau, F., Lambert, T., Stoyneva, M. P., Bouillon, S., and Borges, A. V.: Phytoplankton dynamics in the Congo River, *Freshwater Biol.*, 62, 87–101, <https://doi.org/10.1111/fwb.12851>, 2017.
- Doctor, D. H., Kendall, C., Sebestyen, S. D., Shanley, J. B., Ohte, N., and Boyer, E. W.: Carbon isotope fractionation of dissolved inorganic carbon (DIC) due to outgassing of carbon dioxide from a headwater stream, *Hydrol. Process.*, 22, 2410–2423, <https://doi.org/10.1002/hyp.6833>, 2008.
- Downing, J. A., Cole, J. J., Duarte, C. M., Middelburg, J. J., Melack, J. M., Prairie, Y. T., Kortelainen, P., Striegl, R. G., McDowell, W. H., and Tranvik, L. J.: Global abundance and size distribution of streams and rivers, *Inland Waters*, 2, 229–236, <https://doi.org/10.5268/IW-2.4.502>, 2012.
- Duvert, C., Butman, D. E., Marx, A., Ribolzi, O., and Huxley, L. B.: CO<sub>2</sub> evasion along streams driven by groundwater inputs and geomorphic controls, *Nat. Geosci.*, 11, 813–818, <https://doi.org/10.1038/s41561-018-0245-y>, 2018.
- Fisher, J. B., Sikka, M., Sitch, S., Ciais, P., Poulter, B., Galbraith, D., Lee, J.-E., Huntingford, C., Viovy, N., Zeng, N., Ahlström, A., Lomas, M. R., Levy, P. E., Frankenberg, C., Saatchi, S., and Malhi, Y.: African tropical rainforest net carbon dioxide fluxes in the twentieth century, *Philos. T. R. Soc. B*, 368, 20120376, <https://doi.org/10.1098/rstb.2012.0376>, 2013.
- Fluet-Chouinard, E., Lehner, B., Rebelo, L.-M., Papa, F., and Hamilton, S. K.: Development of a global inundation map at high spatial resolution from topographic downscaling of coarse-scale remote sensing data, *Remote Sens. Environ.*, 158, 348–361, <https://doi.org/10.1016/j.rse.2014.10.015>, 2015.
- Frankignoulle, M., Borges, A., and Biondo, R.: A new design of equilibrator to monitor carbon dioxide in highly dynamic and turbid environments, *Water Res.*, 35, 1344–1347, [https://doi.org/10.1016/S0043-1354\(00\)00369-9](https://doi.org/10.1016/S0043-1354(00)00369-9), 2001.

- Gaillardet, J., Dupré, B., Louvat, P., and Allègre C. J.: Global silicate weathering and CO<sub>2</sub> consumption rates deduced from the chemistry of large rivers, *Chem. Geol.*, 159, 3–30, [https://doi.org/10.1016/S0009-2541\(99\)00031-5](https://doi.org/10.1016/S0009-2541(99)00031-5), 1999.
- Gillikin, D. P. and Bouillon, S.: Determination of  $\delta^{18}\text{O}$  of water and  $\delta^{13}\text{C}$  of dissolved inorganic carbon using a simple modification of an elemental analyzer – isotope ratio mass spectrometer (EA-IRMS): an evaluation, *Rapid Commun. Mass Spectr.*, 21, 1475–1478, <https://doi.org/10.1002/rcm.2968>, 2007.
- Gran, G.: Determination of the equivalence point in potentiometric titrations Part II, *The Analyst*, 77, 661–671, <https://doi.org/10.1039/AN9527700661>, 1952.
- Hamilton, S. K., Sippel, S. J., and Melack J. M.: Comparison of inundation patterns among major South American floodplains, *J. Geophys. Res.*, 107, LBA 5-1-LBA 5-14, <https://doi.org/10.1029/2000JD000306>, 2002.
- Happell, J., Chanton, J. P., and Showers, W.: The influence of methane oxidation on the stable isotopic composition of methane emitted from Florida Swamp forests, *Geochim. Cosmochim. Ac.*, 58, 4377–4388, [https://doi.org/10.1016/0016-7037\(94\)90341-7](https://doi.org/10.1016/0016-7037(94)90341-7), 1994.
- Hedges, J. I., Clark, W. A., Quay, P. D., Richey, J. E., Devol, A. H., and de M. Santos, U.: Compositions and fluxes of particulate organic material in the Amazon River, *Limnol. Oceanogr.*, 31, 717–738, <https://doi.org/10.4319/lo.1986.31.4.0717>, 1986.
- Hotchkiss, E. R., Hall Jr, R. O., Sponseller, R. A., Butman, D., Klaminder, J., Laudon, H., Rosvall, M., and Karlsson, J.: Sources of and processes controlling CO<sub>2</sub> emissions change with the size of streams and rivers, *Nat. Geosci.*, 8, 696–699, <https://doi.org/10.1038/ngeo2507>, 2015.
- Hu, M., Chen, D., and Dahlgren, R. A.: Modeling nitrous oxide emission from rivers: a global Assessment, *Glob. Change Biol.*, 22, 3566–3582, <https://doi.org/10.1111/gcb.13351>, 2016.
- Hughes, R. H. and Hughes, J. S.: A directory of African wetlands, IUCN, ISBN 2-88032-949-3, 820 pp., 1992.
- Huotari, J., Haapanala, S., Pumpanen, J., Vesala, T., and Ojala, A.: Efficient gas exchange between a boreal river and the atmosphere, *Geophys. Res. Lett.*, 40, 5683–5686, <https://doi.org/10.1002/2013GL057705>, 2013.
- Kindler, R., Siemens, J., Kaiser, K., Walmsley, D. C., Bernhofer, C., Buchmann, N., Cellier, P., Eugster, W., Gleixner, G., Grunwald, T., Heim, A., Ibrom, A., Jones, S. K., Jones, M., Klumpp, K., Kutsch, W., Steenberg Larsen, K., Lehuger, S., Loubet, B., McKenzie, R., Moors, E., Osborne, B., Pilegaard, K., Rebmann, C., Saunders, M., Schmidt, M. W. I., Schrumpf, M., Seyfferth, J., Skiba, U., Soussana, J.-F., Sutton, M. A.; Tefs, C., Vowinkel, B., Zeeman, M. J., and Kaupenjohann, M.: Dissolved carbon leaching from soil is a crucial component of net ecosystem carbon balance, *Glob. Change Biol.*, 17, 1167–1185, <https://doi.org/10.1111/j.1365-2486.2010.02282.x>, 2011.
- Klaus, M., Geibrink, E., Jonsson, A., Bergström, A.-K., Bastviken, D., Laudon, H., Klaminder, J., and Karlsson, J.: Greenhouse gas emissions from boreal inland waters unchanged after forest harvesting, *Biogeosciences*, 15, 5575–5594, <https://doi.org/10.5194/bg-15-5575-2018>, 2018.
- Kokic, J., Sahlée, E., Sobek, S., Vachon, D., and Wallin, M. B.: High spatial variability of gas transfer velocity in streams revealed by turbulence measurements, *Inland Waters*, 8, 461–473, <https://doi.org/10.1080/20442041.2018.1500228>, 2018.
- Koné, Y. J. M., Abril, G., Kouadio, K. N., Delille, B., and Borges, A. V.: Seasonal variability of carbon dioxide in the rivers and lagoons of Ivory Coast (West Africa), *Estuar. Coast.*, 32, 246–260, <https://doi.org/10.1007/s12237-008-9121-0>, 2009.
- Koné, Y. J. M., Abril, G., Delille, B., and Borges, A. V.: Seasonal variability of methane in the rivers and lagoons of Ivory Coast (West Africa), *Biogeochemistry*, 100, 21–37, <https://doi.org/10.1007/s10533-009-9402-0>, 2010.
- Kosten, S., Piñeiro, M., de Goede, E., de Klein, J., Lamers, L. P. M., and Ettwig, K.: Fate of methane in aquatic systems dominated by free-floating plants, *Water Res.*, 104, 200–207, 2016.
- Kroeze, C., Dumont, E., and Seitzinger, S. P.: Future trends in emissions of N<sub>2</sub>O from rivers and estuaries, *J. Integr. Environ. Sc.*, 7, 71–78, <https://doi.org/10.1080/1943815X.2010.496789>, 2010.
- Lambert, T., Bouillon, S., Darchambeau, F., Massicotte, P., and Borges, A. V.: Shift in the chemical composition of dissolved organic matter in the Congo River network, *Biogeosciences*, 13, 5405–5420, <https://doi.org/10.5194/bg-13-5405-2016>, 2016.
- Laraque, A., Mietton, M., Olivry, J. C., and Pandi, A.: Impact of lithological and vegetal covers on flow discharge and water quality of Congolese tributaries from the Congo river, *Rev. Sci. Eau.*, 11, 209–224, 1998.
- Laraque, A., Bricquet, J. P., Pandi, A., and Olivry, J. C.: A review of material transport by the Congo River and its tributaries, *Hydrolog. Process.*, 23, 3216–3224, <https://doi.org/10.1002/hyp.7395>, 2009.
- Lauerwald, R., Laruelle, G. G., Hartmann, J., Ciais, P., and Regnier, P. A. G.: Spatial patterns in CO<sub>2</sub> evasion from the global river network, *Global Biogeochem. Cy.*, 29, 534–554, <https://doi.org/10.1002/2014GB004941>, 2015.
- Lauerwald, R., Regnier, P., Camino-Serrano, M., Guenet, B., Guimberteau, M., Ducharme, A., Polcher, J., and Ciais, P.: OR-CHILEAK (revision 3875): a new model branch to simulate carbon transfers along the terrestrial–aquatic continuum of the Amazon basin, *Geosci. Model Dev.*, 10, 3821–3859, <https://doi.org/10.5194/gmd-10-3821-2017>, 2017.
- Le, T. T. H., Fettig, J., and Meon, G.: Kinetics and simulation of nitrification at various pH values of a polluted river in the tropics, *Ecohydrol. Hydrobiol.*, 19, 54–65, 2019.
- Liptay, K., Chanton, J., Czepiel, P., and Mosher, B.: Use of stable isotopes to determine methane oxidation in landfill cover soils, *J. Geophys. Res.*, 103, 8243–8250, <https://doi.org/10.1029/97JD02630>, 1998.
- Liss, P. S. and Slater, P. G.: Flux of gases across the air sea interface, *Nature*, 247, 181–184, <https://doi.org/10.1038/247181a0>, 1974.
- Liu, S. and Raymond, P. A.: Hydrologic controls on pCO<sub>2</sub> and CO<sub>2</sub> efflux in US streams and rivers, *Limnol. Oceanogr. Lett.*, 3, 428–435, <https://doi.org/10.1002/10l2.10095>, 2018.
- Lynch, J. K., Beatty, C. M., Seidel, M. P., Jungst, L. J., and DeGrandpre, M. D.: Controls of riverine CO<sub>2</sub> over an annual cycle determined using direct, high temporal resolution pCO<sub>2</sub> measurements, *J. Geophys. Res.*, 115, G03016, <https://doi.org/10.1029/2009JG001132>, 2010.
- Maavara, T., Lauerwald, R., Laruelle, G. G., Akbarzadeh, Z., Bouskill, N. J., Van Cappellen, P., and Regnier, P.: Nitrous oxide emissions from inland waters: Are IPCC estimates too high?, *Glob. Change Biol.*, 25, 473–488, <https://doi.org/10.1111/gcb.14504>, 2018.

- Malhi, Y., Adu-Bredu, S., Asare, R. A., Lewis, S. L., and Mayaux, P.: African rainforests: past, present and future, *Philos. T. R. Soc. B*, 368, 20120312, <https://doi.org/10.1098/rstb.2012.0312>, 2013.
- Mann, P. J., Spencer, R. G. M., Dinga, B. J., Poulsen, J. R., Hernes, P. J., Fiske, G., Salter, M. E., Wang, Z. A., Horing, K. A., Six, J., and Holmes, R. M.: The biogeochemistry of carbon across a gradient of streams and rivers within the Congo Basin, *J. Geophys. Res.-Biogeosci.*, 119, 687–702, <https://doi.org/10.1002/2013JG002442>, 2014.
- Maurice, L., Rawlins, B. G., Farr, G., Bell, R., and Goddy, D. C.: The influence of flow and bed slope on gas transfer in steep streams and their implications for evasion of CO<sub>2</sub>, *J. Geophys. Res.-Biogeosci.*, 122, 2862–2875, <https://doi.org/10.1002/2017JG004045>, 2017.
- Marwick, T. R., Tamoooh, F., Ogwoka, B., Teodoru, C., Borges, A. V., Darchambeau, F., and Bouillon, S.: Dynamic seasonal nitrogen cycling in response to anthropogenic N loading in a tropical catchment, Athi–Galana–Sabaki River, Kenya, *Biogeosciences*, 11, 1–18, <https://doi.org/10.5194/bg-11-1-2014>, 2014.
- Marx, A., Dusek, J., Jankovec, J., Sanda, M., Vogel, T., van Geldern, R., Hartmann, J., and Barth, J. A. C.: A review of CO<sub>2</sub> and associated carbon dynamics in headwater streams: A global perspective, *Rev. Geophys.*, 55, 560–585, <https://doi.org/10.1002/2016RG000547>, 2017.
- McDowell, M. J. and Johnson, M. S.: Gas transfer velocities evaluated using carbon dioxide as a tracer show high streamflow to be a major driver of total CO<sub>2</sub> evasion flux for a headwater stream, *J. Geophys. Res.-Biogeosci.*, 123, 2183–2197, <https://doi.org/10.1029/2018JG004388>, 2018.
- Melack, J. M., Hess, L. L., Gastil, M., Forsberg, B. R., Hamilton, S. K., Lima, I. B. T., and Novo, E. M. L. M.: Regionalization of methane emissions in the Amazon Basin with microwave remote sensing, *Glob. Change Biol.*, 10, 530–544, <https://doi.org/10.1111/j.1365-2486.2004.00763.x>, 2004.
- Meybeck, M.: Global chemical weathering of surficial rocks estimated from river dissolved loads, *Am. J. Sci.*, 287, 401–428, <https://doi.org/10.2475/ajs.287.5.401>, 1987.
- Millero, F. J.: The thermodynamics of the carbonate system in seawater, *Geochem. Cosmochim. Ac.*, 43, 1651–1661, [https://doi.org/10.1016/0016-7037\(79\)90184-4](https://doi.org/10.1016/0016-7037(79)90184-4), 1979.
- Morana, C., Borges, A. V., Roland, F. A. E., Darchambeau, F., Descy, J.-P., and Bouillon, S.: Methanotrophy within the water column of a large meromictic, tropical lake (Lake Kivu, East Africa), *Biogeosciences*, 12, 2077–2088, <https://doi.org/10.5194/bg-12-2077-2015>, 2015.
- Nkounkou, R. R. and Probst, J. L.: Hydrology and geochemistry of the Congo river system, *Mitt. Geol-Palaont. Inst. Univ. Hamburg, SCOPE/UNEP*, 64, 483–508, 1987.
- O'Loughlin, F., Trigg, M. A., Schumann, G. J.-P., and Bates, P. D.: Hydraulic characterization of the middle reach of the Congo River, *Water Resour. Res.*, 49, 5059–5070, <https://doi.org/10.1002/wrcr.20398>, 2013.
- Peter, H., Singer, G. A., Preiler, C., Chifflard, P., Steniczka, G., and Battin, T. J.: Scales and drivers of temporal pCO<sub>2</sub> dynamics in an Alpine stream, *J. Geophys. Res.-Biogeosci.*, 119, 1078–1091, <https://doi.org/10.1002/2013JG002552>, 2014.
- Powell, R. L., Yoo, E.-H., and Still, C. J.: Vegetation and soil carbon-13 isoscapes for South America: integrating remote sensing and ecosystem isotope measurements, *Ecosphere*, 3, 1–25, <https://doi.org/10.1890/ES12-00162.1>, 2012.
- Prairie, Y. T., Bird, D. F., and Cole, J. J.: The summer metabolic balance in the epilimnion of southeastern Quebec lakes, *Limnol. Oceanogr.*, 47, 316–321, <https://doi.org/10.4319/lo.2002.47.1.0316>, 2002.
- Raymond, P. A., Zappa, C. J., Butman, D., Bott, T. L., Potter, C., Mulholland, P., Laursen, A. E., McDowell, W. H., and Newbold, D.: Scaling the gas transfer velocity and hydraulic geometry in streams and small rivers, *Limnol. Oceanogr. Fluids Environ.*, 2, 41–53, <https://doi.org/10.1215/21573689-1597669>, 2012.
- Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., Butman, D., Striegl, R., Mayorga, E., Humborg, C., Kortelainen, P., Dürr, H., Meybeck, M., Ciais, P., and Guth, P.: Global carbon dioxide emissions from inland waters, *Nature*, 503, 355–359, <https://doi.org/10.1038/nature12760>, 2013.
- Reiman, J. H. and Xu, J. Y.: Diel variability of pCO<sub>2</sub> and CO<sub>2</sub> outgassing from the Lower Mississippi River: Implications for riverine CO<sub>2</sub> outgassing estimation, *Water*, 11, 1–15, <https://doi.org/10.3390/w11010043>, 2019.
- Richardson, D. C., Newbold, J. D., Aufdenkampe, A. K., Taylor, P. G., and Kaplan, L. A.: Measuring heterotrophic respiration rates of suspended particulate organic carbon from stream ecosystems, *Limnol. Oceanogr.-Method.*, 11, 247–261, <https://doi.org/10.4319/lom.2013.11.247>, 2013.
- Richey, J. E., Devol, A. H., Wofsy, S. C., Victoria, R., and Riberio, M. N. G.: Biogenic gases and the oxidation and reduction of carbon in Amazon River and floodplain waters, *Limnol. Oceanogr.*, 33, 551–561, <https://doi.org/10.4319/lo.1988.33.4.0551>, 1988.
- Richey, J. E., Melack, J. M., Aufdenkampe, A. K., Ballester, V. M., and Hess, L.: Outgassing from Amazonian rivers and wetlands as a large tropical source of atmospheric CO<sub>2</sub>, *Nature*, 416, 617–620, <https://doi.org/10.1038/416617a>, 2002.
- Runge, J.: Large Rivers: Geomorphology and Management, edited by: Gupta, A., John Wiley & Sons., 293–309, 2008.
- Santos, I. R., Maher, D. T., and Eyre, B. D.: Coupling automated radon and carbon dioxide measurements in coastal waters, *Environ. Sci. Technol.*, 46, 7685–7691, <https://doi.org/10.1021/es301961b>, 2012.
- Saunois, M., Bousquet, P., Poulter, B., Peregon, A., Ciais, P., Canadell, J. G., Dlugokencky, E. J., Etiope, G., Bastviken, D., Houweling, S., Janssens-Maenhout, G., Tubiello, F. N., Castaldi, S., Jackson, R. B., Alexe, M., Arora, V. K., Beerling, D. J., Bergamaschi, P., Blake, D. R., Brailsford, G., Brovkin, V., Bruhwiler, L., Crevoisier, C., Crill, P., Kovey, K., Curry, C., Frankenberger, C., Gedney, N., Höglund-Isaksson, L., Ishizawa, M., Ito, A., Joos, F., Kim, H.-S., Kleinen, T., Krummel, P., Lamarque, J.-F., Langenfelds, R., Locatelli, R., Machida, T., Maksyutov, S., McDonald, K. C., Marshall, J., Melton, J. R., Morino, I., Naik, V., O'Doherty, S., Parmentier, F.-J. W., Patra, P. K., Peng, C., Peng, S., Peters, G., Pison, I., Prigent, C., Prinn, R., Ramonet, M., Riley, W. J., Saito, M., Sanyini, M., Schroeder, R., Simpson, I. J., Spahni, R., Steele, P., Takizawa, A., Thornton, B. F., Tian, H., Tohjima, Y., Viovy, N., Voulgarakis, A., van Weele, M., van der Werf, G., Weiss, R., Wiedinmyer, C., Wilton, D. J., Wiltshire, A., Worthy, D., Wunch, D. B., Xu, X., Yoshida, Y., Zhang, B., Zhang, Z., and Zhu, Q.: The global methane budget, *Earth Syst. Sci. Data*, 8, 697–751, <https://doi.org/10.5194/essd-8-697-2016>, 2016.

- Sawakuchi, H. O., Bastviken, D., Sawakuchi, A. O., Krusche, A. V., Ballester, M. V. R., and Richey, J. E.: Methane emissions from Amazonian Rivers and their contribution to the global methane budget, *Glob. Change Biol.*, 20, 2829–2840, <https://doi.org/10.1111/gcb.12646>, 2014.
- Sawakuchi, H. O., Bastviken, D., Sawakuchi, A. O., Ward, N. D., Borges, C. D., Tsai, S. M., Richey, J. E., Ballester, M. V. R. and Krusche, A. V.: Oxidative mitigation of aquatic methane emissions in large Amazonian rivers, *Glob. Change Biol.*, 22, 1075–1085, <https://doi.org/10.1111/gcb.13169>, 2016.
- Scofield, V., Melack, J. M., Barbosa, P. M., Amaral, J. H. F., Forsberg, B. R., and Farjalla, V. F.: Carbon dioxide outgassing from Amazonian aquatic ecosystems in the Negro River basin, *Biogeochemistry*, 129, 77–91, <https://doi.org/10.1007/s10533-016-0220-x>, 2016.
- Seitzinger, S. P. and Kroeze, C.: Global distribution of nitrous oxide production and N inputs in freshwater and coastal marine ecosystems, *Global Biogeochem. Cy.*, 12, 93–113, <https://doi.org/10.1029/97GB03657>, 1998.
- Simpson, H. and Herczeg, A.: Stable isotopes as an indicator of evaporation in the River Murray, Australia, *Water Resour. Res.*, 27, 1925–1935, <https://doi.org/10.1029/91WR00941>, 1991.
- Spencer, R. G. M., Hernes, P. J., Aufdenkampe, A. K., Baker, A., Gulliver, P., Stubbins, A., Aiken, G. R., Dyda, R. Y., Butler, K. D., Mwamba, V. L., Mangangu, A. M., Wabakanganzi, J. N., and Six, J.: An initial investigation into the organic matter biogeochemistry of the Congo River, *Geochim. Cosmochim. Ac.*, 84, 614–627, <https://doi.org/10.1016/j.gca.2012.01.013>, 2012.
- SCA (Standing committee of Analysts): Ammonia in waters. Methods for the examination of waters and associated materials, 16 pp., 1981.
- Stanley, E. H., Casson, N. J., Christel, S. T., Crawford, J. T., Loken, L. C., and Oliver, S. K.: The ecology of methane in streams and rivers: patterns, controls, and global significance, *Ecol. Monogr.*, 86, 146–171, <https://doi.org/10.1890/15-1027>, 2016.
- Still, C. J. and Powell, R. L.: Continental-scale distributions of vegetation stable carbon isotope ratios, edited by: West, J. B., Bowen, G. J., Dawson, T. E., Tu, K. P., Isoscapes, the Netherlands, Springer Netherlands, 179–193, 2010.
- Takahashi, T., Sutherland, S. C., Wanninkhof, R., Sweeney, C., Feely, R. A., Chipman, D. W., Hales, B., Friederich, G., Chavez, F., Sabine, C., Watson, A., Bakker, D. C. E., Schuster, U., Metzl, N., Yoshikawa-Inoue, H.I., Ishii, M., Midorikawa, T., Nojiri, Y., Körtzinger, A., Steinhoff, T., Hoppema, M., Olafsson, J., Arnarson, T. S., Tilbrook, B., Johannessen, T., Olsen, A., Bellerby, R., Wong, C. S., Delille, B., Bates, N. R., and de Baar, H. J. W.: Climatological mean and decadal change in surface ocean  $p\text{CO}_2$ , and net sea-air  $\text{CO}_2$  flux over the global oceans, *Deep-Sea Res. Pt. II*, 56, 554–577, <https://doi.org/10.1016/j.dsr2.2008.12.009>, 2009.
- Tamooh, F., Borges, A. V., Meysman, F. J. R., Van Den Meerseche, K., Dehairs, F., Merckx, R., and Bouillon, S.: Dynamics of dissolved inorganic carbon and aquatic metabolism in the Tana River basin, Kenya, *Biogeosciences*, 10, 6911–6928, <https://doi.org/10.5194/bg-10-6911-2013>, 2013.
- Teodoro, C. R., Nyoni, F. C., Borges, A. V., Darachambeau, F., Nyambe, I., and Bouillon, S.: Spatial variability and temporal dynamics of greenhouse gas ( $\text{CO}_2$ ,  $\text{CH}_4$ ,  $\text{N}_2\text{O}$ ) concentrations and fluxes along the Zambezi River mainstem and major tributaries, *Biogeosciences*, 12, 2431–2453, <https://doi.org/10.5194/bg-12-2431-2015>, 2015.
- Tyler, S. C., Bilek, R. S., Sass, R. L., and Fisher, F. M.: Methane oxidation and pathways of production in a Texas paddy field deduced from measurements of flux,  $\delta^{13}\text{C}$ , and  $\delta\text{D}$  of  $\text{CH}_4$ , *Global Biogeochem. Cy.*, 11, 323–348, <https://doi.org/10.1029/97GB01624>, 1997.
- Ulseth, A. J., Hall Jr, R. O., Canadell, M. B., Madinger, H. L., Niayifar, A., and Battin, T. J.: Distinct air–water gas exchange regimes in low- and high-energy streams, *Nat. Geosci.*, 12, 259–263, <https://doi.org/10.1038/s41561-019-0324-8>, 2019.
- Upstill-Goddard, R. C., Salter, M. E., Mann, P. J., Barnes, J., Poulsen, J., Dinga, B., Fiske, G. J., and Holmes, R. M.: The riverine source of  $\text{CH}_4$  and  $\text{N}_2\text{O}$  from the Republic of Congo, western Congo Basin, *Biogeosciences*, 14, 2267–2281, <https://doi.org/10.5194/bg-14-2267-2017>, 2017.
- Ward, N. D., Krusche, A. V., Sawakuchi, H. O., Brito, D. C., Cunha, A. C., Sousa Moura, J. M., da Silva, R., Yager, P. L., Keil, R. G., and Richey, J. E.: The compositional evolution of dissolved and particulate organic matter along the lower Amazon River–Óbidos to the ocean, *Mar. Chem.*, 177, 244–256, <https://doi.org/10.1016/j.marchem.2015.06.013>, 2015.
- Ward N. D., Sawakuchi, H. O., Neu, V., Less, D. F. S., Valerio, A. M., Cunha, A. C., Kampel, M., Bianchi, T. S., Krusche, A. V., Richey, J. E., and Keil, R. G.: Velocity-amplified microbial respiration rates in the lower Amazon River, *Limnol. Oceanogr. Lett.*, 3, 265–274, <https://doi.org/10.1002/lol2.10062>, 2018.
- Wassenaar, L. I., Coplen, T. B., and Aggarwal, P. K.: Approaches for achieving long-term accuracy and precision of  $\delta^{18}\text{O}$  and  $\delta^2\text{H}$  for waters analyzed using laser absorption spectrometers, *Environ. Sci. Technol.*, 48, 1123–1131, <https://doi.org/10.1021/es403354n>, 2014.
- Weiss, R. F.: Determinations of carbon dioxide and methane by dual catalyst flame ionization chromatography and nitrous oxide by electron capture chromatography, *J. Chromatogr. Sci.*, 19, 611–616, <https://doi.org/10.1093/chromsci/19.12.611>, 1981.
- Weiss, R. F. and Price, B. A.: Nitrous oxide solubility in water and seawater, *Mar. Chem.*, 8, 347–359, [https://doi.org/10.1016/0304-4203\(80\)90024-9](https://doi.org/10.1016/0304-4203(80)90024-9), 1980.
- Wissmar, R. C., Richey, J. E., Stallard, R. F., and Edmond, J. M.: Plankton metabolism and carbon processes in the Amazon river, its tributaries, and floodplain waters, Peru–Brazil, May–June 1977, *Ecology*, 62, 1622–1633, <https://doi.org/10.2307/1941517>, 1981.
- Yoshida, N., Iguchi, H., Yurimoto, H., Murakami, A., and Sakai, Y.: Aquatic plant surface as a niche for methanotrophs, *Front. Microbiol.*, 30, 1–9, <https://doi.org/10.3389/fmicb.2014.00030>, 2014.
- Zhou, L., Tian, Y., Myneni, R. B., Ciais, P., Saatchi, S. L., Yi, Y., Shilong, P., Chen, H., Vermote, E. F., Song, C., and Hwang, T.: Widespread decline of Congo rainforest greenness in the past decade, *Nature*, 509, 86–90, <https://doi.org/10.1038/nature13265>, 2014.